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Bu Sayının Hakem Listesi (alfabetik sıra)
The Referees List of This Issue (in alphabetical order)

AÇIKGÖZ Zümrüt	Ege Üniversitesi Ziraat Fakültesi Zootečni Bölümü
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GÜL Mehmet	Atatürk Üniversitesi Veteriner Fakültesi Hayvan Besleme ve Beslenme Hastalıkları Anabilim Dalı
GÜLBAHAR M. Yavuz	Ondokuz Mayıs Üniversitesi Veteriner Fakültesi Patoloji Anabilim Dalı
GÜLER Leyla	Konya Veteriner Kontrol ve Araştırma Enstitüsü
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İMİK Halit	Atatürk Üniversitesi Veteriner Fakültesi Hayvan Besleme ve Beslenme Hastalıkları Anabilim Dalı
KABAK Murat	Ondokuz Mayıs Üniversitesi Veteriner Fakültesi Anatomi Anabilim Dalı
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ÖZTÜRK Dilek	Mehmet Akif Ersoy Üniversitesi Veteriner Fakültesi Mikrobiyoloji Anabilim Dalı
ÖZTÜRK Ergin	Ondokuz Mayıs Üniversitesi Ziraat Fakültesi Yemler ve Hayvan Besleme Anabilim Dalı
PAPUÇCUOĞLU Serhat	İstanbul Üniversitesi Veteriner Fakültesi Dölerme ve Suni Tohumlama Anabilim Dalı
PEKCAN Zeynep	Kırıkkale Üniversitesi Veteriner Fakültesi Cerrahi Anabilim Dalı
SEZEN Yavuz	Gebze İleri Teknoloji Enstitüsü
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Bu Sayının Hakem Listesi (alfabetik sıra)
The Referees List of This Issue (in alphabetical order)

TİMURKAN Sema	Fırat Üniversitesi Veteriner Fakültesi Histoloji ve Embriyoloji Anabilim Dalı
TÜRKER Hakan	Abant İzzet Baysal Üniversitesi Fen Edebiyat Fakültesi Biyoloji Bölümü
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YILDIZ Hüseyin	Uludağ Üniversitesi Veteriner Fakültesi Anatomi Anabilim Dalı
YÖRÜK Mecit	Yüzüncü Yıl Üniversitesi Veteriner Fakültesi Histoloji ve Embriyoloji Anabilim Dalı

İÇİNDEKİLER (CONTENTS)

ARAŞTIRMA MAKALELERİ (RESEARCH ARTICLES)

Sayfa
(Page)

Studies on Chewing Lice (Phthiraptera: Amblycera, Ischnocera) Species from Domestic and Wild Birds in Turkey DIK B, YAMAC E, USLU U	553
Investigation of the Effects of Seasonal Variation on the Microbial-N Flow to the Small Intestine and Prediction of Feed Intake in Grazing Karayaka Sheep SALMAN M, CETINKAYA N, SELCUK Z, GENÇ B	561
Effects of Graded Levels of Crude Glycerine Addition to Diets on Growth Performance, Carcass Traits and Economic Efficiency in Broiler Chickens SEHU A, KUCUKERSAN S, COSKUN B, KOKSAL BH	569
Cloning and Expression of <i>Cellulosimicrobium cellulans</i> β-1,3-Glucanase Gene in <i>Lactobacillus plantarum</i> to Create New Silage Inoculant for Aerobic Stability OZCAN BD, OZCAN N, BAYLAN M, GUZEL AI	575
The Effects of D-Ala², D-Leu⁵-Enkephalin (DADLE)'s Continuous Perfusion on Erythrogram and Biochemical Profile in Rats AKYAZI I, ERASLAN E, AKTARAN BALA D, CIRAKLI ZL, COTELIOGLU U	583
Parasitic Aortitis due to <i>Onchocerca armillata</i> in Slaughtered Cattle in the Southeastern Region of Turkey OZYILDIZ Z, YILMAZ R, OZSOY SY, OZKUL IA	589
The Effects of Dietary Rosemary (<i>Rosmarinus officinalis</i> L.) Oil Supplementation on Performance, Carcass Traits and Some Blood Parameters of Japanese Quail Under Heat Stressed Condition CIFTCI M, SIMSEK UG, AZMAN MA, CERCI IH, TONBAK F	595
The Relationships Among Some Udder Traits and Somatic Cell Count in Holstein-Friesian Cows ALIC URAL D	601
Effects of Acrylamide Treatment on Oxidant and Antioxidant Levels in Rats YERLIKAYA FH, TOKER A, YENER Y	607
Effect of Hyaluronic Acid/Carboxymethylcellulose and Flunixin Meglumine Combination on the Prevention of Postoperative Intraabdominal Adhesions: An Experimental Study in Rabbits KOM M	613
Risk Factors Associated with Passive Immunity, Health, Birth Weight and Growth Performance in Lambs: II. Effects of Passive Immunity and Some Risk Factors on Growth Performance During the First 12 Weeks of Life GOKCE E, ATAKISI O, KIRMIZIGUL AH, ERDOGAN HM	619
Effects of FGA Sponge and Ovsynch Based Protocols on Reproductive Performance of Fat-tailed Ewes During the Breeding Season KULAKSIZ R, UCAR O, DASKIN A	629
The Axon Number of the Rat Sciatic Nerve: A Stereological Study RAGBETLI C, YALAMA M, KATI I, ULKAY MB, ESENER OB, BOZKURT HH, KOYUN N, RAGBETLI MC	635
The Impacts of Prebiotic Supplementation on Humoral and Cell-Mediated Immune Responses of Broilers Under Combined Stresses Caused by Feed Restriction and <i>Salmonella enteritidis</i> Challenge SADEGHI AA, MADDAGHAN A, SHAWRANG P, AMINAFSHAR M	641
Effects of Low Doses of Bisphenol A on Primordial Germ Cells in Zebrafish (<i>Danio rerio</i>) Embryos and Larvae AKBULUT C, KIZIL C, YON ND	647
Importance of Vaccination Against Atrophic Rhinitis in Pigs on Average Daily Gain and Mortality Rate STOJANAC N, GAGRČIN M, STEVANČEVIĆ O, UROŠEVIĆ M, MLAĐAN V	655
Kontrol Günü Süt Verimlerinin Zaman Serisi Yöntemi ile Modellenmesi KARAMAN E, FIRAT MZ	659

Molecular Characteristics of *Staphylococcus aureus* Isolates from Buffaloes Milk

SHAYEGH J, BARZEGARI A, MIKAILI P 665

Comparison of Growth Curves of Broiler under Different Stocking Densities by Gompertz Model

SEKEROGLU A, TAHTALI Y, SARICA M, GULAY MS, ABACI SH, DUMAN M 669

Effects of Dietary Inclusion of Plant Extract Mixture and Copper into Layer Diets on Egg Yield and Quality, Yolk Cholesterol and Fatty Acid Composition

KAYA A, KAYA H, MACIT M, CELEBI S, ESENBUGA N, YORUK MA, KARAOGLU M 673

The Statistical Analysis of Some Volumetric Measurements in the Japanese Quails' Head with Different Feather Color: A Computed Tomography Study

DUYMUS M, DEMIRASLAN Y, AKBULUT Y, ORMAN G, ASLAN K, OZCAN S 681

The Anatomy of the Cardiac Veins in Storks (*Ciconia ciconia*)

YOLDAS A, OZMEN E, AKSOY G 687

Cirit Atlarında Vücut Ölçüleri

YILDIRIM F, YILDIZ A 693

Effect of Varieties on Potential Nutritive Value of Pistachio Hulls

BOGA M, GUVEN I, ATALAY AI, KAYA E 699

KISA BİLDİRİ (SHORT COMMUNICATION)**Geleneksel Yöntemle Üretilen Sucuklarda *Listeria monocytogenes*, *Staphylococcus aureus* ve Koliform Varlığının Araştırılması**

YALCIN H, CAN OP 705

Comparison of Three Methods for Routine Detection of *Staphylococcus aureus* Isolated from Bovine Mastitis

RUSENOVA N, GEBREYES W, KOLEVA M, MITEV J, PENEV T, VASILEV N, MITEVA T 709

OLGU SUNUMU (CASE REPORT)**Bir Ceylanda (*Gazella gazella*) Generalize Reaktif Amiloidoz Olgusu**

OZDEMİR O, HATİPOĞLU F, KARAMAN M 713

Türkiye'de Köpeklerde *Babesia canis canis*'in Klinik ve Parazitolojik Olarak İlk Tespiti

GOKCE E, KIRMIZIGUL AH, TASCI GT, UZLU E, GUNDUZ N, VATANSEVER Z 717

A Case Report: Isolation of *Alysiella filiformis* from Pig's Lungs

ZUTIC J, RADOSAVLJEVIC V, RADANOVIC O, IVETIC V, PAVLOVIC I, ZUTIC M 721

DERLEME (REVIEW)**Possible Natural Toxins in Organic Livestock Farming**

SEKKIN S, KUM C 725

Studies on Chewing Lice (Phthiraptera: Amblycera, Ischnocera) Species from Domestic and Wild Birds in Turkey ^{[1] [2]}

Bilal DİK ¹  Elif YAMAÇ ² Uğur USLU ¹

[1] This study was supported by Selçuk University Research Fund (BAP Project number: 11401042)

[2] This study has been presented as poster at the 21th National Biology Congress, Ege University, Science Faculty, İzmir, 2-7 September 2012

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Summary

This study was performed to detect chewing lice species occurring on domestic and wild birds in Turkey between October 2010 and May 2012. For this aim, the birds were brought to Veterinary Faculty of Selçuk University and Biology Department of Science Faculty of Anadolu University were examined for their lice. The louse specimens collected from the birds were preserved in eppendorf tubes contains ethyl alcohol 70%. They were cleared in Potassium Hydroxide (KOH) 10% for a day, washed within distilled water for a day and transferred to alcohol 70%, 80%, 90% and 99% at consecutive days. Then, they were mounted on the slides in Canada balsam and identified to species. Among studied 26 bird species in 24 genera and 17 families belonging to 9 orders, seventeen species (65.38%) were found to be infested with lice and no louse specimen was found in 9 bird species (34.62%). The forty-one specimens (48.23%) of examined 85 birds were found to be infested with the lice, and 32 lice species were detected. Besides, the genera *Falcolipeurus*, *Coloceras* and *Struthiolipeurus*, the species; *Brueelia apiastri* (Denny, 1842); *Brueelia munda* (Nitzsch, 1866); *Coloceras piageti* (Johnston ve Harrison, 1912); *Coloceras israelensis* (Tendeiro, 1974); *Degeeriella nesus* (Giebel, 1866); *Degeeriella phylctopygus* (Nitzsch, 1861); *Degeeriella rufa* (Burmeister, 1838); *Falcolipeurus quadripustulatus* (Burmeister, 1838); *Laemobothrion vulturis* (Fabricius, J.C., 1775); *Menacanthus oriol* (Blagovestchensky, 1951); *Phlopterus fringillae* (Scopoli, 1772); *Struthiolipeurus struthionis* (Gervais, 1844) were recorded for the first time from the birds in Turkey. *Columbicola columbae* was found on the Long-eared Owl as a first record.

Keywords: *Ardeicola*, *Falcolipeurus*, *Brueelia*, *Ciconophilus*, *Coloceras*, *Colpocephalum*, *Laemobothrion*, *Piagetiella*, *Struthiolipeurus*, *Phlopterus*

Türkiye'deki Evcil ve Yabani Kuşlarda Bulunan Çiğneyici Bit (Phthiraptera: Amblycera, Ischnocera) Türleri Üzerine Araştırmalar

Özet

Bu araştırma Türkiye'deki evcil ve yabani kuşlarda görülen bit türlerini belirlemek amacıyla Ekim 2010 -Mayıs 2012 tarihleri arasında yapılmıştır. Bu amaçla Selçuk Üniversitesi Veteriner Fakültesi Kliniklerine ve Anadolu Üniversitesi Fen Fakültesi Biyoloji Bölümü'ne getirilen hasta, yaralı veya ölü kuşlarla, karayollarında ölü olarak bulunan kuşlar laboratuvar ortamında bit yönünden incelenmiştir. Toplanan bitler, içinde %70 alkol bulunan eppendorf tüplerde saklanmıştır. Bitler %10'luk Potasyum Hidroksit (KOH) içinde bir gün süreyle saydamlaştırılmış, 24 saat süreyle distile suda yıkanmış ve birer gün süreyle %70, %80, %90 ve %99'luk alkol serilerinden geçirildikten sonra, Kanada balsamı ile lam üzerine yapıştırılmış ve tür seviyesinde teşhis edilmişlerdir. İncelenen 9 takım, 17 aile, 24 cinse ait 26 kuş türünden 17'si (%65.38) bitlerle enfeste bulunmuş, 9 kuş (%34.62) türünde ise herhangi bir bite tesadüf edilmemiştir. Muayene edilen 85 kuşun 41'i (%48.23) bitlerle enfeste bulunmuş ve 32 bit türü saptanmıştır. *Falcolipeurus*, *Coloceras* ve *Struthiolipeurus* cinsleri ile; *Brueelia apiastri* (Denny, 1842); *Brueelia munda* (Nitzsch, 1866); *Coloceras piageti* (Johnston ve Harrison, 1912); *Coloceras israelensis* (Tendeiro, 1974); *Degeeriella nesus* (Giebel, 1866); *Degeeriella phylctopygus* (Nitzsch, 1861); *Degeeriella rufa* (Burmeister, 1838); *Falcolipeurus quadripustulatus* (Burmeister, 1838); *Laemobothrion vulturis* (Fabricius, J.C., 1775); *Menacanthus oriol* (Blagovestchensky, 1951); *Phlopterus fringillae* (Scopoli, 1772); *Struthiolipeurus struthionis* (Gervais, 1844) Türkiye'deki kuşlardan ilk kez bildirilmiştir. Ayrıca, Kulaklı Orman Baykuşunda *C. columbae*'ye ilk kez rastlanmıştır.

Anahtar sözcükler: *Ardeicola*, *Falcolipeurus*, *Brueelia*, *Ciconophilus*, *Coloceras*, *Colpocephalum*, *Laemobothrion*, *Piagetiella*, *Struthiolipeurus*, *Phlopterus*



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INTRODUCTION

One of the most common bird diseases is parasitic ones which are mostly caused by ectoparasites. Lice and mites are known as common ectoparasites of the birds. Although parasitic lice species on birds were previously considered order or suborder Mallophaga, according to recent classification, bird lice are belonging to the suborders Ischnocera and Amblycera in the order Phthiraptera. More than 250 genera and 6.000 lice species were described until now; about 4.500 species are valid species ^[1].

Although, Turkey has 468 species of bird ^[2], there is limited number of investigations about the lice species found on the birds in Turkey ^[3-6]. The number of lice species reported from the birds in Turkey has been reached about 100 species ^[7-38]. Although, most of the studies about lice species have been focused on only chickens ^[3,7,9,11,39-41].

As a result of the published studies, some information has been obtained with regard to the lice fauna found on bird species in Turkey. On the other hand, existing information has included 20% of known bird species in Turkey. Thus, it can be argued that our knowledge is not sufficient and we need to perform much more studies on the lice fauna of birds in Turkey. This study was performed to detect chewing lice species occurring on domestic and wild birds in Turkey.

MATERIAL and METHODS

This study was carried out to detect the lice species on the birds in Turkey between October 2010 and May 2012. The chewing lice were collected from 85 domestic

and wild bird specimens that were found to be injured or died in the field or obtained from hunters during legal bird hunting seasons. The feathers of each bird were carefully examined and lice were removed with forceps. After visual examination, Bolfo (1% (w/w) 2-Isopropoxyphenyl N-methyl-carbamate) or Biyo Avispray (0.09% (v/w) Tetramethrin and 0.45 (w/w) piperonyl butoxide) were applied to birds placed in a white basin and were kept at least 15 min. The collected lice specimens were fixed in 70% ethanol. After clearing in 10% KOH for 24 h, they were washed with distilled water for 24 h and passed in alcohol series 70%, 80%, 90% and 99% in four consecutive days. The lice specimens were mounted on the slides in Canada balsam by using stereo-zoom microscope, and they were kept for drying in an incubator 50°C for a few weeks. The lice species were examined under light microscope and were identified to species according to the relevant literatures ^[42-47].

RESULTS

Only one bird individual in the orders Struthioniformes, Pelecaniformes, Ciconiiformes and Coraciiformes, two bird samples in the orders Columbiformes and Psittaciformes, three bird samples in the order Strigiformes and eight bird specimens in the orders Falconiformes and Passeriformes were examined. While, seventeen of twenty six bird species (65.38%) were found to be infested, no louse was found on studied nine bird species (34.62%). It was determined that forty one of a total of eighty five birds (48.23%) has one louse species on their body, at least. A total of 32 lice species were determined, which of 11 and 21 species belongs to the suborders Amblycera and Ischnocera, respectively. The comprehensive information about the lice species found on the birds was presented in Table 1.



Fig 1. *Brueelia apiastri*, female, original
Şekil 1. *Brueelia apiastri*, dişi, orijinal

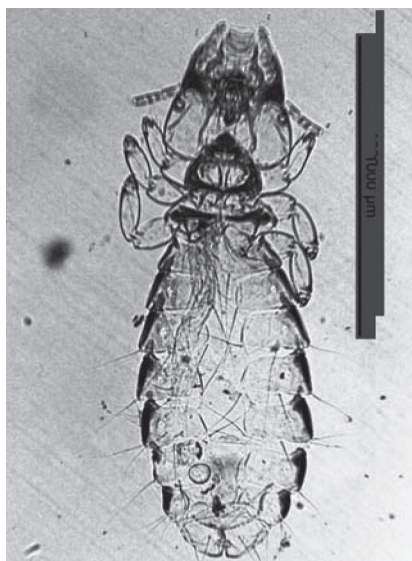


Fig 2. *Brueelia munda*, female, original
Şekil 2. *Brueelia munda*, dişi, orijinal

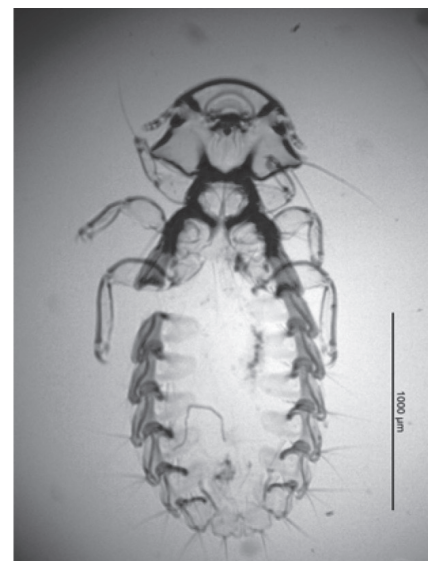


Fig 3. *Coloceras israelensis*, female, original
Şekil 3. *Coloceras israelensis*, dişi, orijinal

Table 1. Studied bird species and detected lice species**Tablo 1.** İncelenen kuş türleri ve tespit edilen bit türleri

Order	Family	Genus	Species	Examined Bird Number	Infested Bird Number	Lice Species
Struthioniformes	Struthionidae	<i>Struthio</i>	<i>S. camelus</i> Ostrich	1	1	<i>Struthiolipeurus struthionis</i> *
Pelecaniformes	Pelecanidae	<i>Pelecanus</i>	<i>P. onocrotalus</i> Great White Pelican	2	2	<i>Colpocephalum eucarenum</i> <i>Pectinopygus forficulatus</i> <i>Piagetiella titan</i>
Ciconiiformes	Ciconiidae	<i>Ciconia</i>	<i>C. ciconia</i> White Stork	1	1	<i>Ardeicola ciconiae</i> <i>Ciconophilus quadripustulatus</i> <i>Colpocephalum zebra</i> <i>Neophilopterus incompletus</i>
Falconiformes	Accipitridae	<i>Aegypius</i>	<i>A. monachus</i> Eurasian Black Vulture	1	1	<i>Falcolipeurus quadripustulatus</i> * <i>Laemobothrion vulturis</i> * <i>Colpocephalum trachelioti</i>
		<i>Circus</i>	<i>C. aeruginosus</i> WesternMarsh-harrier	2	1	<i>Colpocephalum turbinatum</i> ** <i>Degeeriella fusca</i>
		<i>Accipiter</i>	<i>A. nisus</i> Eurasian Sparrowhawk	3	2	<i>Colpocephalum nanum</i> <i>Degeeriella nisus</i> *
			<i>A. brevipes</i> Levant Sparrowhawk	1	-	-
		<i>Buteo</i>	<i>B. buteo</i> Common Buzzard	2	2	<i>Degeeriella fulva</i> <i>Kurodaia fulvofasciata</i>
			<i>B. rufinus</i> Long-legged Buzzard	13	11	<i>Colpocephalum nanum</i> <i>Craspedorrhynchus platystomus</i> <i>Degeeriella fulva</i> <i>Laemobothrion maximum</i>
	Falconidae	<i>Pernis</i>	<i>P. apivorus</i> European Honey-buzzard	1	1	<i>Degeeriella phlyctopygus</i> *
		<i>Falco</i>	<i>F. tinnunculus</i> Common Kestrel	1	1	<i>Degeeriella rufa</i> *
Columbiformes	Columbidae	<i>Columba</i>	<i>C. livia</i> Rock Pigeon	4	4	<i>Columbicola columbae</i> <i>Coloceras israelensis</i> *
		<i>Streptopelia</i>	<i>S. decaocto</i> Eurasian Collared-dove	2	1	<i>Coloceras piageti</i> *
Psittaciformes	Psittacidae	<i>Melopsittacus</i>	<i>M. undulatus</i> Budgerigar	8	-	-
	Cacatuidae	<i>Nymphicus</i>	<i>N. hollandicus</i> Cockatiel	1	-	-
Strigiformes	Strigidae	<i>Athene</i>	<i>A. noctua</i> Little Owl	2	-	-
		<i>Asio</i>	<i>A. otus</i> Long-eared Owl	4	4	<i>Strigiphilus barbatus</i> <i>Columbicola columbae</i> **
Coraciiformes	Meropidae	<i>Merops</i>	<i>M. apiaster</i> European Bee-eater	1	1	<i>Brueelia apiastri</i> * <i>Meropoecus sp</i>
Passeriformes	Motacillidae	<i>Motacilla</i>	<i>M. alba</i> White Wagtail	1	-	-
	Sylviidae	<i>Sylvia</i>	<i>S. atricapilla</i> Blackcap	1	-	-
	Corvidae	<i>Pica</i>	<i>P. pica</i> Eurasian Magpie	1	-	-
	Oriolidae	<i>Oriolus</i>	<i>O. oriolus</i> Eurasian Golden Oriole	1	1	<i>Brueelia munda</i> * <i>Menacanthus oriolus</i> *
	Sturnidae	<i>Sturnus</i>	<i>S. vulgaris</i> Common Starling	4	4	<i>Brueelia nebulosa</i> <i>Menacanthus eurysternus</i> **
	Passeridae	<i>Passer</i>	<i>P. domesticus</i> House Sparrow	22	3	<i>Menacanthus eurysternus</i> ** <i>Philopterus fringillae</i> *
	Fringillidae	<i>Fringilla</i>	<i>F. coelebs</i> Eurasian Chaffinch	1	-	-
		<i>Carduelis</i>	<i>C. cannabina</i> Eurasian Linnet	4	-	-
Total				85	41	

* First record in Turkey, ** First record from this host in Turkey

Brueelia apiastri (Denny, 1842) (Fig. 1); *Brueelia munda* (Nitzsch, 1866) (Fig. 2); *Coloceras israelensis* (Tendeiro, 1974) (Fig. 3); *Coloceras piageti* (Johnston ve Harrison, 1912) (Fig. 4); *Degeeriella nisus* (Giebel, 1866) (Fig. 5); *Degeeriella phylctopygus* (Nitzsch, 1861) (Fig. 6); *Degeeriella rufa* (Burmeister, 1838) (Fig. 7); *Falcolipeurus quadripustulatus* (Burmeister, 1838) (Fig. 8); *Laemobothrion vulturis* (Fabricius, J.C., 1775) (Fig. 9); *Menacanthus oriolis* (Blagovestchensky, 1951) (Fig. 10); *Philopterus fringillae* (Scopoli, 1772) (Fig. 11); *Struthiolipeurus struthionis* (Gervais, 1844) (Fig. 12) were reported for the first time in Turkey.

