

## RESEARCH ARTICLE

# Dynamics of Oxidants, Antioxidants and Hormones During Different Phases of Pregnancy in Hairy Goats

Nebi ÇETİN<sup>1,a (\*)</sup> Funda EŞKİ<sup>2,b</sup> Leyla MİS<sup>3,c</sup> Zahid NASEER<sup>4,d</sup> Memiş BOLACALI<sup>5,e</sup><sup>1</sup> Department of Obstetrics and Gynecology, Faculty of Veterinary Medicine, Van Yüzüncü Yil University, TR-65080 Van - TURKEY<sup>2</sup> Department of Obstetrics and Gynecology, Faculty of Ceyhan Veterinary Medicine, Cukurova University, TR-01330 Adana - TURKEY<sup>3</sup> Department of Physiology, Faculty of Veterinary Medicine, Van Yuzuncu Yil University, TR-65080 Van - TURKEY<sup>4</sup> Department of Clinical Studies, Faculty of Veterinary and Animal Sciences, Pir Mehr Ali Shah Arid Agriculture University, PK-46000 Rawalpindi - PAKISTAN<sup>5</sup> Department of Animal Science, Faculty of Veterinary Medicine, Siirt University, TR-56100 Siirt - TURKEYORCID: <sup>a</sup> 0000-0001-6073-4215; <sup>b</sup> 0000-0002-9242-9271; <sup>c</sup> 0000-0002-5110-2862; <sup>d</sup> 0000-0001-6335-9633; <sup>e</sup> 0000-0002-4196-2359

Article ID: KVFD-2020-24861 Received: 22.08.2020 Accepted: 06.01.2021 Published Online: 06.01.2021

## Abstract

The aim of the present study was to observe the variation in oxidant (MDA), antioxidants (SOD, GSH, GSH-Px) and hormones (P4 and E2) levels in pregnant hairy goats during breeding season. In this study, twenty hairy goats were synchronized by using sponges containing progesterone (fluorogestone acetate). The animals showing oestrus were inseminated twice, first at 18th -24th h and second at 36th - 48th h of oestrus. On 35th and 42nd days after insemination, pregnancy diagnosis examination was performed with transrectal ultrasonography. The blood samples were collected from pregnant goats at 0, 11, 24, 57, 100, 134, 141 days of gestation and immediately after the parturition. Serum samples collected at estrus and different stages of gestation were analyzed for MDA, SOD, GSH-Px, GSH, P4 and E2 concentrations using standard protocols. The results showed that MDA level did not change in pregnant goats. The SOD, GSH and GSH-Px levels also shown a similar pattern throughout pregnancy period. The peak (P<0.05) level of progesterone was recorded between 11th to 134th days of gestation. At time of estrus (day 0) and late gestation (day 134 and 141), the concentrations estradiol reached at maximal level (P<0.05) in hairy goats. In conclusion, the oxidants and antioxidants do not change with respect to dynamics of progesterone or estradiol level during gestation period in black hairy goats.

**Keywords:** Antioxidants, Oxidants, Pregnancy, Hormones, Hairy Goat

## Kıl Keçilerinde Gebeliğin Farklı Aşamalarında Oksidanların, Antioksidanların ve Hormonların Dinamikleri

### Öz

Bu çalışmanın amacı, üreme sezonundaki gebe kıl keçilerinde oksidan (MDA), antioksidan (SOD, GSH, GSH-Px) ve hormon (P4 ve E2) seviyelerindeki farklılıkları gözlemlemektir. Bu çalışmada 20 adet kıl keçisi progesteron içeren (fluorogeston asetat) vaginal süngerler kullanılarak senkronize edildi. Östrüs gösteren hayvanlar ilk olarak östrüsün 18-24. saatlerinde daha sonra 36-48. saatlerde olmak üzere iki defa tohumlandı. Tohumlama sonrası 35-42. günlerde transrektal ultrasonografi ile gebelik muayenesi yapıldı. Gebe keçilerden gebeliğin 0, 11, 24, 57, 100, 134, 141. günlerinde ve doğumdan hemen sonra kan örnekleri alındı. Gebeliğin farklı dönemlerinde toplanan serum örnekleri standart protokoller kullanılarak MDA, SOD, GSH-Px, GSH, P4 ve E2 konsantrasyonları için analiz edildi. Sonuçlar, MDA seviyesinin gebe keçilerde değişmediğini gösterdi. SOD, GSH ve GSH-Px seviyeleri de gebelik süresince benzer kaldı. Gebeliğin 11 ile 134. günleri arasında progesteron en yüksek seviyededeydi (P<0.05). Gebeliğin 0, 134 ve 141. günlerinde estradiol konsantrasyonu en yüksek seviyeye ulaştı. Sonuç olarak oksidanlar ve antioksidanlar, progesteron ve estradiol seviyelerine kıyasla gebelik süresince herhangi bir değişiklik göstermedi.