DISCUSSION

It was stated that, 3910 out of 5642 described lice species from birds had been approved as valid species

about ten years ago. Among them, 2737 and 1173 species are belonging to the suborders Ischnocera and Amblycera, respectively^[1]. In spite of increased number of investigations about lice fauna found on bird species; the numbers of investigated bird and reported lice species are still insufficient in Turkey. Approximately, 20% of total bird species have been examined for their lice species and no louse was found on some of them. The results of the studies carried out on chewing lice fauna in the birds in Turkey until now have showed that, the numbers of the lice species recorded have been reached about 100.

In this study, 85 bird individuals of 26 species in 24 genera, 17 families belonging to the 9 orders were examined. Therefore we can argue that the presented study is the most comprehensive investigation on the lice species on birds in Turkey. Forty-one bird individuals

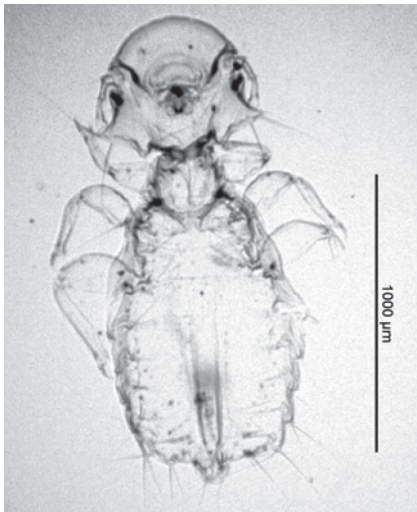


Fig 4. *Coloceras piageti*, male, original
Şekil 4. *Coloceras piageti*, erkek, orijinal



Fig 5. *Degeeriella nisus*, female, original
Şekil 5. *Degeeriella nisus*, dişi, orijinal



Fig 6. *Degeeriella phylctopygus*, female, original
Şekil 6. *Degeeriella phylctopygus*, dişi, orijinal



Fig 7. *Degeeriella rufa*, female, original
Şekil 7. *Degeeriella rufa*, dişi, orijinal



Fig 8. *Falcolipeurus quadripustulatus*, female, orig.
Şekil 8. *Falcolipeurus quadripustulatus*, dişi, orij.

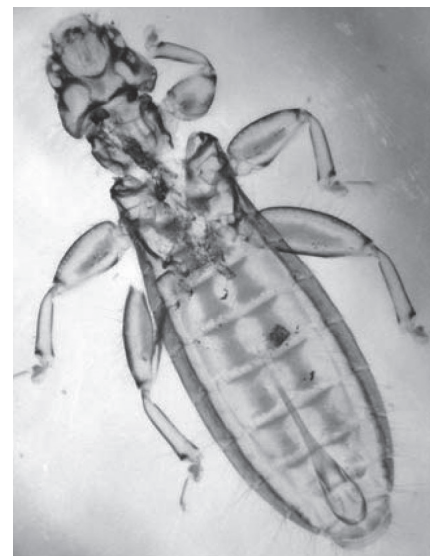


Fig 9. *Laemobothrion vulturis*, male, original
Şekil 9. *Laemobothrion vulturis*, erkek, orijinal



Fig 10. *Menacanthus orioli*, female, original

Şekil 10. *Menacanthus orioli*, dişi, orijinal

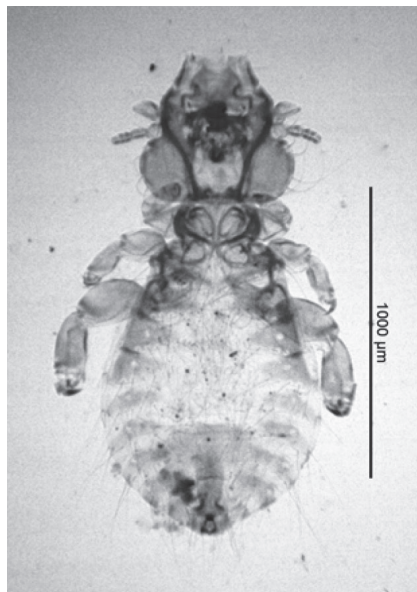


Fig 11. *Philopterus fringillae*, female, original

Şekil 11. *Philopterus fringillae*, dişi, orijinal



Fig 12. *Struthiolipeurus struthionis*, male, original

Şekil 12. *Struthiolipeurus struthionis*, erkek, orij.

belonging to seventeen species (65.38%) were found to be infested by the lice, while no louse was found on nine bird species (34.62%). Totally, 32 lice species, which of 11 species in six genera and 21 species in 13 genera are belonging to the suborders Amblycera and Ischnocera were detected, respectively. While the bird individuals and species numbers was not same for each order, infestation rates were 100% for Struthioniformes, Pelecaniformes, Ciconiiformes and Coraciiformes, 83.33% for Columbiformes, 79.17% for Falconiformes, 66.66% for Strigiformes and 22.86% for Passeriformes. But, it could not be detected any louse specimen on the order Psittaciformes.

The numbers of recorded lice species on the orders Falconiformes and Passeriformes were more than the others, due to the bird samples examined in these orders were much more from the others. Twenty-four individuals belonging to the eight bird species in the order Falconiformes were examined and 13 lice species were detected. For the order Passeriformes, total 35 individuals belonging to eight bird species were examined and five lice species were identified. Although, the order Passeriformes was the second order for lice species; the numbers of lice species detected on the birds in this order were less than the others, compared to the examined bird numbers.

Chewing lice usually parasitise on their specific host, but some of them are cosmopolitan and can found on different bird species. Twenty-eight lice species were found on their normal host; *C. columbae*, *C. nanum*, *D. fulva* and *M. eurysternus* were occurred on different bird species, as well as their normal hosts, in this study. *Colpocephalum nanum* was collected from Eurasian Sparrow Hawk and Long-legged Buzzard in this study. This louse species

has also been reported on common Buzzard and Long-legged Buzzard in Turkey ^[16,21,27], previously. *Menacanthus eurysternus* has been recorded from several bird species throughout the world ^[1], but not in Turkey until now. Dik et al. ^[41] recorded that, *Brueelia nebulosa*, *Sturnidoecus sturni* and *Myrsidea cucullaris*, but no found *Menacanthus eurysternus* on Eurasian Starlings (*Sturnus vulgaris*) in a previous study. Three European Starlings and two House Sparrows (*Passer domesticus*) were found to be infested by *M. eurysternus*. Thus, *M. eurysternus* is recorded from these hosts for the first time and this species a new record for Turkey lice fauna. *Columbicola columbae* is found on rock pigeon (*Columba livia*). However, this species was found on a Long-eared Owl (*Asio otus*) as well as Rock Pigeon in this study.

In a previous study, no louse species was found on 13 house sparrows ^[31]. Seventy-two individuals from over 10 different bird species belonging to the order Passeriformes had been examined near Aras River in Kars and infestation rate was determined as 12.50% ^[35]. In another report for louse species found on the birds belonging to the order Passeriformes in Kuyucuk Lake in Kars, 51 bird specimens representing 22 genera were examined and five lice species were detected from 11 bird individuals, and infestation rate was detected as 21.57% ^[37]. In present study, 35 bird specimens belonging to the eight species in the order Passeriformes were examined, infestation rate was found as 22.86% and five lice species; *B. munda*, *B. nebulosa*, *M. eurysternus*, *M. orioli* and *P. fringillae* were detected. Infestation rate was relatively low in this study as well as the other studies ^[31,35,37], because of only three of examined 22 House Sparrows had been infested for the louse specimens. This result related House Sparrow in this study is similar with the results of Dik ^[31] and its reasons

have been discussed in a previous paper ^[37]. As a result, *B. munda*, *M. oriol* and *P. fringillae* detected on Passeriformes birds were recorded for the first time in Turkey.

The genera *Colpocephalum*, *Kurodaia*, *Laemobothrion*, *Craspedorrhynchus* and *Degeeriella* were reported from the Falconiformes birds, previously. *Laemobothrion maximum*, *C. nanum*, *Colpocephalum* sp., *D. fulva* and *C. platystomus* have been reported from Long legged Buzzard (*Buteo rufinus*) ^[16,20,21,24,27,31,34]. We have also confirmed the presence of *L. maximum*, *C. nanum*, *C. platystomus* and *D. fulva* on Long-legged Buzzard in this study. Besides, it was found that prevalence of *C. platystomus* and *D. fulva* were higher and of *L. maximum* was lower than the other species. Dik and Yamaç ^[24] have reported that, *C. trachelioti* was found on Eurasian Black Vulture (*Aegypius monachus*) in Turkey. *Colpocephalum trachelioti*, *F. quadripustulatus* and *L. vulturis* were found on the Eurasian Black Vulture in this study. The genus *Falcolipeurus* and the species *L. vulturis* were firstly reported from Turkey. *Kurodaia fulvofasciata* was found on Common Buzzard (*Buteo buteo*) and reported as a first record for Turkey in a previous study ^[31]. İnci et al. ^[34] determined that all of lice species, except *K. fulvofasciata*, found on Common Buzzards also has been detected on the Long-legged Buzzards. Two Common Buzzard samples examined and they were infested with *D. fulva* and *K. fulvofasciata* in this study. Besides, İnci et al. ^[34] have also recorded that, *D. fulva* and *Colpocephalum* sp. from European Honey-Buzzards in Middle Anatolia. Additionally, *D. phlyctopygus* had been found on European Honey-Buzzard as a first record for Turkey in that study ^[34]. In previously studies; the Western Marsh-Harrier (*Circus aeruginosus*), Northern Goshawk (*Accipiter gentilis*) and Common Kestrel (*Falco tinnunculus*) were reported as hosts of *D. fusca* ^[27], *Colpocephalum polonium* ^[36], and *Colpocephalum subzerafae* and *Laemobothrion tinnunculi* ^[38], respectively in Turkey. There was no detected any louse species on the Northern Goshawk in present study, while the Common Kestrel was infested by *D. rufa* and one of the Western Marsh-Harrier by *C. turbinatum*, *D. fusca*. Thus, *D. rufa* was reported for the first time in Turkey. *Colpocephalum turbinatum* occurs on Rock Pigeon ^[4] and some raptor species ^[1]. However, this species was recorded for the first time on Western Marsh-Harrier in Turkey. But, *C. subzerafae* and *L. tinnunculi* reported from Common kestrel by Ulutaş Esatgil et al. ^[38] could not found in this study.

Studies on the lice species occurring on the Owls (Strigiformes) are very limited in Turkey. Dik and Uslu ^[22] reported that, *Strigiphilus strigis* has been recorded from Eurasian Eagle-Owl (*Bubo bubo*) for the first time in Turkey. *Strigiphilus barbatus* has been firstly reported by Dik ^[31] and then by the other researchers ^[34,36] from Long-eared Owl (*Asio otus*) in Turkey. Two owl species were examined in this study and no louse species was found on the Little Owl (*Athene noctua*), while the four Long-eared Owl specimens were found to be infested with the lice. Three

of them were infested by *S. barbatus* and the other was parasitized with *Columbicola columbae*, although this species is a specific for Rock Pigeon. As far as we know, this is the first record of this host-parasite association. This association can be explained by transferring of louse specimens from Rock Pigeon to Long-eared Owl while hunting. Louse individuals collected from the Owl was alive. Therefore, adaptation of *C. columbae* to owl as a new host could be another alternative explanation for this association.

Lice species in the genera *Bonomiella*, *Hohorstiella*, *Campanulatus*, *Coloceras*, *Colpocephalum*, *Columbicola* and *Physconelloides* occurs on the birds in the order Columbiformes ^[1]. *Hohorstiella lata*, *Colpocephalum turbinatum*, *Campanulatus compar* and *C. columbae* have been recorded from Rock Pigeon in Turkey, previously ^[4,6,8,10,14,36,40]. The results of those studies showed that, the prevalence of *C. columbae* and *C. compar* were higher than the other species ^[4,6,8,10,14]. In a study, *Columbicola bacillus* was reported from Eurasian Collared Dove (*Streptopelia decaocto*) as a new record for this host in Turkey ^[32]. In this study, six bird specimens (4 Rock Pigeons and 2 Eurasian Collared Doves) in the order Columbiformes were examined and, all of the specimens with except of one Eurasian Collared Dove were infested by the lice species. As a result, *Columbicola columbae* and *Coloceras israelensis* on the Rock Pigeon and *Coloceras piageti* on the Eurasian Collared Dove were found. The last two species; *C. israelensis* and *C. piageti* were the first records for Turkey lice fauna. On the other hand, *C. compar*, *H. lata* and *C. bacillus* were not detected in the study.

Piagetiella titan, *Pectinopygus forficulatus* and *Colpocephalum eucarenum* have been recorded on Great White Pelican previously in Turkey ^[15,18,23]. According to results of those reports, *P. titan* and *P. forficulatus* were the most common lice species on Great White Pelican, but; *C. eucarenum* was found rarely on the pelicans. Two Great White Pelican specimens were examined in this study, and both of them were infested by *P. titan* and *P. forficulatus*. *Colpocephalum eucarenum* was detected only one pelican. While *P. titan* and *P. forficulatus* were obtained more numbers, only two male *C. eucarenum* specimens were found on the birds. These findings are similar to the results of the previous studies. Although, Dik ^[15] has reported that *P. titan* can cause erosive stomatitis in a white pelican, erosive stomatitis was not detected on the pelicans infested by *P. titan* in this study.

As far as we know, there is no study about the lice species on Struthioniformes in Turkey. Price et al. ^[1] reported that the genus *Struthiolipeurus* and the species *S. struthionis* in this genus has been occurred on Ostrich (*Struthio camelus*). In the present study, one Ostrich specimen was examined and a lot of lice specimens were collected from this bird, and they were identified as *S. struthionis*, as the first record for Turkey.

There is a little information about the lice species found on the birds belonging to the order Ciconiiformes in Turkey. Dik and Uslu ^[19] have reported that, presence of *Ardeicola ciconiae*, *Ciconophilus quadripustulatus*, *Colpocephalum zebra* and *Neophilopterus incompletus* from White Stork (*Ciconia ciconia*) in Konya. In addition to that, White Stork, Eurasian Bittern (*Botaurus stellaris*) and Great Egret (*Ardea alba*) samples have been examined for the louse, recently ^[31,34]. Although, there was no found any louse species on the Eurasian Bitter, *A. ciconiae*, *C. quadripustulatus*, *C. zebra* and *N. incompletus* were detected on the White Storks. Also, it was stated that, *Comatomenopon elongatum* was recorded as a first record on Great Egret for Turkey in that study ^[34]. In this study, all of the species reported from White Stork by Dik and Uslu ^[19] were detected from the White Stork specimen.

There is no any detailed study on the lice species of the order Psittaciformes in Turkey. Although, some of the bird samples belonging to this order were examined, but any louse specimen was found ^[31]. Dik ^[32] reported that *Afrimenopon waar* from budgerigar (*Melopsittacus undulatus*) as a new record for Turkey. In the current study, eight budgerigars and one cockatiel (*Nymphicus hollandicus*) were examined, but any louse specimen was obtained on all of them, probably they were pet animals and provided adequate living conditions by humans.

Among the bird species belonging to the order Coraciiformes; only European Bee-eater (*Merops apiaster*) has been examined for louse, previously in Turkey. *Meromenopon meropis* and *Meropoecus meropis* have been detected on the birds in those studies ^[35,36]. In this study, only one European Bee-eater was examined and *Meropoecus* sp. and *Brueelia apiastri* were detected on the bird while *Meromenopon meropis* was not found. *B. apiastri* was recorded for the first time from the Eurasian Bee-eater in Turkey in this study. *Meropoecus* specimens could not be identified to species; because all of them were nymph stages.

As a result, a total of 85 bird specimens belonging to the 26 species in the orders Struthioniformes, Pelecaniformes, Ciconiiformes, Coraciiformes, Columbiformes, Psittaciformes, Strigiformes, Falconiformes and Passeriformes were examined in this study. Seventeen (65.38%) of these species were infested with lice species and no louse specimen was found in nine bird species (34.62%). Forty-one of 85 bird specimens (48.23%) were found to be infested with lice specimens and a total of 32 lice species belonging to the suborders Amblycera (11 species) and Ischnocera (21 species) were recorded. The results of the study showed that, the genera *Falcolipeurus*, *Coloceras* and *Struthiolipeurus*, and the species; *Brueelia apiastri*; *Brueelia munda*; *Coloceras piageti*; *Coloceras israelensis*; *Degeeriella nesus*; *Degeeriella phylctopygus*; *Degeriella rufa*; *Falcolipeurus quadripustulatus*; *Laemobothrion vulturis*; *Menacanthus oriole*; *Philopterus fringillae*; *Struthiolipeurus*

struthionis were reported for the first time in Turkey. Also, *C. columbae* was reported from Long-eared Owl, as a first record.

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REFERENCES

1. Price RD, Hellenenthal RA, Palma RL, Johnson KP, Clayton DH: The Chewing Lice: World checklist and biological overview. Illinois Natural History Survey Special Publication, 24. x + 501 pp, 2003.
2. Şekercioğlu ÇH, Anderson S, Akçay E, Bilgin R, Can ÖE, Semiz G, Tavşanoğlu Ç, Yokeş MB, Soyumert A, İpekdağ K, Sağlam İK, Yücel M, Dalfes HN: Turkey's globally important biodiversity in crisis. Biological Conservation, 2011.
3. Mimioğlu MM: Türkiye'de tavuklarda Mallophaga'lar (tavuk bitleri) ve en uygun mücadele metotları üzerine araştırmalar. Ankara Üniv Vet Fak Yayın: 32, Çalışmalar: 16, Doçentlik tezi, Ankara Üniversitesi Basımevi, Ankara, 60 s., 1952.
4. Merdivenci A: Türkiye'nin Entomolojik Coğrafyası. Unat EK, Yaşarol Ş, Merdivenci A (Eds): Türkiye'nin Parazitolojik Coğrafyası. s.114-154. Ege Üniversitesi Tıp Fakültesi Yayınları No: 42, Ege Üniversitesi Basımevi, İzmir, 1965.
5. Güralp N, Mayilmayıl A: Samsun'da sülünlerde (*Phasianus colchicus*) görülen sekal Trichostrongylose ile Mallophaga enfeksiyonlarının etken ve sağaltımları. Ankara Üniv Vet Fak Derg, 18 (2): 271-275, 1971.
6. Tiğin Y: Ehli güvercinlerde (*Columba livia*) bulunan ektoparazitler. Ankara Üniv Vet Fak Derg, 20 (2-3): 372-390, 1973.
7. Dik B, Yaman M, Köse M, Gülbahçe S: Konya'da tavuklarda bulunan Mallophaga türleri. Türkiye Parazitol Derg, 23 (3): 327-330, 1999.
8. Gıcık Y: Ankara ve çevresinde yaban güvercinlerinde ektoparazitler. Kafkas Üniv Vet Fak Derg, 5 (1): 71-74, 1999.
9. Köroğlu E, Şaki CE, Aktaş M, Dumanlı N, Angın M: Elazığ ve yöresinde tavuklarda bulunan bit (Mallophaga) türleri ve bunların yayılışı. Fırat Üniv Sağlık Bil Derg, 13 (2): 57-60, 1999.
10. Köroğlu E, Şimşek S: Elazığ yöresi güvercinlerinde (*Columba livia*) bulunan ektoparazitler ve yayılış oranları. Fırat Üniv Sağlık Bil Dergisi, 15 (1): 195-198, 2001.
11. Okursoy S, Yılmaz F: Bursa yöresinde tavuklarda görülen bit türleri ve bunların dağılımı. Türkiye Parazitol Derg, 26 (1): 71-75, 2002.
12. Aksin N: Elazığ yöresi yabani kekliklerde bulunan Mallophaga türleri. Turk J Vet Anim Sci, 27, 559-565, 2003.
13. Aksin N: Elazığ yöresinde yabani kazlarda bit enfestasyonu. Turk J Vet Anim Sci, 28, 87-90, 2004.
14. Şenlik B, Güleğen E, Akyol V: Bursa yöresindeki evcil güvercinlerin (*Columba livia domestica*) ektoparazitleri. Türkiye Parazitol Derg, 29 (2): 100-102, 2005.
15. Dik B: Erosive stomatitis in a white pelican (*Pelecanus onocrotalus*) caused by *Piagetiella titan* (Mallophaga: Menoponidae). J Vet Med B, 53, 153-154, 2006.
16. Dik B: Mallophaga species on long-legged buzzards (*Buteo rufinus*). New records from Turkey. Türkiye Parazitol Derg, 30 (3): 226-230, 2006.
17. Dik B, Uslu U: Konya'da Halkalı sülünlerde (*Phasianus colchicus*) *Cuculotogaster heterographus* (Mallophaga: Lipeuridae) enfestasyonu. Türkiye Parazitol Derg, 30 (2): 125-127, 2006.
18. Dik B, Uslu U: The first recording of *Piagetiella titan* (Menoponidae: Mallophaga) on a white pelican (*Pelecanus onocrotalus*, Linnaeus) in Turkey.

Türkiye Parazitol Derg, 30 (2): 128-131, 2006.

19. Dik B, Uslu U: Beyaz Leyleklerde (*Ciconia ciconia* Linnaeus, 1758) görülen Mallophaga (Insecta) türleri. *Türkiye Parazitol Derg*, 30 (3): 220-225, 2006.

20. Gülanber A, Kaya Ü, Vaassen EWAM, Yavuz E: Chewing-lice on long-legged Buzzard. *Indian Vet J*, 83 (11): 1238-1239, 2006.

21. Dik B, Aydenizöz-Özkayhan M: Mallophaga species on long-legged buzzards (*Buteo rufinus*) in Turkey. *Türkiye Parazitol Derg*, 31 (4): 298-301, 2007.

22. Dik B, Uslu U: Türkiye'de bir Puhu'da (*Bubo bubo interpositus*) *Strigiphilus strigis* (Mallophaga: Philopteridae). *Türkiye Parazitol Derg*, 31 (1): 69-71, 2007.

23. Dik B, Uslu U: Türkiye'de, Beyaz Pelikanlarda (*Pelecanus onocrotalus*, Linnaeus) görülen Mallophaga türleri. *Türkiye Parazitol Derg*, 32 (1): 71-76, 2008.