**Anahtar sözcükler:** Antioksidanlar, Oksidanlar, Gebelik, Hormonlar, Kıl Keçisi

### How to cite this article?

Çetin N, Eşki F, Mis L, Naseer Z, Bolacali: Dynamics of oxidants, antioxidants and hormones during different phases of pregnancy in hairy goats. *Kafkas Univ Vet Fak Derg*, 27 (1): 117-121, 2021. DOI: 10.9775/kvfd.2020.24861

### (\*) Corresponding Author

Tel: +90 432 225 1128/24503 Fax: +90 432 225 1127

E-mail: [nebicetin@hotmail.com](mailto:nebicetin@hotmail.com) (N. Çetin)



This article is licensed under a Creative Commons Attribution-NonCommercial 4.0 International License (CC BY-NC 4.0)

## INTRODUCTION

During the pregnancy phase, body of the dam and conceptus demands the surplus supply of oxygen. The high demand of oxygen promotes the reactive oxygen species (ROS) production level during implantation to parturition [1]. The normal ROS level is required for the process of steroidogenesis, embryonic development, maternal recognition and implantation during pregnancy. Any imbalance in ROS level leads to cellular disintegration in the dam and conceptus that could result in embryonic mortality, placental degeneration, and pregnancy failure [2]. The pregnancy associated oxidative stress is balanced by regulation of exogenous and endogenous antioxidants level [3] and pre-conception increase in antioxidants favors the establishment of pregnancy [4,5]. Previously, the mechanism of ROS generation and antioxidant response during gestation have been documented in bovine [6] and ovine [7] but little information is available in pregnant goats [8] with respect to antioxidant response and oxidative stress to hormonal changes. Enzymatic scavenging mechanism i.e. glutathione peroxidase, catalase (CAT) and superoxide dismutase (SOD) against oxidative damage prevent dam and fetus from oxidative stress during the pregnancy. Alteration in enzymatic antioxidants occurs due to multiple fetuses, change of environment, low body condition scores and provision of low quality forage during pregnancy [9]. A change in oxidants and antioxidants is also associated with the status of the animal such as pregnancy, parturition and lactation [10].

Cotyledonary placenta is present in the goats and performs similar functions for optimal fetal growth and pregnancy maintenance alike to other species [11,12]. In contrast, the corpus luteum is considered the sole organ for pregnancy maintenance in caprine compared to bovine or ovine where the shift of corpus luteum (CL) to placenta for progesterone production occurs in mid of gestation [13]. Under such conditions, mechanisms underlying oxidants (MDA)-antioxidants (SOD, GSH, GSH-Px) variability in response to progesterone or estradiol level in pregnant goats need to be elucidated. Therefore, the present study was aimed to evaluate the malondialdehyde (MDA) concentrations, antioxidant enzyme activities superoxide dismutase (SOD), glutathione peroxidase (GSH-Px) and total glutathione (GSH) and progesterone (P<sub>4</sub>) and estradiol (E<sub>2</sub>) during the different phases of pregnancy in hairy goats.

## MATERIAL AND METHODS

### Ethical Approval

Prior to the execution of study, an approval was obtained from Animal Welfare and Ethical Committee Van Yüzüncü Yıl University (VAN YUHADYEK), Van, Turkey (Decision No: 2015/07-B).

### Animals

In the present study, about twenty (n=20) 2-5 years old multiparous black hairy goats with no history of reproductive problems were selected. The research was conducted in Small Ruminants Research Center, Van Yüzüncü Yıl University, Van, Turkey. The goats were reared under semi-intensive conditions where each goat was given approximately 0.50 kg/head of concentrate daily in addition to grazing. The black hairy goat breed is present in Eastern Anatolia Region and this breed exhibits seasonal breeding pattern in this region. For optimum conditions and to minimize the variability in experiments, the peak breeding season (September to November, 2019) was chosen to conduct the study.