24. Dik B, Yamaç E: Türkiye'de bir Kara Akbaba'da (*Aegypius monachus* L.) ilk *Colpocephalum trachelioti* (Amblycera: Menoponidae). *Türkiye Parazitol Derg*, 32 (2): 149-152, 2008.

25. Karataş A, Göçmen B, Karataş A: A new record for Turkish lice fauna: *Dennyus hirundinis* (Linnaeus, 1761) (Mallophaga: Menoponidae). *Türkiye Parazitol Derg*, 32 (1): 77-78, 2008.

26. Dik B: Türkiye'de, Çobanaldatanlarda (*Caprimulgus europaeus* L.)'da ilk *Mulcticola hypoleucus* (Denny, 1842) (Phthiraptera: Ischnocera) olgusu. *Türkiye Parazitol Derg*, 33 (3): 212-214, 2009.

27. Dik B, Uslu U: Konya Hayvanat Bahçesi'ndeki Kanatlı Hayvanlarda Görülen Çiğneyici Bit (Phthiraptera: Amblycera, Ischnocera) Türleri. *Türkiye Parazitol Derg*, 33 (1): 43-49, 2009.

28. Aksin N: Chewing lice and feather mites on wild partridges. *Indian Vet J*, 87, 940-941, 2010.

29. Aksin N: The presence of Chewing-lice (Insecta: Phthiraptera) on Wild Quails. *J Anim Vet Adv*, 9 (9): 1377-1379, 2010.

30. Dik B, Şekercioglu ÇH, Kirpik MA, İnak S, Uslu U: Chewing lice (Phthiraptera) species found on Turkish Shorebirds (Charadriiformes). *Kafkas Univ Vet Fak Derg*, 16 (5): 867-874, 2010.

31. Dik B: Türkiye'deki evcil ve yabani kanatlılarda bulunan Çiğneyici bit (Phthiraptera) türler. *Türkiye Parazitol Derg*, 34 (1): 55-60, 2010.

32. Dik B: New records of chewing lice (Phthiraptera) from some bird species in Turkey. *Türkiye Parazitol Derg*, 34, 168-173, 2010.

33. İnci A, Yıldırım A, Dik B, Düzlü Ö: Current knowledge of Turkey's louse

fauna. *Türkiye Parazitol Derg*, 34, 212-220, 2010.

34. İnci A, Dik B, Kibar M, Yıldırım A, Düzlü Ö: Chewing Lice (Phthiraptera) Species on Wild Birds in Cappadocia Region, Turkey. *Türkiye Parazitol Derg*, 34, 174-178, 2010.

35. Dik B, Şekercioglu ÇH, Kirpik MA: Chewing lice (Phthiraptera) species found on birds along the Aras river, Iğdır, Eastern Turkey. *Kafkas Univ Vet Fak Derg*, 17 (4): 567-573, 2011.

36. Dik B, Yamaç E, Uslu U: Chewing lice (Phthiraptera) found on wild birds in Turkey. *Kafkas Univ Vet Fak Derg*, 17 (5): 787-794, 2011.

37. Dik B, Kirpik MA, Şekercioglu ÇH, Şaşmaz Y: Chewing lice (Phthiraptera) found on songbirds (Passeriformes) in Turkey. *Türkiye Parazitol Derg*, 35 (1): 34-39, 2011.

38. Ulutaş-Esatgil M, Efil İ, Tüzer E: Two chewing lice species, *Colpocephalum subzerafae* and *Laemobothrion tinnunculi*, on a Kestrel (*Falco tinnunculus*) in Istanbul: Two new records from Turkey. *Kafkas Univ Vet Fak Derg*, 18 (Suppl-A): A241-A244, 2012.

39. Aldemir O: Kars ilinde tavuklarda bulunan ektoparazitler. *Türkiye Parazitol Derg*, 28 (3): 154-157, 2004.

40. Gülanber A, Tüzer E, Çetinkaya H: A survey on lice infestations of pigeons in Istanbul, Turkey. *Istanbul Univ Vet Fak Derg*, 28 (1): 231-234, 2002.

41. Dik B, Uslu U, Derinbay-Ekici Ö, Işık N: Türkiye'de, sığırcıklarda (*Sturnus vulgaris*, L.) görülen bit (Phthiraptera; Ischnocera Amblycera) türleri. *Türkiye Parazitol Derg*, 33 (4): 316-320, 2009.

42. Clay T: Revisions of Mallophaga Genera. *Degeeriella* from Falconiformes. *Bull Brit Mus (Nat Hist) Ent*, 7, 123-207, 1958.

43. Clay T, Hopkins GHE: The early literature on Mallophaga. *Bull Brit Mus (Nat Hist) Ent*, 3, 221-266, 1954.

44. Price RD, Beer JR: Species of *Colpocephalum* (Mallophaga: Menoponidae) Parasitic upon the Falconiformes. *Can Entomol*, 95, 731-763, 1963.

45. Tandan BK: Mallophaga from birds of the Indian subregion. Part VI. *Falcolipeurus* Bedford. *Proc R Ent Soc Lond (B)*, 33, 173-180, 1964.

46. Tendeiro J: Estudos Sobreos Goniodideos (Mallophaga, Ischnocera) dos Columbiformes. XIV. Género *Coloceras* Taschenberg, 1882. *Rev Ciênc Vet*, 6, 199-524, 1973.

47. Tendeiro J: Estudos Sobreos Goniodideos (Mallophaga, Ischnocera) dos Columbiformes. XVI. Observações cedicionais sobre o género *Nitzschella* Kéler, 1939, com descrição de cinco species novas. *Rev Ciênc Vet*, 7, 144-236, 1974.

The Effects of Seasonal Variation on the Microbial-N Flow to the Small Intestine and Prediction of Feed Intake in Grazing Karayaka Sheep ^[1]

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Summary

The objectives of the present study were to estimate the microbial-N flow to the small intestine and to predict the digestible organic matter intake (DOMI) in grazing Karayaka sheep based on urinary excretion of purine derivatives (xanthine, hypoxanthine, uric acid and allantoin) by the use of spot urine sampling under field conditions. In the trial, 10 Karayaka sheep from 2 to 3 years of age were used. The animals were grazed in a pasture for ten months and fed with concentrate and vetch plus oat hay for the other two months (January and February) indoors. Highly significant linear and cubic relationships ($P<0.001$) were found among months for purine derivatives index, purine derivatives excretion, purine derivatives absorption, microbial-N and DOMI. Through urine sampling and the determination of levels of excreted urinary PD and the Purine Derivatives:Creatinine ratio (PDC index), microbial-N values were estimated and they indicated that the protein nutrition of the sheep was insufficient. In conclusion, the prediction of protein nutrition of sheep under the field conditions may be possible with the use of spot urine sampling, urinary excreted PD and PDC index. The mean purine derivative levels in spot urine samples from sheep were highest in June, July and October. Protein nutrition of pastured sheep may be affected by weather changes, including rainfall. Spot urine sampling may useful in modeling the feed consumption of pasturing sheep. However, further studies are required under different field conditions with different breeds of sheep to develop spot urine sampling as a model.

Keywords: Karayaka sheep, Spot sampling, Urinary purine derivatives, PDC index, microbial-N, Feed intake

Merada Otlayan Karayaka Koyunlarında Mevsimsel Değişimin İnce Bağırsağa Geçen Mikrobiyal-N Miktarı ve Yem Tüketimine Etkisi

Özet

Merada otlayan Karayaka koyunlarında spot idrar örnekleme ile idrarla atılan purin türevlerinden (ksantin, hipoksantin, ürik asit ve allantoin) ince bağırsağa geçen mikrobiyal-N miktarı ve sindirilebilir organik madde tüketiminin (SOMT) mevsimsel olarak belirlenmesi amaçlanmıştır. Araştırmada 2-3 yaşlarında 10 adet Karayaka koyunu kullanılmıştır. Hayvanlara yem materyali olarak 10 ay mera ve 2 ayda (Ocak, Şubat) kapalı ağıllarda fabrika yemi ve fiğ +yulaf otu ile beslenmiştir. Purin türevleri indeksi, idrarla atılan purin türevleri, absorbe edilen purin türevleri, mikrobiyal-N ve SOMT yönünden aylara göre bir değerlendirme yapıldığında gruplar arasında lineer ve kübik ($P<0.001$) bir ilişki tespit edilmiştir. Mikrobiyal-N değerlerine bakıldığında koyunların protein yönünden yetersiz beslendikleri görülmüştür. Sonuç olarak spot idrar toplanarak denemenin yapıldığı benzer saha koşullarında otlatılan Karayaka koyunlarda protein beslenmesini tahmin etmekte idrarla atılan purin türevleri ve Purin Türevleri: Kreatinin (PDC indeksi) kullanılabilir. Ortalama PT miktarı Karayaka koyunları için spot idrar toplamada en yüksek değerler Haziran, Temmuz ve Ekim aylarında belirlenmiştir. Bu aylardaki iklimsel değişikliklerden (yağış fazlalığı) dolayı meradaki koyunların protein beslenmesi etkilenmiştir. Ayrıca spot idrar toplama tekniği meradaki koyunların yem tüketimini belirlemede bir model olabilir. Bunun içinde farklı saha koşullarında farklı koyun ırklarıyla benzer çalışmaların yapılmasına ihtiyaç vardır.

Anahtar sözcükler: Karayaka koyunu, Spot örnekleme, İdrar purin türevleri, PDC indeksi, Mikrobiyal-N, Yem tüketimi



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INTRODUCTION

The Karayaka sheep is one of the indigenous breeds reared on the coastline of the Black Sea region of Turkey. Karayaka sheep are well adapted to the wet climate of the region and there is a total population of approximately one million ^[1].

The use of spot urine sampling has been proposed to predict protein nutrition and digestible organic matter intake in grazing sheep ^[2] and goats ^[3]. Microbial protein flow to the small intestine has been estimated total of purine derivatives excreted in the urine of ruminants ^[4,5]. Rumen microbes constitute the major source of protein supply to the ruminants. The purines from the rumen microbes are metabolized and excreted in the urine as their derivatives, hypoxanthine, xanthine, uric acid and allantoin. In sheep, hypoxanthine and xanthine are converted to uric acid by xanthine oxidase, and uric acid is further converted to allantoin by uricase. All four compounds are excreted in the urine of sheep. The synthesis of microbial protein is dependent on ruminal ammonia nitrogen supply ^[6] and digestible organic matter intake ^[7,8]. Reports that estimate microbial-N flow to the small intestine are mostly from European sheep breeds ^[2,8-11].

The objectives of the present study were to examine the microbial-N flow to the small intestine and to predict DOMI in grazing Karayaka sheep on the basis of urinary excretion of PD by the use of spot urine sampling under field conditions and to investigate the effects of seasonal variation on ruminal microbial synthesis.

MATERIAL and METHODS

The study was approved by the Local Ethics Committee on Animal Experiments of Ondokuz Mayıs University (OMU, 30.12.2009, HADYEK/139).

Animal and Feed Materials

A total of 10 Karayaka sheep aged between 2 and 3 years, and with live weight ranging from 42.4 to 47.6 kg, were used in the this study that was undertaken in Akyazı village of Bafra town, Samsun province, Turkey. The animals were grazed in a pasture for ten months and fed a concentrate and vetch plus oat hay for the other two months (January and February) indoors.

Plant samples were taken by hand clipping to ground level at the beginning of the grazing experiment and repeated every month. They were collected from one square meter area in six different locations in a 0.5-1 ha area ^[12]. The important part of meadow and pasture was formed by *Agropyron cristatum*, *Lotus corniculatus* L., *Agropyron elongatum*, *Bromus inermis*, *Convolvulus* sp., *Trifolium pretense*, *Trifolium repense* and *Dactylis glomerata*. All plant

samples were dried to a constant weight in a forced-air oven at 65°C for 48 h. Ash content was determined by heating in a muffle furnace at 550°C ^[13]. Organic matter (OM) was calculated as DM-ash. Nitrogen (N) content was analyzed with the Kjeldahl method, with the use of a semi-automated N analyzer. Neutral detergent fibre (NDF) and acid detergent fibre (ADF) were determined according to the methodology of VAN SOEST *et al.* ^[14].

The levels of purine derivatives were determined according to the methodology of CHEN *et al.* ^[15] using a spectrophotometer (Shimadzu UV-1700). Allantoin is first hydrolyzed under weak alkaline conditions and at 100°C to allantoinic acid which is further hydrolysed to urea and glyoxylic acid in a weak acid solution. The glyoxylic acid is reacted with phenylhydrazine hydrochloride to produce a phenylhydrazone derivative of the acid. The product forms an unstable chromophore with potassium ferricyanide. The colour was read at 522 nm. Xanthine and hypoxanthine are converted to uric acid by treatment with xanthine oxidase and are thus determined as uric acid, the amount of which is determined by its absorbance at 293 nm, although other compounds may also absorb at this wavelength. When samples are treated with uricase, uric acid is converted to allantoin and other compounds that do not absorb UV at 293 nm. Therefore, the reduction in absorbance reading after treatment with uricase is correlated with the concentration of uric acid in the sample. After treatment, the absorbance of the standards should be zero, if the conversion is complete. Creatinine reacts with picrate ion formed in alkaline medium and a red-orange colour develops. The colour produced from the sample is then compared in a colorimeter at 505 nm with that produced by a known amount of creatinine under the same conditions ^[5].

Mean monthly rainfall and air temperature (°C) in Bafra in 2010 and 2011 were collated by the Samsun Meteorological Office and are presented in Fig. 1.

Urinary Sample Collection

Spot urinary samples were collected from each sheep between 10:00 am and 12:00 pm (2 h after grazing) on two consecutive days per month for a year. Urine samples were collected by closing the mouth and nose of the sheep by hand. Samples were usually collected over a 10 to 35 sec period. After collection, a 10 ml sub-sample was taken, acidified with 1 ml of 10% H₂SO₄ and diluted to 1:4 with distilled water. The samples were stored at -20°C until analysis for purine derivatives.

Estimation of Microbial-N Supply

The microbial-N supply was estimated using the following equation for sheep ^[5].

$$Y = 0.84X + (0.150 W^{0.75} e^{-0.25X})$$

Where X (mmol/d) is absorption of purines (X mmol/d)

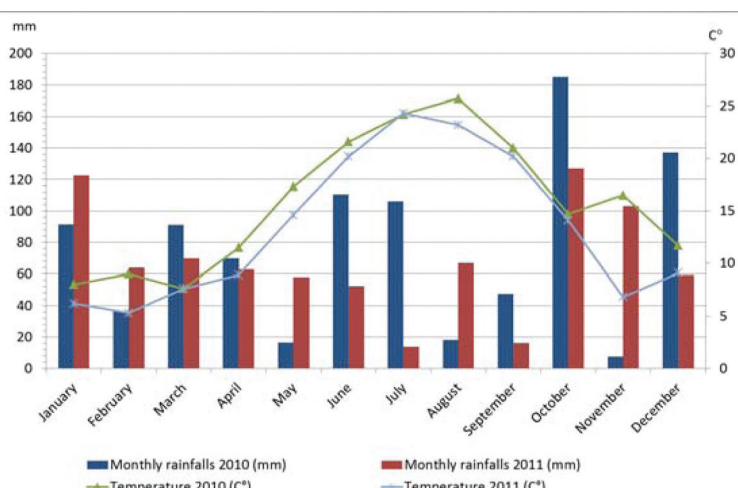


Fig 1. Mean monthly rainfall and air temperature (°C) in Bafrı in 2010 and 2011

Şekil 1. Bafrı'da 2010 ve 2011 yıllarında ortalama aylık yağış ve sıcaklık miktarları (°C)

and Y (mmol/d) is PD excretion in the urine. The calculation of X based on the above non-linear equation can be performed by means of the Newton Raphson iteration process, as shown below:

$$X_{(n+1)} = X_n - [f(X_n) / f'(X_n)]; \text{ where } f(X) = 0.84 X + 0.150 W^{0.75} e^{-0.25X} - Y, \text{ and the derivative of } f(X): f'(X) = 0.84 - 0.038 W^{0.75} e^{-0.25X}$$

An initial value of $X_1 = Y / 0.84$ is fed into the above equation to calculate X_2 , and after the process is repeated for 4 iterations, X_5 should have reached a constant value.

The supply of microbial-N was estimated below from the relationship derived by CHEN and GOMES [11]. The assumptions used in the above equations are: digestibility of microbial purines is 0.83. the N content of purines is 70 mg N/mmol and the ratio of purine-N: total-N in mixed rumen microbes is 11.6:100 [5]. Therefore, the equation to determine microbial-N is: Microbial N (gN/d) = $[X \text{ (mmol/d)} \times 70] / [0.116 \times 0.83 \times 1000] = 0.727X$

Microbial-N estimated from purine derivatives excretion corresponds to the quantity of microbial biomass reaching the duodenum, rather than that synthesized within the rumen. It is expressed as grams of microbial-N per kilogram of digestible organic matter apparently digested in the rumen. The model assumes that the digestible organic matter in the rumen is 65% of digestible organic matter intake [16].

Estimation of PDC Index

PDC index is determined by calculating the creatinine concentrations and total purine derivatives in the urine. The following equation is used to index the PDC:

$$\text{PDC index} = (PD/C) \times W^{0.75}$$

where W is the body weight (kg), and PD and C are purine derivatives and creatinine concentrations, respectively in mmol/L.

PD excretion (mmol/d) = PDC index $\times C$ where C is the daily creatinine excretion (mmol/kg $W^{0.75}$) for a specific

breed of animals, which should have been previously measured from the complete urine collection. Average daily creatinine excretion was taken to be 503 $\mu\text{mol/kg CA}^{0.75}$ for sheep [5].

Estimation of Digestible Organic Matter Intake (DOMI)

The spot measurement of the PDC index can provide an estimate of feed intake. Digestible organic matter intake was calculated with the following equation [5].

$$\text{DOMI (g/d)} = 59.7 \times \text{PDC} - 678$$

Statistical Analysis

Data were summarized with descriptive statistics for means, and the standard errors of the means were analyzed with analysis of variance (ANOVA), using the Least Square Method of the GLM procedure of the SAS [17]. The differences between the groups were analyzed via 3rd order orthogonal polynomials. All results were summarized as mean \pm standard error of mean (SEM). Ordinary linear regression and Pearson correlation analyses were performed with the use of variables, namely allantoin, microbial-N, DOMI and purine derivatives excretion, and digestible organic matter digested in the rumen.

RESULTS

The dry matter (DM), organic matter (OM), crude protein (CD), neutral detergent fiber (NDF) and acid detergent fiber (ADF) values of meadow and pasture samples (g/kg DM) collected monthly are presented in Table 1. Levels of creatinine and allantoin, uric acid, hypoxanthine and xanthine in spot urine samples are shown in Table 2. The purine derivatives index (PDC index), purine derivatives excretion, purine derivatives absorption, microbial-N and DOMI for spot urine samples are shown in Table 3. Monthly variations in both mean values of the PDC index and microbial-N (Fig. 2) and mean values of DOMI are presented in Fig. 3.

Table 1. The nutrient composition of meadow and pasture samples collected monthly (g/kg DM)**Tablo 1.** Aylık olarak toplanan çayır ve mera örneklerinin besin madde kompozisyonu (g/kg KM)

Period			DM	OM	CP	NDF	ADF
Years	Months						
2010	June		913.0	838.4	134.0	437.2	351.7
	July		931.2	866.5	148.0	476.8	337.7
	August		892.5	801.2	106.5	500.2	392.1
	September		909.3	830.9	143.7	479.4	334.9
	October		905.6	824.5	131.1	442.1	313.5
	November		913.0	837.6	107.5	468.4	345.5
	December		904.3	827.3	101.2	543.7	415.5
2011	January	1	902.1	829.4	118.0	613.0	469.6
	February	2	896.0	832.7	138.5	232.7	113.7
	March		917.5	851.3	127.5	410.8	264.0
	April		923.4	819.4	154.0	403.4	246.8
	May		906.8	840.3	135.7	451.6	344.0

1- Mixed vetch and oat hay were used to feed sheep in January and February, 2- Concentrate was used to feed sheep in January and February

Table 2. Mean concentrates (mmol /L) of allantoin, uric acid, hypoxanthine plus xanthine and creatinine in spot urine samples collected from grazing Karayaka sheep**Tablo 2.** Merada otlayan Karayaka koyunlarından toplanan spot idrar örneklerinde allantoin, ürik asit, hipoksantin + ksantin ve kreatinin ortalama değerleri (mmol/L)

Grazing Period		n	Allantoin	Uric Acid	Hypoxanthine +Xanthine	Creatinine	Purine Derivatives
Year	Months						
2010	June	9	7.67±0.37	1.76±0.18	0.65±0.06	6.39±0.19	10.08±0.44
	July	9	7.71±0.38	1.40±0.08	0.68±0.06	6.30±0.32	9.79±0.43
	August	10	6.17±0.18	1.25±0.06	0.61±0.06	6.75±0.23	8.02±0.18
	September	10	6.81±0.29	1.77±0.09	0.75±0.07	5.74±0.21	9.32±0.29
	October	10	8.62±0.58	1.59±0.12	0.68±0.04	6.49±0.28	10.90±0.66
	November	9	7.28±0.27	1.64±0.19	0.35±0.03	4.72±0.22	9.27±0.31
	December	10	5.04±0.22	1.96±0.10	0.32±0.02	5.44±0.27	7.32±0.21
2011	January	9	4.84±0.18	0.87±0.05	0.42±0.04	5.00±0.16	6.13±0.20
	February	8	6.31±0.31	1.29±0.13	0.53±0.03	6.64±0.28	8.12±0.31
	March	10	5.64±0.28	0.59±0.04	0.50±0.05	5.60±0.27	6.73±0.30
	April	10	5.12±0.20	1.29±0.09	0.54±0.06	5.61±0.15	6.95±0.24
	May	9	5.55±0.33	1.28±0.12	0.48±0.05	5.46±0.20	7.31±0.35
Significance of main effects							
Linear			***	***	NS	NS	***
Quadratic			***	NS	***	***	***
Cubic			***	NS	***	NS	***

*** $P < 0.001$, NS: Non Significant, L: Linear, Q: Quadratic, C: Cubic

DISCUSSION

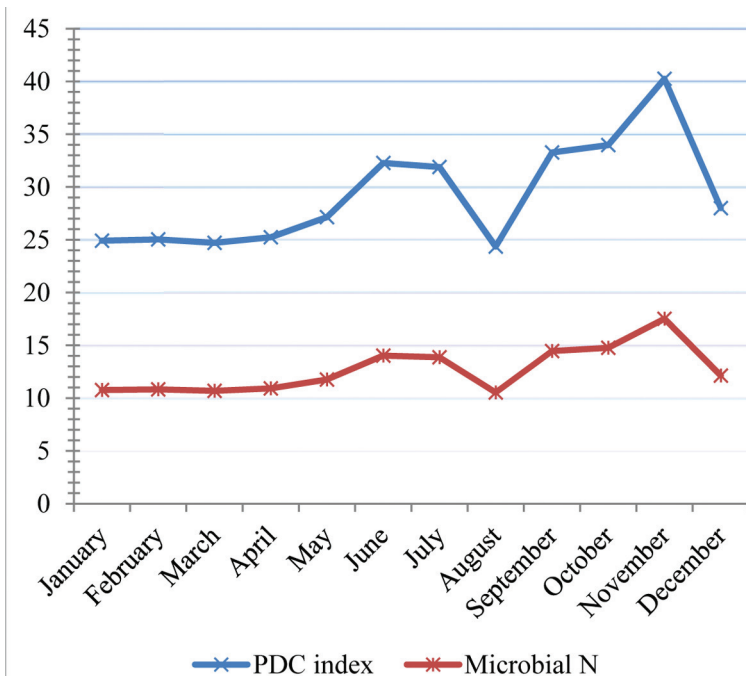
Measurement of microbial protein supply to sheep has been a major area of study in the context of their protein nutrition. An estimate of microbial protein contribution to the intestinal protein flow is incorporated into the new protein evaluation systems already being used in a number

of countries. The supply of microbial protein to the animal per unit of feed ingested varied from 14 to 60 g microbial-N/kg digestible organic matter fermented in the rumen ^[16]. This variation is due to the influence of various factors related to the diet or rumen environment. As seen in *Table 1*, CP levels of plant samples collected monthly did not fluctuate all year around. The lowest CP levels were

Table 3. Mean levels of purine derivatives index (PDC index), purine derivatives excretion, purine derivatives absorption, microbial-N supply and DOMI in spot urine samples collected from grazing Karayaka sheep**Tablo 3.** Merada otlayan Karayaka koyunlarından toplanan spot idrar örneklerinde PDC indeksi, pürin türevleri ekskresyonu, pürin türevleri absorpsiyonu, mikrobal-N ve DOMI ortalama değerleri

Grazing Period		n	PDC Index	PD Excretion (mmol/d)	Purine Absorption (mmol/d)	Microbial-N Supply		DOMI g/d
Year	Months					(g of N/d)	(g of N/kg of DOMR)	
2010	June	9	32.29±1.51	16.24±0.76	19.28±0.92	14.02±0.67	20.87±0.73	1249.44±90.31
	July	9	31.90±1.02	16.04±0.52	19.06±0.62	13.86±0.45	20.86±0.78	1226.26±61.17
	August	10	24.36±0.74	12.25±0.37	14.48±0.45	10.53±0.33	21.06±0.70	776.13±44.04
	September	10	33.27±1.17	16.74±0.59	19.89±0.70	14.46±0.51	20.98±0.70	1308.28±69.59
	October	10	33.98±1.40	17.09±0.70	20.31±0.85	14.77±0.62	19.76±0.73	1350.41±83.61
	November	9	40.26±1.20	20.25±0.60	24.10±0.72	17.52±0.52	18.12±0.73	1725.41±71.46
	December	10	28.00±1.14	14.08±0.57	16.69±0.69	12.14±0.50	17.49±0.73	993.41±67.84
2011	January	9	24.90±0.69	12.53±0.35	14.81±0.43	10.77±0.31	21.24±0.70	808.77±41.49
	February	8	25.04±0.99	12.60±0.50	14.89±0.61	10.83±0.44	17.26±0.70	816.91±59.32
	March	10	24.71±0.92	12.43±0.46	14.69±0.56	10.68±0.41	17.29±0.70	797.47±54.68
	April	10	25.23±0.94	12.69±0.47	15.01±0.57	10.91±0.42	15.69±0.73	828.42±56.06
	May	9	27.14±0.96	13.65±0.48	16.18±0.59	11.76±0.43	19.34±0.70	942.39±57.47
Significance of main effects								
Linear			***	***	***	***	***	***
Quadratic			NS	NS	NS	NS	NS	NS
Cubic			***	***	***	***	*	*

*** $P < 0.001$, * $P < 0.05$, NS: Non Significant, DOMR: Digestible organic matter fermented in the rumen, calculated as $0.65 \times \text{DOMI}$ (g of N/kg of DOMR), DOMI: Digestible organic matter intake g/d

**Fig 2.** Monthly changes of mean values of PDC index and Microbial-N (g N/d) for grazing Karayaka sheep**Şekil 2.** Merada otlayan Karayaka koyunlarında ortalama PDC index ve Mikrobal N değerlerinin aylık değişimi

observed in August and December. They are in agreement with estimated mean microbial-N and DOMI values (Fig. 2 and 3) for Karayaka sheep.

Creatinine is produced from creatine in muscle, and is excreted in the urine. Its excretion is correlated with the muscle mass. When expressed as 'mmol/per kg $W^{0.75}$,

the daily excretion is relatively constant. The value is approximately 0.5 mmol/kg $W^{0.75}$ /d in sheep [5]. The excretion of creatinine is affected minimally by the amount of protein and non-protein nitrogen consumed. In the current study, creatinine concentrations in the spot urine samples were higher in August, October and February than in other months but overall values were not significantly

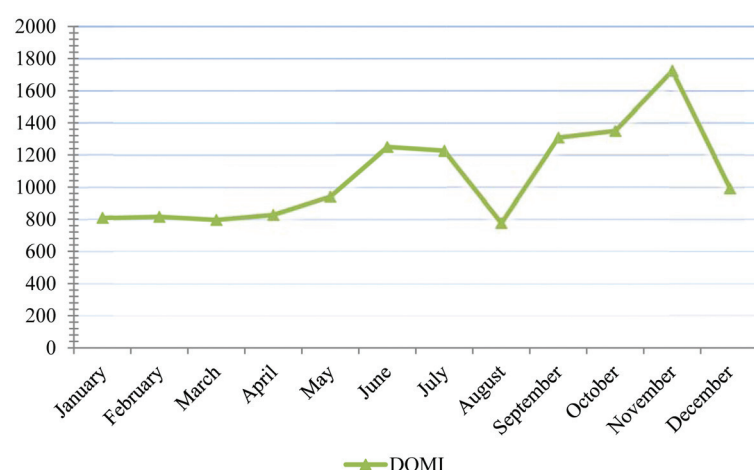


Fig 3. Monthly changes of mean values of DOMI (g/d) for grazing Karayaka sheep

Şekil 3. Merada otlayan Karayaka koyunlarında ortalama DOMI (g/gün) değerlerinin aylık değişimi

influenced by season (Table 2). There was no linear or cubic relationship between creatinine excretion and month but a quadratic relationship was determined. It was also the case that season did not affect creatinine excretion in urine.

The method for estimating microbial-N production from urinary purine derivatives assumes that duodenal nucleic acids are mostly of microbial origin [15]. It has been reported that after intestinal digestion and absorption, the purine base catabolites are proportionally recovered in urine, mostly as allantoin, but also as hypoxanthine, xanthine and uric acid [18]. The present study reports 69-84%, 9-27% and 4-8%, for allantoin, uric acid and xanthine plus hypoxanthine, respectively. For the same catabolites, Chen and Gomes [11] reported the proportions to be 60-80%, 10-30% and 5-10% for allantoin, uric acid and xanthine plus hypoxanthine, respectively. Furthermore, allantoin excretion in urine was reported to be 80-85% of total purine derivatives. The profile of PD excretion in grazing Karayaka sheep was similar to that reported in previous studies [19-21]. The results obtained may indicate that the proportion of purine derivatives is independent of diet.

In the current study, the levels of allantoin in the urine of grazing sheep ranged from 4.84-8.62 mmol/L on a monthly basis across the sampling period. The study also determined a positive correlation ($r=0.615$, $P<0.01$) between the amount of allantoin excreted in urine and rumen microbial protein flowing into the small intestine, as described by the equation:

$$Y=0.378X+1.594$$

Where y is the amount of allantoin excreted in urine and x is rumen microbial protein flowing into the small intestine. This equation showed that allantoin excretion was able to enhance duodenal flow of microbial protein. Studies of cattle [22,23] and sheep [7] have indicated a high correlation between the excretion of purine derivatives and rumen microbial protein flow into the small intestine ($R^2 = 0.97$). In the present study, the average allantoin amounts in spot urine samples for sheep were lowest in January,

April and December, whereas the highest values were determined in June, July and October. In 2010, allantoin amounts in June, July and October were higher than in other months (Table 2). That phenomenon may reflect the impact of vegetation changes due to higher than average rainfall during those months.

The higher excretion of PD in October and November clearly indicates enhanced microbial protein synthesis, since significant relationships have already been reported between urinary PD excretion and the levels of nucleic acid infused in the abomasum [4,15] and duodenum [9,24]. Orellana-Boero *et al.* [25] found that the excretion of purine derivatives in the urine increased linearly ($r = 0.867$) with digestible organic matter intake. The principle is that duodenal purine bases are efficiently absorbed in the small intestine [24,26]. Urinary PD excretion is used to predict ruminal microbial protein synthesis. The daily excretion of purine derivatives and the microbial-N supply in grazing Karayaka sheep were found to be in the range of 12.25-20.25 and 10.53-17.52 mmol/d, respectively (Table 3). These values for Karayaka sheep are within the range of those published for different sheep breeds [2,10]. The urinary PD excretion values obtained in goats [3] and wethers are similar to those observed in sheep [10,27,28]. Hence, purine derivatives in spot urine samples may provide a practical indicator of microbial protein supply status in grazing ruminants.

The estimated average monthly DOMI values (Table 3) in grazing Karayaka sheep were within the range of 776 to 1725 g/day. The estimation of DOMI from PDC index (Table 3) through the use of spot urine sampling from grazing Karayaka sheep showed that this technique may be applied in grazing animals where DOMI cannot be measured directly. Many reports have confirmed a linear relationship between allantoin excretion and both the level of feed intake and flow of nucleic acids in the duodenum [29]. Laurent *et al.* [30] determined that allantoin excretion is correlated with digestible organic matter intake, and also that allantoin excretion ($r=0.54$, $P<0.01$) can be used as an index of rumen microbial protein synthesis.

The present study also determined that digestible organic matter fermented in the rumen was converted to a similar proportion of microbial-N in all months, ranging from 16 to 21 g N/kg of digestible organic matter apparently digested in the rumen (Fig. 2 and 3; Table 3), which was similar the range reported by Yu *et al.*^[31]. The higher amount of urinary allantoin reported in the present study in June, July, September, October and November was due to the increased digestible organic matter intake. The present study also determined that there were linear and cubic relationship between the PDC index, urinary excretion of purine derivatives, microbial-N and DOMI and month (Table 3), and that there was a seasonal influence on these parameters.

In the current study, the estimated microbial-N values appear insufficient for adequate protein nutrition. Monthly mean values of PDC index, microbial-N and DOMI were relatively stable. However, fluctuations were observed between June and December (Fig. 2 and 3). The lowest values for DOMI, PDC index and microbial-N were observed in August and December (Fig. 2 and 3). This is due to meadow and pasture conditions reflecting the driest period of the year (Fig. 1). In conclusion, protein nutrition of pastured sheep may be affected by weather changes. Spot urine sampling may serve as the basis for modeling their feed requirements. Furthermore, it may provide a basis for the preparation of balanced diets to meet the protein requirements of sheep by closely approximating the amount of rumen microbial-N flowing into the small intestine. However, further studies are required under different field conditions and with different breeds of sheep to develop the spot urine sampling technique into a model.

REFERENCES

- 1. Turkish Statistical Institute (TurkStat):** Livestock Statistics, Url:<http://www.turkstat.gov.tr>, Accessed: 24.05.2008.
- 2. Chen XB, Mejia AT, Kyle DJ, Ørskov ER:** Evaluation of the use of the purine derivative: creatinine ratio in spot urine and plasma samples as an index of microbial protein supply in ruminants: Studies in sheep. *J Agr Sci*, 125, 137-143, 1995.
- 3. Cetinkaya N, Salman M, Genc B:** Estimation of the microbial N flow to small intestine in Saanen goats and kids based on urinary excretion of purine derivatives by the use of spot urine sampling technique. *Kafkas Univ Vet Fak Derg*, 16 (1): 75-79, 2010.
- 4. Fujihara T, Ørskov ER, Reeds PJ, Kyle DJ:** The effect of protein infusion on urinary excretion of purine derivatives in ruminants nourished by intragastric infusion. *J Agric Sci Camb*, 109, 7-12, 1987.
- 5. IAEA:** A training package the technique for estimating microbial protein supply in ruminants based on determination on purine derivatives in urine. Joint FAO/IAEA Division of Nuclear Applications in Food and Agriculture. Vienna, Austria, 2003.
- 6. Wanapat M, Pimpa O:** Effect of ruminal NH₃-N levels on ruminal fermentation, purine derivatives, digestibility and rice straw intake in swamp buffaloes. *J Anim Sci*, 12, 904-907, 1999.
- 7. Puchala R, Kulasek GW:** Estimation of microbial protein flow from the rumen of sheep using microbial protein flow from the rumen of sheep using microbial nucleic acid and urinary excretion of purine derivatives. *Can J Anim Sci*, 72, 821-830, 1992.
- 8. Makkar HPS, Chen XB:** Estimation of microbial protein supply in ruminants using urinary purine derivatives. Springer. 1st ed., p.212, 2004.
- 9. Antoniewicz AN, Hienemann WW, Hanks EM:** The effect of changes in the flow of nucleic acids on allantoin excretion in the urine of sheep. *J Agric Sci*, 95, 395-400, 1980.
- 10. Chen XB, Chen YK, Franklin MF, Ørskov ER, Shand WJ:** The effect of feed intake and body weight on purine derivative excretion and microbial protein supply. *J Anim Sci*, 70, 1534-1542, 1992.
- 11. Chen XB, Gomes JM:** Estimation of Microbial Protein Supply to Sheep and Cattle Based on Urinary Excretion of Purine Derivatives and Overview of the Technical Detail. International Feed Resources Unit. Rowett Research Institute, Bucksburn Aberdeen, UK. Occasional Publication, pp.1-21, 1992.
- 12. Karabolut A, Canbolat Ö:** Feed Evaluation and Analyze Methods. Uludag University Publ., no: 2,05.048,0424, Bursa, 2005.
- 13. AOAC:** Official Methods of Analysis. 15th ed., Association of Official Agricultural Chemists, Washington, DC, 1995.
- 14. Van Soest PJ, Robertson JB, Lewis BA:** Methods for dietary fiber, neutral detergent fiber, and nonstarch polysaccharides in relation to animal nutrition. *J Dairy Sci*, 74, 3583-3597, 1991.
- 15. Chen XB, Hovell FD, De B, Ørskov ER, Brown DA:** Excretion of purine derivatives by ruminants: Effect of exogenous nucleic acid supply on purine derivative excretion by sheep. *Br J Nutr*, 63, 131-142, 1990.
- 16. Agricultural Research Council (ARC):** The nutrient requirements of ruminant livestock. Suppl. No. 1, Commonwealth Agricultural Bureaux, Slough, 1984.
- 17. SAS Statistical Software:** SAS Compus drive, Carry, NC 27513 USA, 2009.
- 18. McAllan AB, Smith RH:** Degradation of nucleic acids in the rumen. *Br J Nutr*, 29, 331-345, 1973.
- 19. Brun-Bellut J, Linberg JE, Hadjipanayiotou M:** Protein nutrition and requirements of adult dairy goats. In: Morand-Fehr P (Ed): Goat Nutrition. pp.82-93, EAAP Publ. No. 46, Pudoc Wageningen Netherlands, 1991.
- 20. Linberg JE:** Urinary allantoin excretion and digestible organic matter intake in dairy goats. *Swed J Agric Res*, 15, 31-37, 1985.
- 21. Ma T, Deng K, Jiang C, Tu Y, Zhang N, Liu J, Zhao Y, Diaao Q:** The relationship between microbial N synthesis and urinary excretion of purine derivatives in Dorperxthin-tailed Han crossbred sheep, <http://dx.doi.org/10.1016/j.smallrumres.2012.09.003>, Accessed: 20.09.2012.
- 22. Cetinkaya N, Yaman S, Baber NHO:** The use of purine derivatives/ creatinine ratio in spot urine samples as an index of microbial protein supply in Yerli Kara crossbred cattle. *Livest Sci*, 100, 91-98, 2006.
- 23. Gonzalez-Ronquillo M, Balcells J, Belenguer A, Castrillo C, Mota M:** A comparison of purine derivatives excretion with conventional methods as indices of microbial yield in dairy cows. *J Dairy Sci*, 87, 2211-2221, 2004.
- 24. Balcells J, Guada JA, Peiro JM:** Simultaneous determination of allantoin and oxypurines in biological fluids by high performance liquid chromatography. *J Chromatogr*, 575, 153-157, 1992.
- 25. Orellana-Boeroa P, Seradib AR, Fondevilac MJ, Noland J, Balcells J:** Modelling urinary purine derivatives excretion as a tool to estimate microbial rumen outflow in alpacas (*Vicugna pacos*). *Small Rumin. Res.*, 107, 101-104, 2012.
- 26. Belenguer A, Yanez D, Balcells J, Ozdemir BNH, Gonzalez-Ronquillo M:** Urinary excretion of purine derivatives and prediction of rumen microbial outflow in goats. *Livest Prod Sci*, 77, 127-135, 2002.
- 27. Carro MD, Cantalapiedra-Hijar G, Ranilla MJ, Molina-Alcaide E:** Urinary excretion of purine derivatives, microbial protein synthesis, nitrogen use, and ruminal fermentation in sheep and goats fed diets of different quality. *J Anim Sci*, 90, 3963-3972, 2012.
- 28. Hindrichsen IK, Osuji PO, Odenyo AA, Madsen J, Hvelplund T:** Effects of supplementation of a basal diet of maize straw with different amounts of *Leucaena diversifolia* on intake, digestibility, nitrogen metabolism and rumen parameters in sheep. *Anim Feed Sci Technol*, 98, 131-142, 2002.

29. Kreuzer M, Kirchgeßner M, Kellner RJ, Roth FX: Effect of varying protein and energy concentration on digestibility of nutrients, nitrogen metabolism and allantoin excretion in wethers. *German J Anim Physiol Anim Nutr* 55, 144-159, 1986.

30. Laurent F, Vignon B: Factors of variations in urinary excretion of allantoin in sheep and goats. *Anim Sci*, 64, 281-282, 1983.

31. Yu P, Boon-ek L, Leury BJ: Effect of dietary protein variation in term of net truly digest intestinal protein (DVE) and rumen degraded protein balance (OEB) on the concentration and excretion of urinary creatinine, purine derivatives and microbial N supply in sheep: Comparison with the prediction from the DVE/OEB model. *Anim Feed Sci Tech*, 93, 71-91, 2001.

Effects of Graded Levels of Crude Glycerine Addition to Diets on Growth Performance, Carcass Traits and Economic Efficiency in Broiler Chickens ^{[1][2]}

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Summary

The aim of the current study was to evaluate the effects of graded levels of crude glycerine addition to diets on growth and economic performance, carcass yield, organ weights and moisture levels of meat in broiler chickens. For this purpose, a total of 270 one day old male chicks (Ross 308) were randomly divided into 3 equal groups which fed with isocaloric and isonitrogenous diets with graded levels (0%, 5% and 10%, respectively) of crude glycerine. Each of experiment group was constituted by 5 subgroups with 18 birds each. The birds had ad libitum access to feed and water until termination of the experiment at d 42. Results indicated that 5% dietary crude glycerine addition improved body weight, body weight gain and feed conversion ratio compare with birds fed with basal diet at d 42 ($P<0.01$). Similarly, dietary 10% crude glycerine supplementation also increased growth performance and improved FCR in broiler chickens at 28th and 42nd days of experiment ($P<0.05$ and $P<0.001$, respectively). Moreover, birds fed with 5% dietary crude glycerine had showed over 20% higher relative economic efficiency compare than birds fed basal diet. As a conclusion, dietary crude glycerine addition improved growth and economic performance and Feed Conversion Ratio of broilers in both supplementation levels.

Keywords: Broiler, Carcass, Economic efficiency, Glycerine, Growth performance

Rasyona Artan Düzeylerde Ham Gliserin İlavesinin Broylerlerde Büyüme Performansına, Karkas Özelliklerine ve Ekonomik Etkinliğe Olan Etkileri

Özet

Bu araştırmanın amacı, rasyona artan düzeyde ham gliserin ilavesinin broylerlerde büyüme ve ekonomik performans, karkas randımanı, iç organ ağırlıkları ve etin nem düzeyi üzerine etkilerinin belirlenmesidir. Bu amaçla toplam 270 adet bir günlük yaşta erkek broyler civciv (Ross 308) her biri izokalorik ve izonitrojenik olan ve artan düzeylerde (sırasıyla %0, %5 ve %10) ham gliserin içeren yemlerle beslenen üç eş gruba ayrılmıştır. Her bir deneme grubu her biri kendi içinde 18 adet civciv içeren 5'er alt gruba ayrılmıştır. Hayvanlar denemenin sonlandığı 42. güne kadar yem ve suya ad libitum olarak ulaşmışlardır. Deneme sonunda %5 düzeyinde ham gliserin ilavesinin canlı ağırlık, canlı ağırlık artışı ve yemden yararlanma oranlarını bazal rasyona beslenen hayvanlara göre önemli ölçüde iyileştirdiği belirlenmiştir ($P<0.01$). Benzer olarak rasyona %10 düzeyinde ham gliserin ilavesi denemenin 28. ve 42. günlerinde büyüme performansını arttırmış ve yemden yararlanma oranını iyileştirmiştir (sırasıyla, $P<0.05$ ve $P<0.001$). Buna ilaveten, %5 ham gliserin ile beslenen hayvanlar bazal rasyona beslenenlere göre %20 daha yüksek göreceli ekonomik etkinlik göstermiştir. Sonuç olarak rasyona ham gliserin ilavesi, her iki düzeyde de, broylerlerde büyüme ve ekonomik etkinlik ile Yemden Yararlanma Oranı'nı iyileştirmiştir.

Anahtar sözcükler: Broyler, Büyüme performansı, Ekonomik etkinlik, Gliserin, Karkas



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INTRODUCTION

In poultry industry, because of increasing prices of energy rich feedstuffs, there is remarkable effort for searching of energy alternatives for chicken diets. In this content, researchers focus on the use glycerine as a less expensive energy source in poultry diets ^[1]. Glycerine, also known as glycerol or glycerin, is a by-product of biodiesel production and usage of this product as an energy source for animal diets has been got attention in recent years ^[2-4]. Glycerine has been evaluated for poultry as a feed ingredient which provides energy for cellular metabolism ^[5-9]. Moreover, several studies have been performed for understanding effects of dietary glycerine addition on performance in broiler chickens ^[4,7-9], quails ^[10], and laying hens ^[11]. Simon et al.^[7] observed that dietary glycerol addition at level of 5 and 10% had improving effect on body weight gain, feed intake and feed efficiency similar to Suchy et al.^[12] who evaluated that effect of pure and raw glycerol addition to diet on production parameters and slaughter traits in male and female broiler chickens. They found that birds fed diet with glycerol had significantly higher body weight, feed intake and carcass trait compare with birds fed with basal diet. On the other hand, Cerrate et al.^[9] reported that feed intake and body weights of birds were decreased with 10% glycerine addition to diets in broilers. However authors stressed that addition of glycerine to diets at a level of 5 and 2.5% had improvement effect on breast yield as a percentage and dressed carcass of birds.

Even though pure glycerine can be used in many different applications including food, cosmetic, drug and weapon industries, and generally recognized as safe for use in animal feed usage of crude glycerine, which find a place to itself in animal feeds recently, has some concern about residual level of methanol sodium, fatty acid and moisture content in it ^[3,9,11]. In this point, because of contradiction results from different studies about the effect of dietary glycerine addition on performance and carcass characteristics in broiler chickens, and insufficient number of study about the usage of crude glycerine in diets of birds; the aim of present trial was evaluated the effect graded level of dietary crude glycerin on growth and economic performance, carcass yield, organ weights and moisture levels of meat in broiler chickens.