### Study Design, Estrus Synchronization and Pregnancy Diagnosis

Initially, the goats were synchronized using progesterone sponges (Chronogest-CR, fluorogestone acetate FGA, 20 mg Intervet®, Istanbul, Turkey) by placing intravaginally for a period of 11 days. Later, the cloprostenol (Estrumate 50 mg, Intervet®, Istanbul, Turkey) and equine chorionic gonadotropin (Chronogest/PMSG, 400 IU, Intervet®, Istanbul, Turkey) were administered intramuscularly 48 h before the removal of sponges. Estrus response detection was started after the removal of sponges till possible window of estrus display using a teaser buck at twelve hours intervals. The estrus goats were inseminated twice (First at 18-24 h and second after 36-48 h of estrus) using frozen-thawed semen ( $75 \times 10^6$  sperm/0.50 mL) through transcervical intrauterine insemination technique [14]. The pregnancy diagnosis was performed at day 35<sup>th</sup> and 42<sup>nd</sup> after insemination through transrectal ultrasonography (7.5 MHz Linear Probe, Honda HS 1500).

### Blood Sampling

The blood samples were collected through jugular vein puncture at 0, 11<sup>th</sup>, and 24<sup>th</sup> days after insemination from all experimental goats (n=20). Upon pregnancy confirmation, the blood samples were collected from pregnant goats (n=10) on 57<sup>th</sup>, 100<sup>th</sup>, 134<sup>th</sup> and 141<sup>st</sup> days of gestation and immediately after the parturition. The blood samples were centrifuged at 3000 rpm for 10 min for serum separation. Then serum samples were stored at -20°C and later assayed for oxidants (MDA), antioxidant (SOD, GSH-Px, Total GSH,) and hormone (P<sub>4</sub> and E<sub>2</sub>) titer.

### MDA, SOD, GSH and GSH-Px Analyses

Serum MDA, GSH, SOD, GSH-Px were measured by commercially available ELISA kits (LOT # OK181040, Rel Assay Diagnostics, Clinical Chemistry Solutions, Gaziantep, Turkey) as described earlier [15]. These kits were based on the spectrophotometric measurement and presence of the oxidants present in serum samples shown the colored compound. The detection principle was based

on the colored compound origination by conversion of ferrous ( $\text{Fe}^{+2}$ ) ions into ferric ( $\text{Fe}^{+3}$ ) ions as a chromogen in an acidic environment. Hydrogen peroxide was used as a calibrator index. The MDA, GSH, SOD, GSH-Px levels were measured by ELISA spectrophotometric device (TECAN Firm, Sunrise model). The results were expressed as  $\mu\text{mol H}_2\text{O}_2$  Eq/L, MDA levels as  $\mu\text{mol/L}$ , SOD activity as U/mL, GSH-Px activity as nmol/min/ml, and total GSH levels as  $\mu\text{m}$ .

### Progesterone ( $\text{P}_4$ ) and Estradiol ( $\text{E}_2$ ) Hormone Analyses

Serum  $\text{E}_2$  and  $\text{P}_4$  concentrations were measured by automated Elecsys Immunoanalyser method (Roche Diagnostics, Mannheim, Germany). The estrogen concentration was expressed in pg/ml following the method described by Souza et al.<sup>[16]</sup>, while progesterone concentration was expressed in ng/ml as reported previously<sup>[17]</sup>.

### Statistical Analyses

The data was analysed using SPSS (IBM SPSS for Windows, Ver.23) statistical software. The samples were analysed by selecting the values 0.80 and Type 1 Error 0.05. Normality distribution was calculated by using Shapiro-Wilk ( $n < 50$ ) test. In case of data unequallity, the non-parametric tests were applied to observe the difference

## RESULTS

The results of serum lipid peroxidation, enzymatic antioxidant activities and hormones at estrus time (0), gestation (day 11, 24, 57, 100, 134 and 141) and immediately after parturition were presented in Table 1. The level of MDA, SOD, GSH-Px and total GSH did not change at estrus (day 0), gestation (day 11, 24, 57, 100, 134 and 141) and after parturition. The progesterone was at peak ( $P < 0.05$ ) level at day 11 with a decreasing pattern at day 24, day 57; however, it again increased ( $P < 0.05$ ) on day 134. The least ( $P < 0.05$ ) level of progesterone was observed at the time of AI and after parturition. In contrast, the estradiol was reached to maximum level on day 0, 134<sup>th</sup> and 141<sup>st</sup> of gestation in goats.