MATERIAL and METHODS

In this experiment, 270 one-day-old male Ross 308 broiler chicks were randomly divided into 3 groups according to the dietary regimen and each group was constituted by 5 subgroups of 18 birds.

The experimental protocol was approved by Local Ethics Committee of Selcuk University.

In the control group, chickens were fed with basal standard diets based on corn and soybean meal by recommendations of NRC ^[13], more specifically with a starter diet [23% crude proteins (CP) and 3100 kcal/kg metabolically energy (ME)] for the first 10 d of trial then with a grower diet [22% CP and 3150 kcal/kg ME] for the between 11 to 28 d of experiment, and then with a finisher diet [21% CP and 3200 kcal/kg ME] for the 3rd and the last 14 days of trial (*Table 1*). Birds from the groups 2 and 3 received standard diets supplemented with 5% or 10% glycerol, respectively for the whole experimental period. Feed and water were provided *ad libitum*. The birds were housed in wire-bottomed pens fitted with electrical heaters during the 42 days experimental period. The temperature started at 33°C (from the 1st day to the 3rd day) and was gradually reduced (2-3°C/week) according to normal management practice. Chicks were maintained on a 24 h constant light schedule until the end of the experiment.

Body weights were determined by pen on days 1, 14, 28 and 42 and feed intakes were measured during each rearing period. Similarly, the cumulated body weight gains (expressed as g/bird and calculated body weight differences of birds from the 1st day to the 14th, 28th and 42nd days, respectively) and feed conversion ratios (expressed as g of consumed feed/weight gain in g) were calculated for each rearing period. On the day 42, 3 birds from each pen for which the body weight was closed to the mean value were slaughtered by cervical dislocation. The weight of the carcass, liver, spleen, gizzard, heart, glandular stomach and cloacal fat were recorded and all values were expressed as percentages of the carcass weight for a same bird. The duodeno-jejunal contents were collected from 3 other birds from each pen slaughtered on the d 42 and the samples were diluted with deionised water for determine pH levels of intestine.

By the end of trial, experiment groups were economically evaluated by using two procedures with a help of consideration the costs of feeds, labor, equipments, nursing and chicken meat from actual market prices; one of them was the total cost needed to obtain one-kilogram carcass weight and second was the net revenue per unit of total.

Data from present study were analyzed by Anova using SPSS 11.50 program (Inc., Chicago, IL, USA). Significant differences among treatment were determined using Duncan's multiple range tests ^[14] with a 5% level of probability.

RESULTS

The growth performance and the feed intakes in broilers according to the dietary treatments are summarized in the *Table 2*. The body weights (BW) and the body weight

Table 1. Feedstuffs and nutrient composition of experiment diets**Tablo 1.** Deneme rasyonlarının yem ham madde içerikleri ve besin madde değerleri

Ingredients	Feeding Stage								
	Starter (0 to 10 d)			Grower (11 to 28 d)			Finisher (29 to 42 d)		
Crude glycerin ¹	0	5	10	0	5	10	0	5	10
Corn	52.33	46.33	40.38	54.13	48.13	42.13	54.8	48.73	42.63
Soybean meal	32	33.05	34.00	31	32	33	31.13	32.2	33.3
Gluten	7	7	7	6.5	6.5	6.5	5	5	5
Vegetable oil	4	4	4	4.5	4.5	4.5	5.5	5.5	5.5
Limestone	1	1	1	0.9	0.9	0.9	0.9	0.9	0.9
DCP	2.25	2.25	2.25	2	2	2	2	2	2
Salt	0.4	0.4	0.4	0.4	0.4	0.4	0.4	0.4	0.4
Vitamin mix ²	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1
Mineral mix ³	0.07	0.07	0.07	0.07	0.07	0.07	0.07	0.07	0.07
Methionine	0.35	0.35	0.35	0.2	0.2	0.2	0.1	0.1	0.1
Lysine	0.50	0.45	0.45	0.2	0.2	0.2	0	0	0
Total	100	100	100	100	100	100	100	100	100
Calculated Analyses									
CP	23	23	23	22	22	22	21	21	21
ME	3107	3106	3107	3152	3153	3153	3201	3201	3200
Ca	1.02	1.03	1.03	0.92	0.93	0.93	0.92	0.93	0.93
Avail. P	0.50	0.50	0.50	0.46	0.46	0.46	0.46	0.46	0.46
Met+sist.	1.13	1.12	1.11	0.96	0.95	0.94	0.82	0.82	0.81
Lizin	1.46	1.44	1.45	1.2	1.21	1.22	1.03	1.04	1.06

¹ Chemical composition of crude glycerin; 4.13% ash, 0.17% salt, 9.11 pH, and 0.45% methanol; ² Vitamin premix supplied the following per kilogram of diet: vit-A 15.000 IU, vit-D₃ 5.000 IU, vit-E 50 mg, vit-K₃ 10 mg, vit-B₁ 4 mg, vit-B₁₂ 8 mg, vit-B₆ 5 mg, vit-B₁₂ 0.025 mg, niacin 50 mg, pantothenic acid 20 mg, folic acid 20 mg, biotin 0.25 mg, vit-C 75 mg, kolin 175 mg; ³ Mineral premix supplied the following per kilogram of diet: Mn 100 mg, Zn 150 mg, Fe 100 mg, Cu 20 mg, I 15 mg, Co 0.5 mg, Se 0.2 mg, Mo 1 mg, Mg 50 mg

gains (BWG) during the first 14 d of experiment showed no significant variance among treatment groups, whereas, BW were significantly increased in birds fed with 5% and 10% crude glycerine (CG) supplemented diet compared to birds fed with basal diet at 28 and 42 days of experiment ($P < 0.05$ and $P < 0.01$, respectively). Furthermore, the mean of BW and BWG results cumulated over the whole experimental period (1 to 42 d) were markedly increased in graded level CG supplemented broilers ($P < 0.05$ and $P < 0.01$, respectively). On the other hand, although the differences were not significant among groups, the highest feed intakes (FI) for the all periods (starter, grower and finisher) were also observed in birds supplemented with 5% CG. Feed conversion ratio (FCR) was not significantly altered in the CG supplemented birds compared to the controls at d 14 while it improved by 5% and 10% CG addition at 28th and 42nd days of experiment ($P < 0.001$ and $P < 0.01$, respectively).

As shown in Table 3, the carcass traits (including carcass yields, moisture of breast and drumstick), the relative organ weights (100 x organ weight/carcass weight) and intestinal pH levels were summarized. Even

though there were no significant variances in carcass traits according to the 3 dietary regimens, the lowest carcass yield was recorded for birds supplemented with 10% CG but differences among treatments were not significant. The relative gizzard ($P < 0.05$) weights were significantly decreased in birds supplemented with 10% CG compared to birds fed with basal diet. By contrast, cloacal fat percentages (not significant) were higher in 10% CG supplemented broilers than control group birds. Intestinal pH levels showed no significant differences between treatment groups, however supplemented birds (5% and 10% glycerine, respectively) present more alkaline pH compared with birds fed with basal diet.

The results for economic evaluation of graded level of dietary CG inclusion were summarized in Table 4. Our findings indicated that 5% dietary CG supplementation had improving effect on relative economic efficiency (REEF) in broiler chickens. Chicks fed with 5% dietary CG showed over 20% higher results than birds fed with basal diet. On the other hand when CG level increased (to 10%) REEF of chickens decreased over 5% than birds fed without any CG addition in their diets.

Table 2. Effects of dietary graded levels of crude glycerin (CG) supplementation on growth performance of broiler chickens**Tablo 2.** Rasyona artan düzeylerde ilave edilen ham gliserinin (HG) broylerlerde büyüme performansına etkileri

Treatment	Day 1	Days 1 to 14				Days 1 to 28				Days 1 to 42			
CG level (%)	BW, g	BW, g	BWG, g	FI, g	FCR ¹	BW, g	BWG, g	FI, g	FCR ¹	BW, g	BWG, g	FI, g	FCR ¹
0	46.21	362.56	316.35	427.39	1.35	1112.60 ^b	1066.39 ^b	1751.33	1.64 ^a	2110.73 ^b	2064.52 ^b	3734.48	1.81 ^a
5	46.12	373.03	326.92	430.28	1.32	1221.96 ^a	1175.84 ^a	1794.29	1.53 ^b	2282.20 ^a	2242.09 ^a	3813.86	1.70 ^b
10	46.14	359.72	313.59	408.28	1.30	1198.17 ^a	1152.03 ^a	1702.24	1.48 ^b	2228.90 ^a	2182.77 ^a	3649.83	1.67 ^b
SEM ²	0.17	3.74	3.76	5.34	0.01	18.67	18.65	16.94	0.02	25.75	25.75	31.27	0.02
P	NS	NS	NS	NS	NS	*	*	NS	***	**	**	NS	**

¹ FCR = Feed conversion ratio was calculated by dividing feed consumption (g) to BW gain (g) per pen basis, ² SEM = Standard error of the mean, NS = Not significant at P>0.05; * P<0.05; ** P<0.01; *** P<0.001, ^{a,b} Means within a treatment and column with different subscripts differ significantly

Table 3. Effects of dietary graded levels of crude glycerin (CG) supplementation on carcass yield (%), meat moisture levels (%), relative organ weights (g/100 g body weight), and intestinal pH in broiler chickens**Tablo 3.** Rasyona artan düzeylerde ilave edilen ham gliserinin (HG) broylerlerde karkas randımanı (%), et nem düzeyleri (%), göreceli organ ağırlıkları (g/100 g canlı ağırlık) ve bağırsak pH üzerine etkileri

CG Levels (%)	Carcass Yield	Moisture of Breast Meat	Moisture of Drumstick	Intestinal pH	Cloacal Fat Pad	Liver	Spleen	Heart	Gizzard	Glandular Stomach
0	74.92	75.92	77.04	6.28	1.34	2.09	0.11	0.46	1.41 ^a	0.37
5	74.59	76.35	75.74	6.30	1.42	1.84	0.09	0.42	1.34 ^{ab}	0.33
10	69.93	76.00	76.16	6.36	1.58	1.97	0.11	0.47	1.19 ^b	0.34
SEM	1.29	0.12	0.31	0.03	0.09	0.05	0.01	0.01	0.03	0.01
P	NS	NS	NS	NS	NS	NS	NS	NS	*	NS

NS = Not significant at P>0.05, ^{a,b} Means within a treatment and column with different subscripts differ significantly

Table 4. Effects of dietary graded levels of crude glycerin (CG) supplementation on economic efficiency in broiler chickens**Tablo 4.** Rasyona artan düzeylerde ilave edilen ham gliserinin (HG) broylerlerde ekonomik verimlilik üzerine etkileri

Cost of Items (Per Bird)	Level of Dietary CG (%)		
	0	5	10
Fixed costs (TL) ^a	1.52	1.52	1.52
Feed cost (TL)	4.48	4.64	4.51
Total cost (TL)	6.00	6.16	6.03
Carcass weight (CW, kg)	1.58	1.71	1.56
Cost/kg CW (TL)	3.80	3.60	3.87
Total revenue (TL) ^b	8.69	9.41	8.58
Net revenue (TL) ^c	2.69	3.25	2.55
Relative economic efficiency ^d	100.00	120.82	94.80

^a- All stable prices (including transport, bird and poultry house costs) of whole rearing period for each bird, ^b- Assuming that the selling price of one kilogram carcass weight is 5.50 TL, ^c- Net revenue per unit total cost, ^d- Relative economic costs of treatments when costs of birds fed with basal diet assume 100%

DISCUSSION

In present study, effects of CG (by-product of biodiesel industry) on the growth performance were evaluated in broiler chickens. It was determined that addition of CG into broiler diets at level of 5% and 10% showed significant promoting effects on growth performance (BW and BWG),

particularly in the growing and finisher period of rearing. The growth performance was similar during the first 14 d of trial in broiler chickens, after than it showed remarkable increased for birds fed with graded level CG supplemented diets. This result might be related with AME_n of glycerine 10% higher than corn ^[13] so it is a replacement of carbohydrates in diet ^[15]. Similarly, FCR results also showed significant variances among treatments not starter but grower and finisher rearing period of experiment. Dietary 5% and 10% CG supplementation resulted in depression of FCR values compare birds fed with basal diet. For present trial, graded level of CG addition to diet had improvement effect on FCR in birds. However, the feed intake has not significantly differed between groups for the whole experimental period but it was slightly increased (not significant) in birds consume graded level of CG in their diets at d 42, probably, palatability increasing affect of CG. These findings showed positive correlation with number of studies ^[4,7,12,16] which have reported dietary glycerol supplementation had positive effects on growth performance in broiler chickens. Suchy et al.^[12] substituted 50% of dietary soybean oil with pure and raw glycerol in a ratio of 1:2 in male and female broilers. They observed that BW of both male and female chickens in experimental groups was significantly higher than birds in control group at d15 and d40 of experiment (P<0.05, P<0.01, respectively). But contrary to our findings in present trial, they found that dietary glycerol addition in different forms (in pure and raw) had increasing effect on FI for whole 40d rearing period in broiler chickens.

Moreover, Simon et al.^[7] also observed improvement effect of dietary glycerol at levels of 5% and 10% on BW and BWG in broiler chickens. These differences between studies probably related to variances of glycerol content especially metabolizable energy density of products.

The results from present trial were in disagreement with previous studies which have reported glycerol addition had negative ^[5] or no effects ^[1,8,9] on growth parameters in birds. Lin et al.^[5] concluded that substitution of dietary energy source at a level of 42.2% with glycerol had significantly detrimental effects on FI. But Simon et al.^[8] observed different level of dietary glycerol addition (from 5% to 25%) had no negative effect on FI and FCR in birds. Similarly, Cerreta et al.^[9] found that birds fed diets with 10% glycerine addition showed significantly less FI than those fed diets with 0 or 5% glycerine and therefore had significantly reduced BW. On a contrary, Mclea et al.^[16] observed that increasing level of glycerin addition had improving effect on FCR in birds. These incompatible results from different studies may be related to variances of dietary glycerol level or different content of glycerol, particularly varied methanol and fatty acid content of glycerol.

Although the 10% of dietary CG addition tended to decreased the carcass yield in the present study, we noticed that inclusion of CG in broiler diets had no significant effect on carcass traits (including moisture of drumstick and breast meat) and relative organ weights (except gizzard) with an agreement with previous studies ^[9,12]. In present trial, we also observed that the relative cloacal fat weight tended to increase in broilers fed with glycerol supplemented, especially at a level of 10% CG due to overestimation of metabolically energy assigned to glycerine ^[18]. Similarly, Lessard et al.^[6] have also found out that inclusion of 5% glycerol had increased the cloacal fat pad weight. However, the variances among treatments had no statistical meaning for this finding for present trial. On the other hand, the relative gizzard weight ($P<0.05$) was significantly depressed in glycerol supplemented birds, especially at a level of 10% CG, compared to the control birds. These findings were in accordance with previous study in which Sehu et al.^[4] have also observed significant depression of relative gizzard weight with 10% dietary glycerol addition in broiler chickens.

Results from economic evaluation indicated that dietary CG inclusion, especially at a level of 5% had improvement effect on REEF in broiler chickens. However, same tendency for REEF was not observed for birds fed with 10% CG included diets. Contrary, these birds showed lesser REEF results compare than birds fed without any dietary CG inclusion, probably cause of they represent same carcass weight like as control birds with more expensive diet costs. This finding showed partly agreement with Abd-Elsamee et al.^[1] results, who found that increasing level of

dietary glycerine addition, particularly at a level of 6%, had improvement effect of economic efficiency in broilers.

In conclusion, the results of the current study indicate that graded level (5% and 10%) crude glycerine addition to diets, especially 5% CG, had improvement effects on growth performance (BW, BWG) and FCR in broilers chickens. Moreover, birds fed with 5% dietary CG had present over 20% higher relative economic efficiency compare than birds fed basal diet. However, all other parameters, including carcass parameters and organ weights (except gizzard), were not significantly affected by dietary CG inclusion. As a result, this experiment indicates that graded level of CG inclusion, particularly at a level of 5% CG, could be effectively used in diets with higher profitability in broiler chickens.

REFERENCES

1. Abd-Elsamee MO, Abdo ZMA, El-Manlawi MAF, Salim IH: Use of crude glycerin in broiler diets. *Egypt Poult Sci*, 30 (1): 281-295, 2010.
2. Thompson JC, He BB: Characterization of crude glycerol from biodiesel production from multiple feed stocks. *Appl Eng Agric*, 22, 261-265, 2006.
3. Min YN, Yan F, Liu FZ, Coto C, Waldroup PW: Glycerin-a new energy source for poultry. *Inter J Poult Sci*, 9 (1): 1-4, 2010.
4. Sehu A, Kucukersan S, Coskun B, Koksall BH, Citil OB: Effects of dietary glycerol addition on growth performance, carcass traits and fatty acid distribution in cloacal fat in broiler chickens. *Rev Med Vet*, 163 (4): 194-200, 2012.
5. Lin MH, Romsos DR, Leveille GA: Effect of glycerol on enzyme activities and on fatty acid synthesis in the rat and chicken, *J Nutr*, 106, 1668-1677, 1976.
6. Lessard P, Lefrancois MR, Bernier JF: Dietary addition of cellular metabolic intermediates and carcass fat deposition in broilers. *Poult Sci*, 72, 535-545, 1993.
7. Simon A, Bergner H, Schwabe M: Glycerol feed ingredient for broiler chickens. *Arch Anim Nutr*, 49, 103-112, 1996.
8. Simon A, Schwabe M, Bergner H: Glycerol supplementation to broilers rations with low crude protein content. *Arch Anim Nutr*, 50, 271-282, 1997.
9. Cerrate S, Yan F, Wang Z, Coto C, Sacakli P, Waldroup PW: Evaluation of glycerin from biodiesel production as a feed ingredient for broilers, *Inter J Poult Sci*, 5 (11): 1001-1007, 2006.
10. Erol H, Yalçin S, Midilli M, Yalçin S: The effects of dietary glycerol on growth and laying performance, egg traits and some blood biochemical parameters in quails. *Rev Med Vet*, 160, 469-476, 2009.
11. Yalcin S, Erol H, Ozsoy B, Onbasilar I, Yalcin S, Uner A: Effects of glycerol on performance, egg traits, some blood parameters and antibody production to SRBC of laying hens. *Livest Sci*, 129, 129-134, 2010.
12. Suchy P, Strakova E, Kroupa L, Herzig I: Pure and raw glycerol in the diet of broiler chickens, its effect on the production parameters and slaughter value. *Arch Tierz*, 54 (3): 308-318, 2011.
13. NRC, National Research Council: Nutrient Requirements for Poultry. 9th ed., pp.26-34, National Academy Press, Washington, DC., 1994.
14. Duncan DB: Multiple range and multiple F Test. *Biometrics*, 11, 1-42, 1955.
15. Dozier III WA, Kerr BJ, Corzo A, Kidd MT, Weber TE, Bregendahl K: Apparent metabolizable energy of glycerin for broiler chickens. *Poult Sci*, 87, 317-322, 2008.
16. Kroupa L, Suchy P, Strakova E, Herzig I: Glycerol as source of energy in broiler chicken fattening. *Acta Vet Brno*, 80, 157-164, 2011.

17. Mclea L, Ball ME, Kilpatrick D, Elliot C: The effect of glycerol inclusion on broiler performance and nutrient digestibility. *Br Poult Sci*, 52 (3): 368-375, 2011.

18. Narayan KA, Mullen JJ, Wakefield T, Calhoun WK: Influence of dietary glycerol on the serum lipoproteins of rats fed a fat-free diet. *J Nutr*, 107, 2153-5163, 1977.

Cloning and Expression of *Cellulosimicrobium cellulans* β -1,3-Glucanase Gene in *Lactobacillus plantarum* to Create New Silage Inoculant for Aerobic Stability^{[1][2]}

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Summary

In this study, the recombinant plasmid pTE353- β G was created by inserting the p353-2 cryptic plasmid region of pLP3537 into pTEG5 recombinant plasmid contains pUC18 and β -1,3-glucanase gene of *Cellulosimicrobium cellulans*. The recombinant plasmid pTE353- β G was then introduced into *Lactobacillus plantarum* by electroporation. Insert analysis of pTE353- β G digested with *SacI* produced 1.9 kbp β -1,3-glucanase gene band on agarose gel as well as 1.9 kbp DNA encoding β -1,3-glucanase gene insert amplified on the recombinant vector via PCR indicated the integration of the gene into the plasmid. Recombinant *L. plantarum* colonies with pTE353- β G on MRS-laminarin-agar plate showed clear positive zones by Congo-red staining that revealed the expression of β -1,3-glucanase encoding gene. The β -1,3-glucanase enzyme of recombinant strain produced the same activity band with *C. cellulans* enzyme in terms of molecular weight, which showed the activity of secreted protein without any proteolytic degradation. Optimal temperature and pH values of *L. plantarum* β -1,3-glucanase have been determined 40°C and 6.0 respectively, by enzymatic analysis. These results revealed that recombinant *L. plantarum* could be considered as a silage inoculant for aerobic spoilage of silage.

Keywords: *Cellulosimicrobium cellulans*, β -1,3-Glucanase, *Lactobacillus plantarum*, Cloning, Gene expression, Recombinant silage inoculant

Aerobik Stabilité için Yeni Silaj İnokülantı Oluşturmak Amacıyla *Cellulosimicrobium cellulans* β -1,3-Glukanaz Geninin *Lactobacillus plantarum*'da Klonlanması ve Ekspresyonu

Özet

Bu çalışmada, rekombinant pTEG5 plazmit DNA'sına (pUC18 + *Cellulosimicrobium cellulans*'ın β -1,3-glukanaz geni) pLP3537 plazmitinin p353-2 kriptik plazmit bölgesinin aktarılması ile rekombinant pTE353- β G plazmit DNA'sı oluşturulmuştur. Rekombinant pTE353- β G plazmiti *Lactobacillus plantarum*'a elektroporasyon tekniği ile transfer edilmiştir. *SacI* enzimi ile kesilmiş pTE353- β G'nin agaroz jelde 1.9 kbç büyüklüğünde gen bandı üretmesinin yanı sıra, β -1,3-glukanaz genini şifreleyen 1.9 kbç büyüklüğündeki DNA parçasının PCR ile rekombinant vektörden amplifiye edilmesi genin plazmite entegrasyonunu göstermektedir. pTE353- β G içeren rekombinant *L. plantarum* kolonileri MRS-laminarin-agar plağında Congo-red boyaması ile β -1,3-glukanaz geninin eksprese olduğunu gösteren açık renkli pozitif zon vermişlerdir. Rekombinant suş tarafından üretilen enzim, *C. cellulans* enzimi ile aynı moleküler ağırlıkta aktivite bandı üretmiş ve böylece enzimin herhangi bir proteolitik parçalanmaya maruz kalmadığı anlaşılmıştır. Enzimatik analizler sonucunda *L. plantarum* tarafından üretilen β -1,3-glukanaz enziminin optimum sıcaklık ve pH değerleri sırasıyla 40°C ve 6.0 olarak bulunmuştur. Bu sonuçlar, rekombinant *L. plantarum*'un silajın aerobik bozulmasını önlemek için inokülant olarak düşünülebileceğini ortaya çıkarmıştır.

Anahtar sözcükler: *Cellulosimicrobium cellulans*, β -1,3-Glukanaz, *Lactobacillus plantarum*, Klonlama, Gen ifadesi, Rekombinant silaj inokülantı



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INTRODUCTION

Endo-1,3- β -glucanases (EC 3.2.1.6 and EC 3.2.1.39) are widely distributed among bacteria and higher plants [1]. These enzymes catalyse the hydrolysis of β -1,3-glucan component found in the yeast cell wall and other β -1,3-glucans such as laminarin, curdlan and pachyman [2,3]. The bacterium *Cellulosimicrobium cellulans* (also known with the synonyms *Cellulomonas cellulans*, *Oerskovia xanthineolytica*, and *Arthrobacter luteus*) has been regarded as a major source of yeast-lytic enzymes, particularly endo- β -1,3-glucanases, proteases and mannanases [4]. Commercially available yeast-lytic glucanase preparations derived from this organism, namely Lyticase, Zymolyase, and Quantazyme, have been produced and widely used for yeast protoplast preparation and yeast DNA isolation [4-6]. Only one of these preparations (Quantazyme, Quantum Biotechnology, Canada) is produced recombinantly and protease-free [4,5].

The primary goal of making silage is to maximize the preservation of original nutrients in the forage crop for feeding at a later date [7]. Therefore, ensiled forages are the most commonly used feeds for ruminants all over the world [8]. *Lactobacillus plantarum* and other *Lactobacillus* species, *Enterococcus faecium* and *Pediococcus* species are most common silage inoculant bacteria and one or more of these bacteria to be included in silage inoculants [7,9]. One of these bacteria, *L. plantarum*, is the most important bacterium used in silage fermentation. However, aerobic spoilage by yeasts and moulds is a major cause of reduced nutritional value of silage and increases the risk of potential pathogenic microorganisms [10]. Various chemical additives with anti-fungal properties, such as propionic acid, sorbate, benzoate, acetic acid, ammonia, urea, some enzyme preparations and *Lactobacillus buchneri* as silage inoculant have been used to prevent or enhance of aerobic stability and decrease of spoilage [11]. However, usage of most of these additives has been restricted because of their other undesirable properties. For instance, propionic acid is difficult to handle because of its corrosive nature as well as at a pH of 4.8, about 50% of the acid being undissociated [7,11], sorbate, benzoate, and acetic acid being too costly [11]. Besides, *L. buchneri* as silage inoculant causes dry matter losses in silage material at early fermentation phase [12].

In the present study, we aimed to express the β -1,3-glucanase gene of *C. cellulans* in *L. plantarum* to create a recombinant silage inoculant to enhance the aerobic stability and decrease the spoilage by secreting the β -1,3-glucanase enzyme by *L. plantarum*.

MATERIAL and METHODS

Strains of Bacteria and Growth Conditions

Lactobacillus plantarum strain 5057 was cultured on MRS broth (Merck, 1.10661) and MRS agar (Merck, 1.10660)

at 37°C without shaking. For culturing of recombinant *L. plantarum*, both MRS-broth and MRS-agar supplemented with ampicillin (50 μ g mL⁻¹). *Cellulosimicrobium cellulans* (*Oerskovia xanthineolytica*, ATCC 21606) was cultured in GYM Streptomyces medium (glucose (0.4% wt/v), yeast extract (0.4% wt/v), malt extract (1% wt/v), pH 7.2) at 28°C with shaking at 250 rpm. Agar (1.2% wt/v) and CaCO₃ (0.2% wt/v) were added into GYM Streptomyces medium for preparation of GYM Streptomyces agar. Recombinant *Escherichia coli* carrying recombinant pTEG5 plasmid DNA was cultured in LB-broth (10 g bacto tryptone (Merck), 5 g yeast extract (Merck) and 10 g NaCl (Merck) per L, pH 7.5) at 37°C with shaking at 250 rpm. Agar (1.5% wt/v) was added into LB medium for LB-agar. Both LB-broth and LB-agar were supplemented with ampicillin (50 μ g mL⁻¹). For activity testing of recombinant *L. plantarum* on MRS-laminarin-agar plate, the plate was stained with Congo-red solution (0.1% wt/v Congo-red) for 15 min and then destained with 1 M NaCl solution for 15 min. Clear haloes on the MRS-laminarin-agar plate indicated the presence of β -1,3-glucanase activity [13].

Plasmids

Recombinant vector pTEG5 was previously created in the Animal Biotechnology and Genetic Engineering Laboratory, Department of Animal Science, Faculty of Agriculture, Çukurova University [14]. The recombinant vector pTE353- β G (pUC18 + β -1,3-glucanase gene + p353-2 cryptic plasmid) was created using *Escherichia coli*/*Lactobacillus* shuttle vector pLP3537 (ampicillin resistant and erythromycin resistant; amp^R, er^R) [15] and pTEG5. pTE353- β G was used for *L. plantarum* transformation [16]. Recombinant *L. plantarum* strain carrying the recombinant plasmid pTE353- β G, was cultured in MRS broth and on MRS agar at 37°C supplemented with ampicillin (50 μ g mL⁻¹). Recombinant plasmid pTEG5 was isolated from *E. coli* cells as described previously [17]. On the other hand, pTE353- β G was isolated from recombinant *L. plantarum* cells as described previously [18].

DNA Modification

The following modifying enzymes were purchased and used for DNA modifications; *SacI*, *SmaI*, *EcoRI*, *HindIII*, and bacterial alkaline phosphatase, bacteriophage T4 DNA ligase as well as Pfu DNA polymerase (Fermentas, Vivantis and Promega Corporation). Restriction enzyme reactions were monitored by examining digestion by agarose gel electrophoresis using standard methods [19]. Linearised plasmid DNA and PCR products were excised from gels and purified using Genomic DNA Purification Kit (Fermentas).

Cloning Procedures

p353-2 cryptic plasmid region (2.4 kbp) was derived from pLP3537 vector by digestion with *EcoRI* and then ligated to *EcoRI* digested pTEG5 vector to construct the pTE353- β G recombinant plasmid.

The pTE353- β G was electrotransformed into *Lactobacillus plantarum* 5057 strain using the modified method^[16] as follows: For electroporation, 50 μ L of the competent *L. plantarum* cells were mixed with 1 μ g of plasmid DNA and transferred to a prechilled electroporation cuvette (1 mm gap). After incubation for 2-3 min, the cells were exposed to a single electrical pulse using a Gene-Pulser (Invitrogen) set at 25 μ F, 200 Ω , 1300 V (13 kV cm⁻¹) and 25 mA resulting in time constant of 5 ms. After electroporation, electrotransformed *L. plantarum* cells were diluted with 500 μ L MRS broth and incubated at 32°C for 4 h. Finally the cells were plated on MRS-agar containing ampicillin (50 μ g mL⁻¹) and incubated at 37°C for 48 h.

PCR and Restriction Endonuclease Analysis

The sequences of the primers used for amplifying of β -1,3-glucanase gene from the recombinant vector were 5'-AGAGCTCGTGGCACTGCACTCGTTCGAGTCT-3' (forward) and 5'-AGAGCTCGACGGGCGCGGTGAGAGCGTCCAG-3' (reverse) based on the gene sequence^[20]. The PCR mixture consisted of 5 μ L of reaction buffer, 1 μ L of 40 mM dNTP mix (200 μ M each final), 1 μ L each of forward and reverse primers (20 pmol each primer), 0.5 μ L of Pfu DNA polymerase (2.5 U/ μ L), 1 μ L of 50% wt/v DMSO (1% wt/v final), and 210 ng of template in a total volume of 50 μ L. The following amplification program was used: Initial denaturation step at 94°C for 2 min, then 30 cycles of denaturation at 98°C for 10 s, annealing and elongation at 68°C for 5 min. A final extension step was performed as 72°C for 5 min. PCR reaction performed with recombinant plasmid analysed by agarose gel (0.8% wt/v) electrophoresis.

For restriction endonuclease analysis, recombinant plasmid pTE353- β G was isolated from *L. plantarum* transformant. To detect the β -1,3-glucanase gene, recombinant plasmid was cleaved with *SacI*+*EcoRI* restriction endonuclease mixture and analysed by agarose gel (0.8% wt/v) electrophoresis. Furthermore, pTE353- β G was digested by *HindIII* to obtain a single linearized band.

Electrophoretic Analysis of Extracellular Proteins

To obtain the extracellular proteins of *C. cellulans* and *L. plantarum* strains from culture supernatants, the overnight grown cells were pelleted by centrifuge. The extracellular extracts (supernatants) were mixed with 1:1 volume of 20% TCA for precipitation. After the incubation at room temperature for overnight, protein pellets were obtained by centrifuge. Air-dried proteins were dissolved in 0.1 M Tris-HCL buffer (pH 8.0).

SDS-PAGE and SDS-Laminarin-PAGE (0.2% laminarin) were done as described previously^[21] with slab gels (12% wt/v acrylamide). After the electrophoresis, the gel was stained for 1 h with Coomassie blue R 250 dye in methanol-acetic acid-water solution (4:1:5 by volume) and destained in the same solution without dye^[22,23]. For activity staining (zymogram analysis), SDS was removed by washing the

gel at room temperature in solutions containing 50 mM Na₂HPO₄, 50 mM NaH₂PO₄ (pH 7.2), isopropanol 20% v/v for 1 h and 50 mM Na₂HPO₄, 50 mM NaH₂PO₄ (pH 7.2) for 1 h, respectively. Renaturation of enzyme proteins was carried out by keeping the gel overnight in a solution containing 50 mM Na₂HPO₄, 50 mM NaH₂PO₄ (pH 7.2), 5 mM β -mercaptoethanol and 1 mM EDTA at 4°C. The gel was then transferred onto a glass plate, sealed with film, and incubated at 30°C for 4 h. The gel was stained in a solution of Congo-red (0.1% Congo-red, 0.2 M NaOH), for 1 h, and destained in 1 M NaCl for 30 min. Clear bands indicated the presence of β -1,3-glucanase activity^[24-26].

Enzyme Assay

β -1,3-Glucanase activity was assayed by adding 1 mL enzyme to 1 mL laminarin (2%, wt/v) in 0.1 M Na-phosphate buffer, pH 6.0 and incubating at 40°C for 30 min. The reaction was stopped by addition of 3 mL of 3,5-dinitrosalicylic acid reagent and A_{540 nm} was measured in a Pharmacia spectrophotometer^[27].

Temperature and pH effects on enzyme activity were assayed at different temperatures ranging from 20 to 100°C and at pH values ranging from 4 to 11 for 30 min. Following buffers were used in the reactions: 50 mM Na-acetate (pH 4-6), 50 mM Na-phosphate (pH 6-8) and 50 mM Tris (pH 8-11)^[28]. Enzyme assay carried out as described previously.

For the measurement of thermal stability, the enzyme was pre-incubated at temperatures between 20 to 100°C for 30 min at optimum pH. The enzyme activity was determined under standard enzyme assay condition. Enzyme assay carried out as described previously.

Supernatants of recombinant *L. plantarum* were taken at predetermined time intervals (0 min, 12, 24, 36, 48, 60, 72 h) of culturing period. Enzyme assay of supernatants carried out as described previously and enzyme production depending on culturing period determined.

RESULTS

Transformation of *Lactobacillus plantarum*

The first generation recombinant vector pTEG5 (pUC18 plus β -1,3-glucanase gene of *C. cellulans*) was created previously^[14]. In present study, the 2.4 kbp *EcoRI*-*EcoRI* p353-2 cryptic plasmid region including replication origin of *L. plantarum* was derived from the pLP3537 and ligated to pTEG5 to create the second generation recombinant plasmid pTE353- β G (Fig. 1).

The pTE353- β G recombinant plasmid was then electrotransformed into *L. plantarum* to express the β -1,3-glucanase gene. Recombinant *L. plantarum* colonies showed β -1,3-glucanase activity on MRS-agar plate supplemented with ampicillin (50 μ g mL⁻¹) and laminarin (0.1% wt/v) with clear

zones around of the recombinant colonies (Fig. 2).

While original *L. plantarum* with no ampicillin resistance gene did not grow on solid growth medium contain ampicillin, recombinant *L. plantarum*/pTE353-βG bearing *Amp* resistance gene on the vector grew well on the same medium (data not shown).

Recombinant pTE353-βG plasmid was isolated from recombinant *L. plantarum* strain. It was then subjected

to restriction fragment length analysis together with PCR amplified DNA encoding the gene on agarose gel electrophoresis (0.8% wt/v). β-1,3-glucanase gene fragment (~1.9 kbp) amplified by PCR and restriction endonuclease digested recombinant plasmid confirmed the success of the cloning experiments. Recombinant plasmid digested with *SacI*+*EcoRI* was yielded the same DNA fragment consisting of pUC18, p353-2 and β-1,3-glucanase gene (Fig. 3).

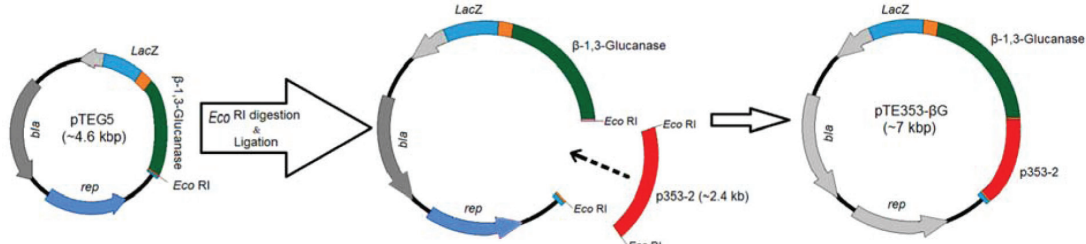


Fig 1. Construction of pTE353-βG plasmid (~7 kbp) by ligating p353-2 cryptic plasmid region including replication origin of *L. plantarum* (~2.4 kbp) into pTEG5 (~4.6 kbp)

Şekil 1. *L. plantarum* replikasyon orijinini içeren p353-2 kriptik plazmit bölgesinin (~2.4 kb) pTEG5'e (~4.6 kb) ligasyonu ile pTE353-βG plazmitinin (~7 kb) oluşturulması

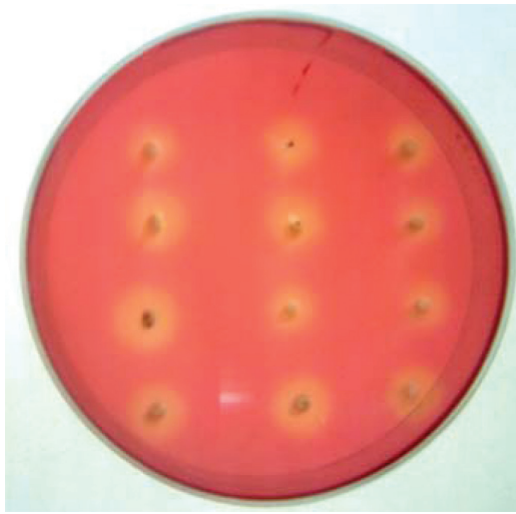


Fig 2. Recombinant *L. plantarum* colonies showing clear β-1,3-glucanase activity on MRS-agar plate with Congo red staining

Şekil 2. MRS-agar plağında Congo-red boyaması ile β-1,3-glukanaz aktivitesi gösteren rekombinant *L. plantarum* kolonileri

Culture supernatants of *C. cellulans*, *L. plantarum*/pTE353-βG, and *L. plantarum* were applied to SDS-PAGE and SDS-Laminarin-PAGE to visualize total proteins and zymogram analysis, respectively. For zymogram analysis, denaturated proteins were renaturated on SDS-Laminarin-PAGE after removing denaturing agents from the gel and then allowing to the enzyme to digest substrate, thereby producing clear zones on the gel. On zymogram analysis, only β-1,3-glucanase protein band of *C. cellulans* with 54.5 kDa in size was showed a clear zone together with extracellular protein counterpart of other recombinant *L. plantarum* strain (Fig. 4). It is clearly indicated that β-1,3-glucanase gene of *C. cellulans* successfully expressed in *L. plantarum* without any significant proteolytic degradation.

Enzyme Properties

The optimum activity of the enzyme isolated from recombinant *L. plantarum* was observed at 40°C. The mean enzyme activities were 91, 86 and 62% at 30, 50 and 60°C

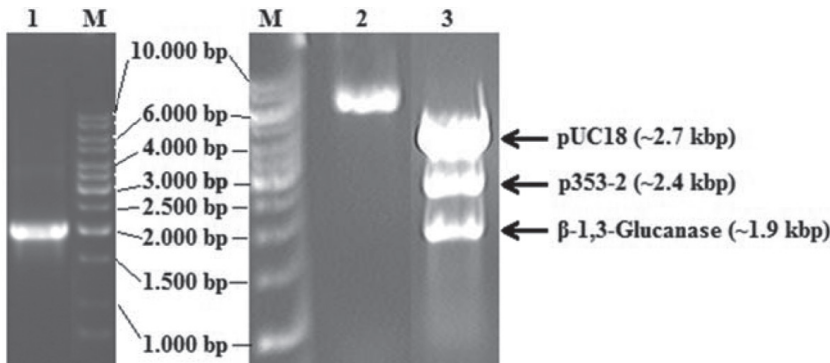


Fig 3. Restriction endonuclease and PCR analysis of pTE353-βG plasmid on agarose gel (M: 1 kbp DNA marker, 1: PCR amplified fragment of β-1,3-glucanase gene from pTE353-βG, 2: pTE353-βG/*HindIII*, 3: pTE353-βG/*SacI*+*EcoRI*)

Şekil 3. pTE353-βG plazmitinin agaroz jelde restriksiyon endonükleaz ve PCR analizleri (M: 1 kb DNA markır, 1: pTE353-βG'dan PCR ile amplifiye edilmiş β-1,3-glukanaz gen fragmenti, 2: pTE353-βG/*HindIII*, 3: pTE353-βG/*SacI*+*EcoRI*)

respectively, whereas only 23% activity was retained after incubation at 70°C for 30 min (Fig. 5A). The enzyme also showed a significant relative activity (80.6%) between pH 5.0 and 7.0 with an optimum pH of 6.0 (Fig. 5B). For thermal stability estimation, the retaining activity was

determined at optimum pH and temperature (Fig. 5C). The retained original enzyme activity obtained from 20 to 60°C was 94% for 15 min. The enzyme was stable for 15 min between at 20 and 40°C, while at 50, 60, 70 and 80°C, 9, 22, 71 and 87% of the original activities were lost, respectively.

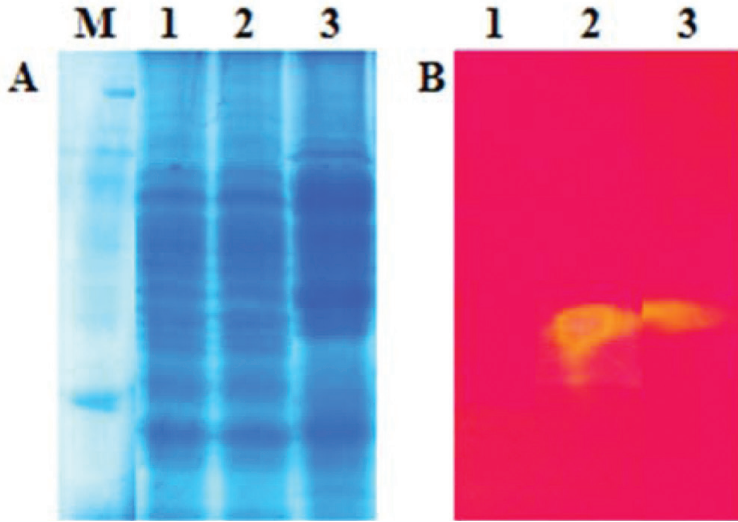


Fig 4. SDS-PAGE (A) and SDS-Laminarin-PAGE (B) analysis of recombinant and non-recombinant bacterial proteins (M: Marker, 1: *L. plantarum*, 2: *L. plantarum*/pTE353-βG, 3: *C. cellulans*)

Şekil 4. Rekombinant ve rekombinant olmayan bakteriyel proteinlerin SDS-PAGE (A) ve SDS-Laminarin-PAGE (B) analizleri (M: Markır, 1: *L. plantarum*, 2: *L. plantarum*/pTE353-βG, 3: *C. cellulans*)

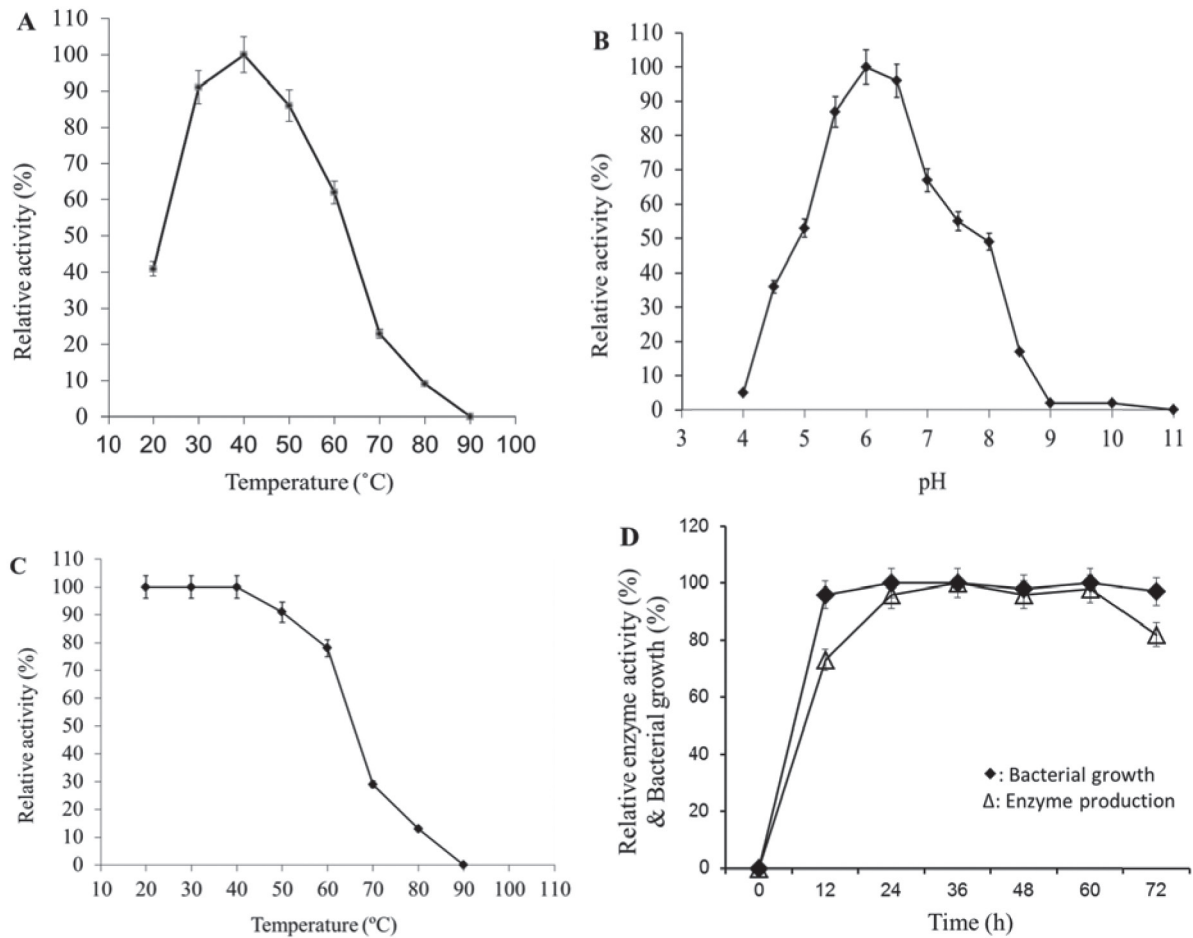


Fig 5. Enzyme properties of recombinant β-1,3-glucanase (A: Effect of temperature, B: Effect of pH, C: Thermal stability, D: Enzyme production depending on bacterial growth)

Şekil 5. Rekombinant β-1,3-glukanazın enzimatik özellikleri (A: Sıcaklığın etkisi, B: pH'nın etkisi, C: Termal stabilite, D: Bakteri gelişimine bağlı enzim üretimi)

The production of β -1,3-glucanase by the recombinant *L. plantarum* was reached maximum activity at 36 h of incubation period (Fig. 5D). After 60 h of incubation period, the enzyme activity started to decrease.