The correlations presented in Table 2 indicated that there was no positive or negative correlations between hormones ( $\text{P}_4$  or  $\text{E}_2$ ) and oxidant (MDA) or antioxidants (SOD, GSH-Px and total GSH activity).

## DISCUSSION

In the present study, we investigated the variation of oxidants and antioxidants levels in association to progesterone and estradiol levels at estrus or during different phases of

**Table 1.** The results of serum lipid peroxidation, enzymatic antioxidant activities hormone at the time of estrus/AI (day 0), blastogenesis (days 11), embryogenesis (days 24), fetal period (days 57, 100, 134 and 141) and after parturition in pregnant goats (values are presented as means  $\pm$  SD)

Variables	Day 0 (Estrus/AI)	Day 11 (B)	Day 24 (E)	FP				After Parturition
				Day 57	Day 100	Day 134	Day 141	
MDA ( $\mu\text{mol/L}$ )	1.57 $\pm$ 0.10	1.57 $\pm$ 0.09	1.56 $\pm$ 0.08	1.53 $\pm$ 0.06	1.59 $\pm$ 0.04	1.59 $\pm$ 0.11	1.56 $\pm$ 0.15	1.59 $\pm$ 0.08
SOD (U/mL)	1.93 $\pm$ 0.57	2.19 $\pm$ 0.49	2.17 $\pm$ 0.58	2.43 $\pm$ 0.36	2.08 $\pm$ 0.43	2.01 $\pm$ 0.62	1.65 $\pm$ 0.54	2.36 $\pm$ 0.91
GSH-Px (nmol/min/mL)	176.0 $\pm$ 100.0	288.4 $\pm$ 131.1	215.4 $\pm$ 373.7	161.9 $\pm$ 60.7	225.4 $\pm$ 135.2	165.2 $\pm$ 97.7	237.6 $\pm$ 146.6	184.3 $\pm$ 71.3
Total GSH ( $\mu\text{m}$ )	3.30 $\pm$ 1.96	5.46 $\pm$ 2.48	4.07 $\pm$ 1.45	2.98 $\pm$ 1.22	4.35 $\pm$ 2.58	3.69 $\pm$ 2.79	4.48 $\pm$ 2.87	3.46 $\pm$ 1.41
$\text{P}_4$ (ng/mL)	0.32 $\pm$ 0.35 <sup>d</sup>	20.37 $\pm$ 7.54 <sup>a</sup>	8.01 $\pm$ 2.84 <sup>c</sup>	9.72 $\pm$ 3.19 <sup>c</sup>	10.82 $\pm$ 3.76 <sup>bc</sup>	15.50 $\pm$ 3.40 <sup>ab</sup>	7.48 $\pm$ 2.91 <sup>c</sup>	0.30 $\pm$ 0.08 <sup>d</sup>
$\text{E}_2$ (pg/mL)	35.20 $\pm$ 16.3 <sup>bc</sup>	13.20 $\pm$ 2.04 <sup>d</sup>	12.40 $\pm$ 1.35 <sup>d</sup>	16.00 $\pm$ 2.55 <sup>d</sup>	23.56 $\pm$ 2.60 <sup>cd</sup>	50.33 $\pm$ 12.69 <sup>ab</sup>	67.89 $\pm$ 24.43 <sup>a</sup>	39.60 $\pm$ 15.58 <sup>bc</sup>

AI: artificial insemination; B: blastogenesis; E: embryogenesis; FP: fetal period; MDA: malondialdehyde; SOD: superoxide dismutase; GSH-Px: glutathione-peroxidase; GSH: glutathione;  $\text{P}_4$ : progesterone;  $\text{E}_2$ : estradiol; Different superscripts (a, b, c, d) in same raw indicate the statistical difference ( $P < 0.05$ )

among the variables. The descriptive statistics were used to express the data (average $\pm$ Standard Deviation). The Friedman test was used to express the repeated measures for different variables at different days of pregnancy. Additionally, the Wilcoxon test was used for the analysis of dependant variables at different gestational days. Spearman correlation coefficient test was applied to determine the relationship between the measurements. A level of  $P < 0.05$  was used to denote the significance difference between the variables.