DISCUSSION

With this study, β -1,3-glucanase gene of *C. cellulans* was cloned and expressed in *L. plantarum*. The enzyme secreted from *L. plantarum* was found to be active with clear zones on MRS-agar plate containing laminarin. On the other hand, zymogram analyses clearly indicated that activity bands with same size surrounded with clear zones confirming the expression of *C. cellulans* β -1,3-glucanase gene in *L. plantarum* successfully without any significant proteolytic degradation.

β -1,3-Glucanase gene of *C. cellulans* (ATCC 21606) was well studied and enzymatic properties were revealed [29]. Furthermore, the nucleotide sequence of the gene was determined [20]. In our previous study, the DNA fragment of gene was generated by PCR from *C. cellulans* genome, ligated into *Sma*I digested pUC18 and then first generation recombinant vector pTEG5 was created [14]. In the present study, p353-2 cryptic plasmid region (2.4 kbp) including replication origin of *L. plantarum* was derived from *Escherichia coli*/*Lactobacillus* shuttle vector pLP3537 then ligated into *Eco*RI digested pTEG5 and second generation recombinant vector pTE353- β G was created. The pTE353- β G recombinant vector was then successfully transferred into *L. plantarum* 5057.

Silage can spoil rapidly if exposed to air during storage or feed out stage of silage. Although a common misconception is that molds are responsible for spoilage of silage, yeasts are the primary microorganisms that cause aerobic spoilage and heating [7]. Some chemical additives and inoculant microorganisms were used to increase the aerobic stability and to prevent the spoilage of silage. Molecular studies on silage inoculants have been increased and then some genes were cloned in lactic acid bacteria for improvement for DM losses.

Aerobic deterioration of silage is a complex process which depends on many factors and usually it is initiated by aerobic yeasts [30]. Various chemical additives with antifungal properties, such as propionic acid, sorbate, benzoate, acetic acid, ammonia, urea, some enzyme preparations and *Lactobacillus buchneri* as silage inoculant have been used to prevent or enhance of aerobic stability and decrease of spoilage [11]. However usages of most of these additives have been restricted because of their other undesirable properties.

Previously, several genes from other microorganisms were cloned and expressed in *L. plantarum* such as α -amylase [31-34], cellulase, xylanase and endoglucanase [8,35], endo-1,4- β -glucanase [36], levanase [37], β -galactosidase [38]

and chitinase [39]. Besides, herbal β -1,3-glucanase genes from soybean [40], jujube fruit [41] and rice [42] were cloned previously. On the other hand, the *C. cellulans* β -1,3-glucanase gene was cloned and expressed in *Bacillus subtilis* and *E. coli* [1-3,20,43]. But this is the first report to the best of our knowledge in which the β -1,3-glucanase gene from *C. cellulans* was cloned and expressed in *L. plantarum* to create new silage inoculant for better aerobic stability.

In our study, the optimum temperature of the recombinant enzyme was found with a wide range between 30-60°C. Although the gene has been cloned in several bacteria including *E. coli* and *B. subtilis*, temperature properties have not been investigated. On the other hand, temperature optimums of the β -1,3-glucanases from strain DSM 10297 and TK-1 were reported as 40 and 60°C, respectively [4]. Our finding is in agreement with optimum temperatures of the enzymes from the strains DSM 10297 and TK-1. On the other hand, optimum pH of our recombinant enzyme was found as 6.0 with a significant relative activity (80.6%) between pH 5.0 and 7.0. The present study indicates that, this enzyme prefers slightly acidic pH for optimal activity. Similar pH conditions for activity of native β -1,3-glucanase from *C. cellulans* have been reported previously [29]. pH values of various silages observed in other studies were reported as 4.3-4.5 (alfalfa silage with 30-35% DM), 4.7-5.0 (alfalfa silage with 45-55% DM), 4.3-4.7 (grass silage with 25-35% DM), 3.7-4.2 (corn silage with 35-40% DM), and 4.0-4.5 (high moisture corn silage with 70-73% DM) [7]. Our enzyme has been protected almost 40 and 55% of its activity in 4.0 and 5.0 pH conditions, respectively. With our results, the recombinant enzyme could be considered as a solution partially for aerobic spoilage at feed out stage. Besides, considering the development of *L. plantarum* at 30°C, this temperature value is quite appropriate for activation of the recombinant enzyme. Our enzyme has been suffered the loss of activation over 60°C for 30 min. So, under the unsuitable silage conditions with 60°C, our enzyme began to denature.

In conclusion, the β -1,3-glucanase gene of *C. cellulans* cloned in *L. plantarum*. Recombinant *L. plantarum* expressing β -1,3-glucanase encoding gene could be considered as a silage inoculant for aerobic spoilage of silage.

REFERENCES

1. Tanabe Y, Pang Z, Oda M: Cloning and sequencing of endo-1,3- β -glucanase from *Cellulosimicrobium cellulans*. *J Biol Macromol*, 8 (3): 60-63, 2008.
2. Salazar O, Molitor J, Lienqueo ME, Asenjo JA: Overproduction, purification, and characterization of β -1,3-glucanase type II in *Escherichia coli*. *Protein Express Purif*, 23 (2): 219-225, 2001.
3. Ferrer P, Halkier T, Hedegaard L, Savva D, Diers I, Asenjo JA: Nucleotide sequence of a β -1,3-glucanase isoenzyme II_A gene of *Oerskovia xanthineolytica* LL G109 (*Cellulomonas cellulans*) and initial characterization of the recombinant enzyme expressed in *Bacillus subtilis*. *J Bacteriol*, 178 (15): 4751-4757, 1996.
4. Ferrer P: Revisiting the *Cellulosimicrobium cellulans* yeast-lytic β -1,3-

glucanases toolbox: A review. *Microb Cell Fact*, 5 (10): 1-8, 2006.

5. Salazar O, Asenjo JA: Enzymatic lysis of microbial cells. *Biotechnol Lett*, 29 (7): 985-994, 2007.

6. Kitamura K, Yamamoto Y: Purification and properties of an enzyme, zymolyase, which lyses viable yeast cells. *Arch Biochem Biophys*, 153 (1): 403-406, 1972.

7. Kung L: Silage fermentation and additives. In: *Directfed Microbial Enzyme and Forage Additive Compendium*. pp. 1-18, Miller Publishing Co. Minnetonka, 2001.

8. Bates EEM, Gilbert HJ, Hazlewood GP, Huckle J, Laurie JI, Mann SP: Expression of a *Clostridium thermocellum* endoglucanase gene in *Lactobacillus plantarum*. *Appl Environ Microbiol*, 55 (8): 2095-2097, 1989.

9. Zahiroddini H, Baah J, McAllister TA: Effects of microbial inoculants on the fermentation, nutrient retention, and aerobic stability of barley silage. *Asian Australas J Anim Sci*, 19 (10): 1429-1436, 2006.

10. Driehuis F, Oude Elferink SJWH, Van Wikselaar PG: Fermentation characteristics and aerobic stability of grass silage inoculated with *Lactobacillus buchneri*, with or without homofermentative lactic acid bacteria. *Grass Forage Sci*, 56, 330-343, 2001.

11. Kung L: Aerobic stability of silages. *Proceedings of the Silage for Dairy Farms*. University of Delaware, Department of Animal and Food Sciences, Harrisburg, PA, 2005.

12. Holzer M, Mayrhuber E, Dannerr H, Braun R: The role of *Lactobacillus buchneri* in forage preservation. *Trends Biotechnol*, 21, 282-287, 2003.

13. Teather RM, Wood PJ: Use of Congo red polysaccharide interactions in enumeration and characterization of cellulolytic bacteria from the bovine rumen. *Appl Environ Microbiol*, 43 (4): 777-780, 1982.

14. Özcan BD, Özcan N, Baylan M, Güzel AI: Cloning and expression of β -1,3-glucanase gene from *Cellulosimicrobium cellulans* in *Escherichia coli* DH5a. *Kafkas Univ Vet Fak Derg*, 19 (3): 523-528, 2013, DOI: 10.9775/kvfd.2012.8225.

15. Posno M, Heuvelmans PTHM, van Giezen MJF, Lokman BC, Leer RL, Pouwels PH: Complementation of the instability of *Lactobacillus* strains to utilize d-xylose with d-xylose catabolism-encoding genes of *Lactobacillus pentosus*. *Appl Environ Microbiol*, 57 (9): 2764-2766, 1991.

16. Alegre MT, Rodriguez MC, Mesas JM: Transformation of *Lactobacillus plantarum* by electroporation with *in vitro* modified plasmid DNA. *FEMS Microbiol Lett*, 241, 73-77, 2004.

17. Birnboim HC, Doly J: A rapid alkaline extraction procedure for screening recombinant plasmid DNA. *Nucleic Acids Res*, 7 (6): 1513-1523, 1979.

18. Hardy KG: *Bacillus* Cloning Methods, In: Glover DM (Ed): *DNA Cloning: A Practical Approach*, Vol: II. 1-36, IRL Press Inc., Oxford, United Kingdom, 1985.

19. Sambrook J, Fritsch EF, Maniatis T: *Molecular Cloning: A Laboratory Manual*. Cold Spring Harbor Laboratory, 1989.

20. Shen S-H, Chretien P, Bastien L, Slilaty SN: Primary sequence of the glucanase gene from *Oerskovia xanthineolytica*. *J Biol Chem*, 266 (2): 1058-1063, 1991.

21. Laemmli UK: Cleavage of structural proteins during the assembly of the head of the bacteriophage T4. *Nature*, 227 (5259): 680-685, 1970.

22. Saul DJ, Williams LC, Grayling RA, Chamley LW, Love DR, Bergquist PL: *celB*, a gene coding for a bifunctional cellulase from the extreme thermophile *Caldocellum saccharolyticum*. *Appl Environ Microbiol*, 56 (10): 3117-3124, 1990.

23. Ozcan N, Cunningham C, Haris WJ: Cloning of a cellulase gene from the rumen anaerobe *Fibrobacter succinogenes* SD35 and partial characterization of the gene product. *Lett Appl Microbiol*, 22 (1): 85-89, 1996.

24. Lee SP, Morikawa M, Takagi M, Imanaka T: Cloning of the *aapT* gene and characterization of its product, α -amylase-pullulanase (AapT), from thermophilic and alkaliphilic *Bacillus* sp. strain XAL601. *Appl Environ Microbiol*, 60 (10): 3764-3773, 1994.

25. Ikeda T, Yamazaki H, Yamashita K, Shinke R: The tetracycline inducible expression of alpha-amylase in *Bacillus subtilis*. *J Ferment Bioengineer*, 74, 58-60, 1992.

26. Coral G, Arikan B, Unaldi MN, Guvenmez H: Some properties of crude carboxymethyl cellulase of *Aspergillus niger* Z10 wild-type strain. *Turk J Biol*, 26, 209-213, 2002.

27. Miller GL: Use of dinitrosalicylic acid for determination of reducing sugar. *Anal Chem*, 31 (3): 424-426, 1959.

28. Özcan N: Cloning and sequencing of a cellulase gene from *Fibrobacter succinogenes* SD35. *PhD Thesis*, Department of Molecular and Cell Biology, University of Aberdeen, Scotland, U.K., 1992.

29. Scott JH, Schekman R: Lyticase: Endoglucanase and protease activities that act together in yeast cell lysis. *J Bacteriol*, 142 (2): 414-423, 1980.

30. Ozduven ML, Kursun Onal Z, Koc F: The effects of bacterial inoculants and/or enzymes on the fermentation, aerobic stability and *in vitro* dry and organic matter digestibility characteristics of triticale silages. *Kafkas Univ Vet Fak Derg*, 16 (5): 751-756, 2010.

31. Fitzsimons A, Hols P, Jore J, Leer RJ, O'Connell M, Delcour J: Development of an amylolytic *Lactobacillus plantarum*, silage strain expressing the *Lactobacillus amylovorus* alpha-amylase gene. *Appl Environ Microbiol*, 60, 3529-3535, 1994.

32. Hols P, Ferain T, Garmyn D, Bernard N, Delcour J: Use of homologous expression-secretion signals and vector-free stable chromosomal integration in engineering of *Lactobacillus plantarum* for alpha-amylase and levanase expression. *Appl Environ Microbiol*, 60 (5): 1401-1413, 1994.

33. Jones S, Warner PJ: Cloning and expression of alpha-amylase from *Bacillus amyloliquefaciens* in a stable plasmid vector in *Lactobacillus plantarum*. *Lett Appl Microbiol*, 11 (4): 214-219, 1990.

34. Scheirlinck T, Mahillon J, Joos H, Dhase P, Michiels F: Integration and expression of alpha-amylase and endoglucanase genes in the *Lactobacillus plantarum* chromosome. *Appl Environ Microbiol*, 55 (9): 2130-2137, 1989.

35. Scheirlinck T, Meutter JD, Arnaut G, Joss H, Claeysens M, Michiels F: Cloning and expression of cellulase and xylanase genes in *Lactobacillus plantarum*. *Appl Microbiol Biotechnol*, 33 (5): 534-541, 1990.

36. Rossi F, Rudella A, Marzotto M, Dellaglio F: Vector-free cloning of a bacterial endo-1,4- β -glucanase in *Lactobacillus plantarum* and its effect on the acidifying activity in silage: Use of recombinant cellulolytic *Lactobacillus plantarum* as silage inoculant. *Antonie van Leeuwenhoek*, 80 (2): 139-147, 2001.

37. Wanker E, Leer RJ, Pouwels PH, Schwab H: Expression of *Bacillus subtilis* levanase gene in *Lactobacillus plantarum* and *Lactobacillus casei*. *Appl Microbiol Biotechnol*, 43 (2): 297-303, 1995.

38. Halbmayer E, Mathiesen G, Nguyen TH, Maischberger T, Peterbauer CK, Eijssink VG, Haltrich D: High-level expression of recombinant beta-galactosidases in *Lactobacillus plantarum* and *Lactobacillus sakei* using a Sakacin P-based expression system. *J Agric Food Chem*, 56 (12): 4710-4719, 2008.

39. Brurberg MB, Haandrikman AJ, Leenhouts KJ, Venema G, Nes IF: Expression of a chitinase gene from *Serratia marcescens* in *Lactococcus lactis* and *Lactobacillus plantarum*. *Appl Microbiol Biotechnol*, 42 (1): 108-115, 1994.

40. Cheong YH, Kim CY, Chun HJ, Moon BC, Park HC, Kim JK, Lee S, Han C, Lee SY, Cho MJ: Molecular cloning of a soybean class III beta-1,3-glucanase gene that is regulated both developmentally and in response to pathogen infection. *Plant Sci*, 154 (1): 71-81, 2000.

41. Tian SP, Yao EJ, Deng X, Xu XB, Qin GZ, Chan ZL: Characterization and expression of β -1,3-glucanase genes in jujube fruit induced by the microbial biocontrol agent *Cryptococcus laurentii*. *Phytopathology*, 97 (3): 260-268, 2007.

42. Yamaguchi T, Nakayama K, Hayashi T, Tanaka Y, Koike S: Molecular cloning and characterization of a novel β -1,3-glucanase gene from rice. *Biosci Biotechnol Biochem*, 66 (6): 1403-1406, 2002.

43. Ferrer P, Hedegaard L, Halkier T, Diers I, Savva D, Asenjo JA: Molecular cloning of a lytic beta-1,3-glucanase gene from *Oerskovia xanthineolytica* LLG109. A beta-1,3-glucanase able to selectively permeabilize the yeast cell wall. *Ann NY Acad Sci*, 782, 555-565, 1996.

The Effects of D-Ala², D-Leu⁵-Enkephalin (DADLE)'s Continuous Perfusion on Erythrogram and Biochemical Profile in Rats ^[1]

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Summary

In this experimental study it was aimed to investigate the effects of chronic continuous DADLE administration on erythrogram and the biochemical profile (ALP, ALT, AST, LDH, glucose, urea, creatinine, albumin, total protein). The study was carried on 48 adult male Wistar rats. DADLE was administered via osmotic mini-pumps, implanted subcutaneously. After the 28-day treatment, plasma glucose levels increased and urea levels decreased significantly in experimental group animals compared to the control group. Differences between the groups were not found to be statistically significant with respect to the other investigated parameters. So we conclude that chronic continuous administration of DADLE via osmotic mini-pumps did not exert any detrimental effect on erythrogram and the biochemical profile in rats.

Keywords: DADLE, Opioid, Osmotic pump, Erythrogram, Biochemical profile

D-Ala², D-Leu⁵-Enkephalin (DADLE)'in Sıçanlarda Sürekli Perfüzyonunun Eritrogram ve Biyokimyasal Profile Etkisi

Özet

Bu deneysel çalışmada DADLE'nin kronik sürekli uygulamasının bazı eritrositer parametreler (eritrogram) ve biyokimyasal profil (ALP, ALT, AST, LDH, glikoz, üre, kreatinin, albümin, total protein) üzerine olan etkilerinin araştırılması amaçlanmıştır. Çalışmada 48 adet yetişkin erkek Wistar sıçan kullanılmıştır. DADLE deri altına implante edilen ozmotik mini pompalar ile uygulanmıştır. Yirmisekiz günlük uygulama sonrası deney grubu hayvanlarının plazma glikoz düzeyleri artarken üre düzeyleri düşmüş ve bu değişikliklerin kontrol grubu ile karşılaştırıldığında istatistiksel önemde olduğu görülmüştür. İncelenen diğer parametreler açısından gruplar arasındaki farklar istatistiksel olarak önemli bulunmamıştır. Bu nedenle DADLE'nin ozmotik mini pompalar ile kronik ve sürekli uygulanmasının sıçanlarda eritrogram ve biyokimyasal profil açısından herhangi bir zararlı etki oluşturmadığı sonucuna varılmıştır.

Anahtar sözcükler: DADLE, Opioid, Ozmotik pompa, Eritrogram, Biyokimyasal profil

INTRODUCTION

Hibernation is an adaptive and protective strategy for a group of animal species and makes it possible for them to survive extended periods of food deprivation and extreme ambient conditions ^[1,2]. Animals in the hibernating state can tolerate body temperatures close to zero ^[3] and excessively decreased blood flow to vital organs ^[4,5].

Dawe and Spurrier ^[6] showed that blood from hibernating animals can induce hibernation when transfused into active euthermic individuals and suggested

for the first time the existence of a triggering substance in the blood. Furthermore they showed that this hibernation induction trigger (HIT) is not species-specific ^[7]. Later it has been found that HIT is present in the albumin fraction of plasma ^[8] and opioid receptors are involved in the induction of hibernation ^[9]. The synthetic peptide DADLE (D-Ala²,D-Leu⁵-Enkephalin; C₂₉H₃₉N₅O₇) is a stable analogue of the endogenous delta opioid enkephalin ^[10,11]. It passes blood-brain barrier after systemic application ^[12] and has a selective agonistic



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property on delta-opioid receptors [10]. DADLE has been found to induce hibernation in hibernant animals after continuous infusion [13].

The fascinating tissue protective adaptation seen during hibernation attracted substantial research interest on HIT and DADLE, so a wide range of studies have been conducted investigating the potential protective efficacy of them [4,14]. Indeed it has been demonstrated that HIT and DADLE prolong the survival of isolated organs (heart and liver) in a multiorgan preservation preparation almost 6-fold, which could have crucial implications for organ transplantation research [15,16]. Summers et al. [17] found that DADLE treatment improves hemodynamic stability and prolongs survival in an animal model of severe hemorrhagic shock. Another *in vivo* study showed that parenteral administration of DADLE protects the liver of rats against ischemia reperfusion injury [18]. Further studies pointed out the protective efficacy of DADLE on the nervous system [10,11,19]. Recently, Borlongan et al. [20] showed that DADLE pre-treatment, administered parenterally, prevents cell death processes and behavioral symptoms caused by experimentally induced stroke in rats.

The aforementioned results together manifest that DADLE has substantial protective efficacy in various cells and tissues and may be regarded as a promising novel therapeutic agent. Nevertheless it is known that prevalence of anemia is highly correlated with opioid use [21], and it has been reported that opioids cause membrane alterations of red blood cells [22]. Besides activation of different opioid receptor subtypes may exert different effects on target cells [23]. To the best of our knowledge no study has so far investigated the side effects of DADLE on erythrogram or the biochemical profile. So the aim of the present study was to investigate the potential side effects of long-term DADLE infusion on some erythrocyte and biochemical parameters.

MATERIAL and METHODS

Animals

Male Wistar albino rats weighing 250-275 g were used for the study. They were randomly allocated to 2 groups, each consisting of 24 rats, as control and experimental groups. All animals were housed in polycarbonate cages with wood shavings bedding in a climate-controlled animal room with 12/12 h light/dark cycle. They were fed standard laboratory chow and tap water *ad libitum*. All experimental procedures were approved by Istanbul University Veterinary Faculty's Animal Experiments Local Ethics Committee (Approval no. 2003/65)

DADLE Application

Rats were anesthetized using xylazine/ketamine (10/75 mg/kg). Osmotic mini pumps (Alzet, USA) containing DADLE (Sigma-Aldrich, Germany) and physiologic saline solution

were implanted subcutaneously into the interscapular region of experimental and control group animals, respectively. Experimental group animals received approximately 2.5 µl (±10%) of DADLE solution (0.5%) per hour, depending on the mean pumping rate of osmotic pumps during 28-day experimental period.

Samples and Tests

At the end of the experimental period all animals were exsanguinated via cardiac puncture under general anesthesia with xylazine/ketamine (10/75 mg/kg, Bayer, Turkey). Blood samples were collected into dry and EDTA tubes. Erythrogram parameters were determined using anticoagulated blood samples by using an electronic cell counter (Medonic 570, Boule Medical, Sweden). Sera were obtained from blood samples collected into dry tubes and were used for the determination of biochemical profile including following parameters: alkaline phosphatase (ALP), alanine aminotransferase (ALT), aspartate aminotransferase (AST), glucose (GLU), creatinine (CREA), lactate dehydrogenase (LDH), albumin (ALB), total protein (TP) and urea (URE) using an automatic biochemical analyzer (Abbott Architect, Abbott Laboratories, USA).