**Table 2.** Correlation analysis between hormones, oxidants and antioxidants in pregnant goats

Variables	MDA	$\text{P}_4$	$\text{E}_2$
MDA ( $\mu\text{mol/L}$ )		-0.02	0.05
SOD (U/mL)	0.21	-0.02	-0.2
GSH-Px (nmol/min/mL)	-0.01	0.06	-0.22
Total GSH ( $\mu\text{m}$ )	-0.02	0.09	-0.2

$P < 0.05$ ; r: Spearman correlation coefficient

pregnancy in goats. The MDA level and blood antioxidant status (SOD, GSH, GSH-Px) in the pregnant goats did not change at any phase of pregnancy. The data shows that no oxidative stress occurs in goats during any phase of pregnancy and oxidants-antioxidants homeostasis is maintained in response to hormonal change.

The level of MDA or CAT, SOD and GSH prior to breeding or mating time also affect the future pregnancy. Although, the ROS is a prerequisite for oocyte maturation, fertilization, embryogenesis, implantation and steroidogenesis from CL or placenta but enormous rise in ROS during synchronization through application of CIDR or vaginal sponges could adversely affect the pregnancy [4]. In contrast, we did not notice any change in MDA or antioxidants levels comparing to nonpregnant or any other phase of pregnancy in the present study, though, the goats were also synchronized using progesterone sponges by placing intravaginally as reported earlier [4,18].

Increasing trend of MDA with decreasing level of antioxidants in blood profile is an indicator of oxidative stress. The current results showed neither MDA nor antioxidants levels change in pregnant goats; although, previous studies in different species describe that pregnancy induces oxidative stress and variation in oxidant-antioxidants is observed [1,2,6]. It is also noted that fluctuations in hormonal changes during the pregnancy did not influence the oxidative and antioxidative parameters. Keeping in view of previous literature, generally rise of oxidative stress is notable during the twinning and multiple pregnancies compared to singleton [19]. In addition, the rise in MDA during pregnancy is an indication of stress that occurred due to climate change, low plane of nutrition or poor body condition [5]. In the present study, unaltered antioxidative homeostasis during the pregnancy might be due to high level of progesterone at each phase of pregnancy and high level of estradiol at estrus or parturition. Because the progesterone and estradiol have the antioxidative properties [20] and hormonal fluctuation during pregnancy could lead to imbalance between oxidative and antioxidants level as seen in multiple pregnancies [21]. In addition, similar level of oxidant-antioxidants during pregnancy in goats in the current study might be linked to similar level of progesterone production solely by CL compared to other species where shift from CL to placenta occur. In current study, we did not notice any oxidative stress in those goats carrying twins because two goats were carrying twins and rest of them having singleton. In contrast to current study, Jimoh et al. [22] observed a decline in antioxidant enzyme activity from 2<sup>nd</sup> to the 3<sup>rd</sup> trimester and assumed that this decline might be due to depletion of the antioxidants in response to higher physiological demand of immunity at neonates birth.

The goat breeds present in subtropical or in tropics shows the seasonality in breeding pattern and majority of the population exhibit estrus activity and conceived during

breeding season with small portion of goats in estrus during non-breeding season. In addition, the extreme climatic conditions during non-breeding season could be major influential factor on cyclicity, variation in serum metabolites, hormones and oxidative/antioxidative marker in sheep and goats [23]. The present study was conducted during the breeding season when small ruminants are present under comfort zone with no significant change in serum metabolites and oxidative markers [24]. The current data showed no variations in oxidative and antioxidative variables throughout the pregnancy in goats that might be linked to the season or husbandry practices. Previously, Teama [25] reported that hot climatic conditions of Egypt influenced the hormonal, serum metabolites, oxidative and antioxidative variables in cyclic goats compared to mild climate. Similarly, Rathwa et al. [24] reported that non-breeding season affect most of the biochemical, hormonal and oxidative/antioxidative markers in sheep in Indian conditions. The changes in stress hormones during non-breeding season lead to change of different body metabolites that could reinforce the incidence of oxidative stress even in nonpregnant goats in Damascus goats [26] in climatic conditions of Southern Turkey. Based on the data, it is indicated that the seasonal comparison concerning to pregnancy and oxidative stress needs to be further elucidated. It is also speculated that breed of species, feeding regimen during pregnancy, and husbandry practices might be contributory factors for onset of oxidative stress in goats.

In conclusion, no oxidative stress occurs at estrus or during pregnancy in black hairy goats carrying single fetus during the breeding season. However, the oxidative stress markers across the different breeding seasons and managerial systems could be investigated to explore the oxidative stress with ameliorative strategies in different tropical and subtropical goat breeds.

## CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

## AUTHOR CONTRIBUTIONS

This work was carried out in collaboration between all authors. Designed the experimental procedures and conducted the research work NÇ, FE, MB. Interpretation and editing of results NÇ, FE, ZN, LM. All authors read and approved the final manuscript.

## REFERENCES

1. de Souza Castagnino D, Härter CJ, Rivera AR, de Lima LD, de Oliveira Silva HG, Biagioli B, de Resende KT, de Almeida Teixeira IAM: Changes in maternal body composition and metabolism of dairy goats during pregnancy. *R Bras Zootec*, 44 (3): 92-102, 2015. DOI: 10.1590/S1806-92902015000300003
2. Mutinati M, Piccinno M, Roncetti M, Campanile D, Rizzo A, Sciorsci



- RL:** Oxidative stress during pregnancy in the sheep. *Reprod Domest Anim*, 48 (3): 353-357, 2013. DOI: 10.1111/rda.12141
- 3. Yıldız H, Kaygusuzoglu E, Kızıl Ö:** Concentrations of serum vitamins A, E and C and  $\beta$ -carotene during pregnancy in cows. *Bull Vet Inst Pulawy*, 49, 199-202, 2005.
- 4. Kuru M, Ögün M, Kulaksız R, Kükürt A, Oral H:** Comparison of oxidative/nitrosative stress, leptin and progesterone concentrations in pregnant and non-pregnant Abaza goats synchronized with controlled internal drug release application. *Kafkas Univ Vet Fak Derg*, 24 (6): 887-892, 2018. DOI: 10.9775/kvfd.2018.20222
- 5. Mohebbi-Fani M, Mirzaei A, Nazifi S, Tabandeh MR:** Oxidative status and antioxidant enzyme activities in erythrocytes from breeding and pregnant ewes grazing natural pastures in dry season. *Revue Méd Vét*, 163 (10): 454-460, 2012.
- 6. Dobson H, Rowan TG, Kippax IS, Humblot P:** Assessment of fetal number, and fetal and placental viability throughout pregnancy in cattle. *Theriogenology*, 40 (2): 411-425, 1993. DOI: 10.1016/0093-691x(93)90278-d
- 7. Erisir M, Benzer F, Kandemir FM:** Changes in the rate of lipid peroxidation in plasma and selected blood antioxidants before and during pregnancy in ewes. *Acta Vet Brno*, 78, 237-242, 2009. DOI: 10.2754/avb200978020237
- 8. Aydın I, Köse AM:** Serum oxidative status and biochemical parameter levels during in Saanen goats. *Eurasian J Vet Sci*, 31 (4): 197-203, 2015. DOI: 10.15312/EurasianJVetSci.2015413490
- 9. Mohebbi-Fani M, Mirzaei A, Nazifi S, Shabbooei Z:** Changes of vitamins A, E, and C and lipid peroxidation status of breeding and pregnant sheep during dry seasons on medium-to-low quality forages. *Trop Anim Health Prod*, 44, 259-265, 2012. DOI: 10.1007/s11250-011-0012-1
- 10. Ryahi SH, Aslani MR, Mohebi AN:** The evaluation of stress oxidative status, some serum biochemical parameters and trace elements of Saanen goats during peripartur period. *Iran J Vet Clin Sci*, 12, 111-120, 2018.
- 11. Al-Gubory KH, Bolifraud P, Germain G, Nicole A, Ceballos-Bicot I:** Antioxidant enzymatic defence systems in sheep corpus luteum throughout pregnancy. *Reproduction*, 128, 767-774, 2004. DOI: 10.1530/rep.1.00389
- 12. Garrel C, Fowler PA, Al-Gubory KH:** Developmental changes in antioxidant enzymatic defences against oxidative stress in sheep placentomes. *J Endocrinol*, 205, 107-116, 2010. DOI: 10.1677/JOE-09-0362
- 13. Buttle HL:** The maintenance of pregnancy in hypophysectomized goats. *J Reprod Fertil*, 52 (2): 255-260, 1978. DOI: 10.1530/jrf.0.0520255
- 14. Dogan I, Nur Z, Gunay U, Sagirkaya H, Soylu MK, Sonmez C:** Estrous synchronization during the natural breeding season in Anatolian black does. *Vet Med*, 50, 33-38, 2005.
- 15. Erel O:** A new automated colorimetric method for measuring total oxidant status. *Clin Biochem*, 38 (12): 1103-1111, 2005. DOI: 10.1016/j.clinbiochem.2005.08.008
- 16. Souza, SS, Alves BG, Alves KA, Santos JDR, Diogenes YP, Bhat MH, Melo LM, Freitas VJF, Teixeira DIA:** Relationship of doppler velocimetry parameters with antral follicular population and oocyte quality in Canindé goats. *Small Ruminant Res*, 141, 39-44, 2016. DOI: 10.1016/j.smallrumres.2016.06.011
- 17. Cinar M, Ceyhan A, Yilmaz O, Erdem H:** Effect of estrus synchronization protocols including PGF<sub>2a</sub> and GnRH on fertility parameters in hair goats during breeding season. *J Anim Plant Sci*, 27 (4): 1083-1087, 2017.
- 18. Oral H, Ögün M, Kuru M, Kaya S:** Evaluation of certain oxidative stress parameters in heifers that were administered short term PRID. *Kafkas Univ Vet Fak Derg*, 21 (4): 569-573, 2015. DOI: 10.9775/kvfd.2015.12969
- 19. Abd El-Hameed AR, Mahmoud KGM, Nawito MF, Sosa ASA, Ezzo OH:** Progesterone concentration, oxidants and antioxidants indices in singleton and multiple bearings ewes and does. *Biosci Res*, 15 (4): 3881-3887, 2018.
- 20. Chainy GBN, Sahoo DK:** Hormones and oxidative stress: An overview. *Free Radic Res*, 54 (1): 1-26, 2019. DOI: 10.1080/10715762.2019.1702656
- 21. Abdel-Ghani MA, El-Sherry TM, Hayder M, Abou-Khalil NS:** Profile of peroxidative injury and antioxidant indicators in singleton, twins and multiple bearing goats throughout pregnancy. *Asian Pac J Reprod*, 5 (5): 400-405, 2016. DOI: 10.1016/j.apjr.2016.07.010
- 22. Jimoh AO, Ojo OA, Ihejirika UDG:** Metabolic and oxidative status of West African dwarf does at different reproductive stages in southwest Nigeria. *Bull Natl Res Cent*, 43:190, 2019. DOI: 10.1186/S42269-019-0223-6
- 23. Ribeiro MN, Ribeiro NL, Bozzi R, Costa RG:** Physiological and biochemical blood variables of goats subjected to heat stress - A review. *J Appl Anim Res*, 46 (1): 1036-1041, 2018. DOI: 10.1080/09712119.2018.1456439
- 24. Rathwa SD, Vasava AA, Pathan MM, Madhira SP, Patel YG, Pande AM:** Effect of season on physiological, biochemical, hormonal, and oxidative stress parameters of indigenous sheep. *Vet World*, 10 (6): 650-654, 2017. DOI: 10.14202/vetworld.2017.650-654
- 25. Teama FEI:** Evaluation of some oxidative-stress and antioxidant markers in goats during estrous cycle under Egyptian environmental conditions. *R Bras Zootec*, 47:e20160382, 2018. DOI: 10.1590/rbz4720160382
- 26. Saribay MK, Naseer Z, Doğruer G, Özsoy B, Ateş CT:** Variations in serum metabolites in response to season, cyclicity, and pregnancy in estrus-synchronized damascus goats. *Trop Anim Health Prod*, 52, 1519-1525, 2020. DOI: 10.1007/s11250-019-02131-6