Statistical Analysis

All statistical analyses were performed with statistical software SPSS (version 11.5.2.1) using the independent samples t-test. All results are reported as means ± SEM. Significance was accepted at $P < 0.05$ level.

RESULTS

Hematological parameters are shown in Table 1. There were no statistically significant differences between groups.

The data on biochemical parameters are shown in Table 2. Glucose level increased and urea level decreased in DADLE treated group compared to the control group, and both differences were found to be statistically significant ($P < 0.05$). There were no statistically significant differences regarding other parameters.

Table 1. Hematological parameters

Tablo 1. Hematolojik parametreler

Parameter	Control (n=24)	DADLE (n=24)
Red blood cells (10 ⁶ /mm ³)	7.91±0.11	8.08±0.09
Hemoglobin (g/dl)	12.27±0.16	12.44±0.12
Hematocrit (%)	40.98±0.54	41.49±0.40
MCH (pg)	15.52±0.15	15.43±0.12
MCHC (g/dl)	29.96±0.11	30.03±0.08
MCV (fl)	51.81±0.42	51.40±0.41
RDW (%)	21.62±0.31	21.29±0.32

MCH: mean corpuscular hemoglobin; **MCHC:** mean corpuscular hemoglobin concentration; **MCV:** mean corpuscular volume; **RDW:** red blood cell distribution width

Table 2. Biochemical parameters**Table 2.** Biyokimyasal parametreler

Parameter	Control (n=24)	DADLE (n=24)
ALP (U/L)	387.61±14.63	409.13±22.40
ALT (U/L)	79.17±3.31	84.75±3.01
AST (U/L)	154.17±12.07	164.83±16.59
LDH (U/L)	362.86±9.96	392.29±16.65
Urea (mg/dl)	37.57±0.74*	34.54±0.88*
Albumin (g/dl)	3.19±0.05	3.17±0.03
Glucose (mg/dl)	161.65±4.07*	178.71±4.42*
Creatinine (mg/dl)	0.40±0.01	0.40±0.01
Total protein (g/dl)	6.92±0.06	6.95±0.07

ALP: alkaline phosphatase; ALT: alanine aminotransferase; AST: aspartate aminotransferase; LDH: lactate dehydrogenase; * Differences between the means of control and experimental groups are statistically significant ($P<0.05$)

DISCUSSION

To the best of our knowledge there are no studies examining the effects of long-term DADLE administration on erythrogram. In the present study 28-day continuous infusion of DADLE did not reveal any significant effect on the investigated parameters related to erythrocytes. Although studies focusing on the side effects of DADLE on hematological parameters are lacking, the effects of other opioids such as morphine on erythrocytes and related parameters have been investigated in various studies. Nie et al.^[24] found that acute and chronic *in vivo* and *in vitro* application of morphine, a mu-opioid receptor agonist, increased the osmotic fragility of red blood cells and shortened their lifetime by affecting the cell membrane. Zeiger et al.^[22] explained the high anemia prevalence among chronic opioid users with increasing levels of opioid receptors on red blood cells due to chronic opioid usage and subsequently increasing deformability of erythrocytes. Even though there is evidence indicating the presence of kappa- and mu-opioid receptors on the erythrocyte membrane^[22,25], we could not encounter any finding about the delta opioid receptors in the literature. The abovementioned detrimental effects of opioids on erythrocytes were attributed both to the ligand-receptor interactions^[22] and to the direct effect of opioid on erythrocyte membrane^[24]. So the absence of any effect of DADLE on red blood cells and related parameters can be due to the lack of delta opioid receptors on erythrocyte membrane or could be interpreted that chronic DADLE administration at this dose does not have any side effects on erythrocyte membrane directly.

Because of its crucial role in biotransformation and metabolism of most opioids as well as various other drugs, liver is predisposed to toxic injury, especially in chronic exposure to these drugs^[26-28]. Indeed it has been reported that parenteral administration of various opioids caused

hepatic damage indicated by increased liver enzymes^[29]. However to the best of our knowledge there are no studies investigating the adverse effects of chronic DADLE treatment on liver. Notwithstanding with the reports about detrimental effects of opioids on liver, Yamanouchi et al.^[18] stated that acute DADLE treatment exerted a hepatoprotective effect against acute liver damage and decreased serum ALT level in rats. In the present study chronic DADLE treatment via osmotic mini-pumps showed neither a beneficial nor a detrimental effect on liver enzymes.

Lactate dehydrogenase is a stable cytosolic enzyme present in a wide variety of mammalian tissues and cells, and leaks out of the cells into the bloodstream as a result of cell membrane damage^[30]. The increase of LDH level in extracellular fluid (e.g., plasma, serum) is a sensitive but nonspecific marker of cell and tissue damage including liver, kidney, heart and muscle^[31,32]. There are various *in vitro* studies in literature, investigating the effects of DADLE on several tissues and cells and it has been reported that DADLE exerted protective effect on these tissues and cells indicated by a decrease of LDH level^[33-36]. Dalargin (D-Ala²,Leu⁵,Arg⁶-enkephalin), another delta opioid receptor (DOR) agonist similar to DADLE, has been found to have a decreasing effect on LDH level in a traumatic shock model in rats^[37]. Atıcı et al.^[28] reported that long-term opioid treatment caused hepatic and renal damage in rats as indicated by a serum LDH increase. Nevertheless we could not find any reports about the *in vivo* effects of chronic DADLE treatment on LDH level. In the present study long-term DADLE treatment did not alter the plasma LDH activity, and this finding can be interpreted as an absence of detrimental effects on tissues including liver and kidney.

There was no significant difference between the creatinine levels of experimental and control groups in the present study. Besides, plasma urea level of DADLE treated animals was significantly ($P<0.05$) lower than that of the control group animals. Nevertheless both parameters were in normal range. Since the DADLE treatment did not alter the creatinine levels of both groups, it can be argued that the glomerular function was not altered by long-term treatment. So the aforementioned decrease in urea level of the experimental group can be based either on the decrease of protein catabolism or on an impairment of urea synthesis. The first possibility of altered protein metabolism by DADLE can be eliminated considering that there was no difference between plasma albumin and total protein levels of groups, and both parameters were in normal range. In this context it can be suggested that the decrease in the urea level of the experimental group is based on its impaired synthesis in liver. However we could not find any data about the effects of long-term DADLE treatment on urea synthesis in the literature to compare and discuss this possibility.

A wide range of studies has been conducted to investigate the effects of opioids on glucose homeostasis^[38-44].

It has been known for several decades that parenterally administered morphine causes hyperglycemia [45]. Bailey and Flatt [42] reported that mu receptor agonist DAMGO (D-Ala², N-Me-Phe⁴, Gly⁵-ol-Enkephalin) and delta receptor agonist DADLE dose dependently increased plasma glucose levels in rats. Beta endorphin, which exhibits high affinity for delta opioid receptors [46], has been found to significantly increase plasma glucose level [47]. Nonetheless Gunion et al. [39] reported that only mu-opioid receptor stimulation resulted in hyperglycemia in rats whereas delta opioid receptor sub-type was ineffective. In our present study long-term DADLE treatment produced a significant ($P < 0.05$) hyperglycemia in the experimental group. This increasing effect of DADLE on plasma glucose level may be due to its interaction with delta opioid receptors as suggested before [40-42].

In summary 28-day continuous infusion of DADLE via osmotic mini-pumps in our present study did not cause any significant alterations on erythrocyte parameters and biochemical profile except for a decrease in the plasma urea level and an increase in the glucose level. The finding of hyperglycemia is in accordance with previous data. Even though there is no relevant data in the literature to compare and discuss the significant decrease in the plasma urea level that we have observed, the levels of experimental and control groups were in normal range. Considering that the inspected biochemical parameters related to liver and kidney functions and the erythrocyte parameters were unaffected by DADLE, it would be reasonable to suggest that DADLE does not have any major detrimental effects at this dose level. Because of its promising tissue protective effects, DADLE deserves further study as a therapeutic agent, and our results could contribute to the further evaluation of chronic effects of DADLE treatment on various tissues, organs and systems.

REFERENCES

1. Carey HV, Andrews MT, Martin SL: Mammalian hibernation: cellular and molecular responses to depressed metabolism and low temperature. *Physiol Rev*, 83 (4): 1153-1181, 2003.
2. Lyman CP, Chatfield PO: Physiology of hibernation in mammals. *Physiol Rev*, 35 (2): 403-425, 1955.
3. Barnes BM: Freeze avoidance in a mammal: body temperatures below 0 degree C in an Arctic hibernator. *Science*, 244 (4912): 1593-1595, 1989.
4. Borlongan CV, Wang Y, Su TP: Delta opioid peptide (D-Ala², D-Leu⁵) enkephalin: Linking hibernation and neuroprotection. *Front Biosci*, 9, 3392-3398, 2004.
5. Drew KL, Rice ME, Kuhn TB, Smith MA: Neuroprotective adaptations in hibernation: Therapeutic implications for ischemia-reperfusion, traumatic brain injury and neurodegenerative diseases. *Free Radic Biol Med*, 31 (5): 563-573, 2001.
6. Dawe AR, Spurrier WA: Hibernation induced in ground squirrels by blood transfusion. *Science*, 163 (3864): 298-299, 1969.
7. Dawe AR, Spurrier WA, Armour JA: Summer hibernation induced by cryogenically preserved blood "trigger". *Science*, 168 (3930): 497-498, 1970.
8. Oeltgen PR, Bergmann LC, Spurrier WA, Jones SB: Isolation of a hibernation inducing trigger(s) from the plasma of hibernating woodchucks. *Prep Biochem*, 8 (2-3): 171-188, 1978.
9. Oeltgen PR, Walsh JW, Hamann SR, Randall DC, Spurrier WA, Myers RD: Hibernation "trigger": Opioid-like inhibitory action on brain function of the monkey. *Pharmacol Biochem Behav*, 17 (6): 1271-1274, 1982.
10. Hayashi T, Hirata H, Asanuma M, Ladenheim B, Tsao LI, Cadet JL, Su TP: Delta opioid peptide [D-Ala², D-Leu⁵]enkephalin causes a near complete blockade of the neuronal damage caused by a single high dose of methamphetamine: Examining the role of p53. *Synapse*, 39 (4): 305-312, 2001.
11. Tsao LI, Cadet JL, Su TP: Reversal by [D-Ala²,D-Leu⁵]enkephalin of the dopamine transporter loss caused by methamphetamine. *Eur J Pharmacol*, 372 (3): R5-7, 1999.
12. Banks WA, Kastin AJ: Peptide transport systems for opiates across the blood-brain barrier. *Am J Physiol*, 259 (1 Pt 1): E1-10, 1990.
13. Oeltgen PR, Nilekani SP, Nuchols PA, Spurrier WA, Su TP: Further studies on opioids and hibernation: Delta opioid receptor ligand selectively induced hibernation in summer-active ground squirrels. *Life Sci*, 43 (19): 1565-1574, 1988.
14. Su TP: Delta opioid peptide[D-Ala²,D-Leu⁵]enkephalin promotes cell survival. *J Biomed Sci*, 7 (3): 195-199, 2000.
15. Chien S, Oeltgen PR, Diana JN, Shi X, Nilekani SP, Salley R: Two-day preservation of major organs with autoperfusion multiorgan preparation and hibernation induction trigger. A preliminary report. *J Thorac Cardiovasc Surg*, 102 (2): 224-234, 1991.
16. Chien S, Oeltgen PR, Diana JN, Salley RK, Su TP: Extension of tissue survival time in multiorgan block preparation with a delta opioid DADLE ([D-Ala², D-Leu⁵]enkephalin). *J Thorac Cardiovasc Surg*, 107 (3): 964-967, 1994.
17. Summers RL, Li Z, Hildebrandt D: Effect of a delta receptor agonist on duration of survival during hemorrhagic shock. *Acad Emerg Med*, 10 (6): 587-593, 2003.
18. Yamanouchi K, Yanaga K, Okudaira S, Eguchi S, Furui J, Kanematsu T: [D-Ala², D-Leu⁵] enkephalin (DADLE) protects liver against ischemia-reperfusion injury in the rat. *J Surg Res*, 114 (1): 305-312, 2003.
19. Hayashi T, Su TP: Chronic [d-Ala², d-Leu⁵]enkephalin treatment increases the nerve growth factor in adult mouse brain. *Eur J Pharmacol*, 464 (2-3): 237-239, 2003.
20. Borlongan CV, Hayashi T, Oeltgen PR, Su TP, Wang Y: Hibernation-like state induced by an opioid peptide protects against experimental stroke. *BMC Biol*, 7, 31, 2009.
21. Brown LS, Hickson MJ, Ajuluchukwu DC, Bailey J: Medical disorders in a cohort of New York city drug abusers: Much more than HIV disease. *J Addict Dis*, 12 (4): 11-27, 1993.
22. Zeiger AR, Patkar AA, Fitzgerald R, Lundy A, Ballas SK, Weinstein SP: Changes in mu opioid receptors and rheological properties of erythrocytes among opioid abusers. *Addict Biol*, 7 (2): 207-217, 2002.
23. Kamiloglu NN, Beytut E, Özsar S: Opioid peptidiler ve bir opioid peptid agonisti olan naloxon'un üreme üzerine etkileri. *Kafkas Univ Vet Fak Derg*, 12 (1): 85-90, 2006.
24. Nie X, Wen ZY, Yan ZY, Huang L, Sun D, Cheng B: Effects of morphine on rheological properties of rat red blood cells. *Clin Hemorheol Microcirc*, 22 (3): 189-195, 2000.
25. Yamasaki Y, Way EL: Inhibition of Ca⁺⁺-ATPase of rat erythrocyte membranes by k-opioid agonists. *Neuropeptides*, 5 (4-6): 359-362, 1985.
26. Tegeder I, Lötsch J, Geisslinger G: Pharmacokinetics of opioids in liver disease. *Clin Pharmacokinet*, 37 (1): 17-40, 1999.
27. Hanna M: The effects of liver impairment on opioids used to relieve pain in cancer patients. *Palliat Med* 25 (5): 604-605, 2011.
28. Atici S, Cinel I, Cinel L, Doruk N, Eskandari G, Oral U: Liver and kidney toxicity in chronic use of opioids: An experimental long term treatment model. *J Biosci*, 30 (2): 245-252, 2005.
29. Needham WP, Shuster L, Kanel GC, Thompson ML: Liver damage from narcotics in mice. *Toxicol Appl Pharmacol*, 58 (2): 157-170, 1981.
30. Parhamifar L, Andersen H, Moghimi SM: Lactate dehydrogenase

assay for assessment of polycation cytotoxicity. In, Ogris M, Oupicky D (Eds): *Methods in Molecular Biology*. pp.13-22, Humana Press, Totowa, 2013.

31. **O'Brien PJ, Watterson CL:** General enzymology. In, Evans GO (Ed): *Animal Clinical Chemistry: A Practical Handbook for Toxicologists and Biomedical Researchers*. 2nd ed., pp.27-28, CRC PressINC, Florida, 2009.
32. **Drent M, Cobben NA. M, Henderson RF, Wouters EFM, van Dieijen-Visser M:** Usefulness of lactate dehydrogenase and its isoenzymes as indicators of lung damage or inflammation. *Eur Respir J*, 9 (8): 1736-1742, 1996.
33. **Hayashi T, Tsao LI, Su TP:** Antiapoptotic and cytotoxic properties of delta opioid peptide [D-Ala(2),D-Leu(5)]enkephalin in PC12 cells. *Synapse*, 43 (1): 86-94, 2002.
34. **Hong SS, Gibney GT, Esquilin M, Yu J, Xia Y:** Effect of protein kinases on lactate dehydrogenase activity in cortical neurons during hypoxia. *Brain Res*, 1009 (1-2): 195-202, 2004.
35. **Seymour EM, Wu S-YJ, Kovach MA, Romano MA, Traynor JR, Claycomb WC, Bolling SF:** HL-1 myocytes exhibit PKC and KATP channel-dependent delta opioid preconditioning. *J Surg Res*, 114 (2): 187-194, 2003.
36. **Zhang J, Haddad GG, Xia Y:** Delta-, but not mu- and kappa-, opioid receptor activation protects neocortical neurons from glutamate-induced excitotoxic injury. *Brain Res*, 885 (2): 143-153, 2000.
37. **Mikhailova NN, Slepishkin VD:** Effectiveness of the use of synthetic opioid peptide in rats with traumatic shock. *Patol Fiziol Eksp Ter*, 4, 27-29, 1995.
38. **Ramabadran K, Bansinath M:** Glucose homeostasis and endogenous opioid peptides. *Int J Clin Pharmacol Ther Toxicol*, 28 (3): 89-98, 1990.
39. **Gunion MW, Rosenthal MJ, Morley JE, Miller S, Zib B, Butler B, Moore RD:** Mu-receptor mediates elevated glucose and corticosterone after third ventricle injection of opioid peptides. *Am J Physiol*, 261 (1 Pt 2): R70-81, 1991.
40. **Green IC, Perrin D, Pedley KC, Leslie RD, Pyke DA:** Effect of enkephalins and morphine on insulin secretion from isolated rat islets. *Diabetologia*, 19 (2): 158-161, 1980.
41. **Hermansen K:** Enkephalins and the secretion of pancreatic somatostatin and insulin in the dog: Studies *in vitro*. *Endocrinology*, 113 (3): 1149-1154, 1983.
42. **Bailey CJ, Flatt PR:** Increased responsiveness to glucoregulatory effect of opiates in obese-diabetic ob/ob mice. *Diabetologia*, 30 (1): 33-37, 1987.
43. **Evans AA, Tunncliffe G, Knights P, Bailey CJ, Smith ME:** Delta opioid receptors mediate glucose uptake in skeletal muscles of lean and obese-diabetic (ob/ob) mice. *Metabolism*, 50 (12): 1402-1408, 2001.
44. **Szeto HH, Soong Y, Wu DL, Cheng PY:** Opioid modulation of fetal glucose homeostasis: Role of receptor subtypes. *J Pharmacol Exp Ther*, 275 (1): 334-339, 1995.
45. **Feldberg W, Gupta KP:** Morphine hyperglycaemia. *J Physiol*, 238 (3): 487-502, 1974.
46. **Cox BM:** Endogenous opioid peptides: A guide to structures and terminology. *Life Sci*, 31 (16-17): 1645-1658, 1982.
47. **van Loon GH, Appel NM:** Beta-endorphin-induced hyperglycemia is mediated by increased central sympathetic outflow to adrenal medulla. *Brain Res*, 204 (1): 236-241, 1981.

Parasitic Aortitis due to *Onchocerca armillata* in Slaughtered Cattle in the Southeastern Region of Turkey

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Summary

In this study, total of 400 aorta suspected with parasitic aortitis was investigated between the years 2011-2012 in slaughtered cattle in the Southeastern of Turkey. Particularly arcus aorta and its surroundings were investigated. Macroscopically 32 of 400 (8%), both macroscopically and microscopically 43 (10.75%) cattle were diagnosed as aortic onchocercosis. The aim of this study is to investigate prevalence and pathology of parasitic aortitis in detail caused by *Onchocerca armillata* in the Southeastern region of Turkey.

Keywords: Cattle, *Onchocerca armillata*, Parasitic aortitis

Türkiye'nin Güneydoğusu'nda Mezbahalarda Kesilen Sığırlarda *Onchocerca armillata*'nın Neden Olduğu Parazitik Aortitis

Özet

Bu çalışmada, Türkiye'nin Güneydoğusu'nda 2011-2012 yılları arasında mezbahalarda kesilen 400 sığır aortasında gözlenen parazitik aortitis olguları incelendi. Aortaların alınıp yeri arcus aorta ve çevresi idi. İncelemeler sonunda 400 hayvandan 32 tanesinde (%8) makroskopik, 43 tanesinde ise (%10.75) hem makroskopik, hem de mikroskopik olarak aortik onkoserkozis tespit edildi. Bu çalışmanın amacı Türkiye'nin Güneydoğusu'nda *Onchocerca armillata* adlı parazitin neden olduğu parazitik aortitis olgularının prevalansı ve patolojisinin ayrıntılı bir biçimde incelenmesidir.

Anahtar sözcükler: Sığır, *Onchocerca armillata*, Parazitik aortitis

INTRODUCTION

Onchocerca armillata a parasite found in the wall of the aorta of cattle ^[1,2] buffalo ^[3], sheep ^[3-5], goat ^[6] and camels ^[7]. *Onchocerca armillata* is mostly observed in the south of the Asian regions and African continent closer to the equator ^[8-10]. In recent years, onchocercosis was also reported in the Tanzania ^[11], Cameroon ^[12], Venezuela ^[13] and in the Greece ^[14]. *Onchocerca* spp. have been reported at skin, tendons, aponeurosis, testicular and breast tissues as well ^[8]. The life cycle and vectors of the *Onchocerca armillata* are unknown. However, black flies and mosquitoes are thought to be vectors ^[12-14]. Onchocercosis often do not show any significant clinical symptoms. Sometimes the bulls show neural symptoms and sometimes microfilaria localized at

ocular regions causes recurrent ophthalmitis ^[8,14]. *Onchocerca armillata* mostly settles in the arcus aorta region. However, can also be found in brachiocephalic truncus, cervical and brachial arteries and the abdominal aorta up to the iliac bifurcation regions ^[1,8]. Affected aortic intima layers have undulant appearance. Numerous tunnels and nodules are found in the media. These nodules show protrusions towards the intima and media layers. Survived and matured microfilaria can pass the circulation ^[2]. Parasitic tunnels are filled with a thin connective tissue without inflammatory cells. Granulomatous nodules present around the dead, degenerate or calcified parasite larvae, contain dominantly eosinophil leukocytes ^[8,11,15,16]. Granulomatous foci in the



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vessel wall can be resorbed over time. Aneurysms can be observed in very old animals due to calcified nodules^[1,15,17]. In recent years, antiparasitic disinfestation decreased the frequency of the onchocercosis^[13].

The aim of this study is to examine the prevalence and pathology of parasitic aortitis that caused by *Onchocerca armillata* in the Southeastern region of Turkey.

MATERIAL and METHODS

This study was conducted in Sanliurfa, Diyarbakir, Hatay and Gaziantep provinces of Turkey between the years 2011-2012. A hundred sample from each province, from a total of 400 cattle aorta, was collected from slaughterhouses. The aortic samples were taken 2 cm above and below including arcus aorta and were fixed in 10% neutral buffered formalin. Fixed aorta lumens were opened and investigated. At suspected places of intima, adventitia or random regions of arcus aorta was trimmed parallel to the long axis of the vessel and passed through all the layers. Then, tissue samples embedded in paraffin by routine methods. Sections were cut 5-6 µm in thickness and were stained with hematoxylin and eosin (HE), Masson's trichrome and Von Kossa.

RESULTS

Macroscopical Findings

Macroscopically 32 of 400 (8%) cattle were diagnosed as aortic onchocercosis. The affected vessel walls, in general, thickened, crusty in consistency and lost flexibility. In a few cases the vessel walls were in appearance of rubber tube and their lumens were severely narrowed.

The intima, media and adventitial layers of the aorta were evaluated in more detail. Accordingly;

Tunica intima: Affected areas revealed tough and an irregular surface. Some of nodules were yellowish gray in color (Fig. 1-A). The nodules of varying sizes (0.5-4.0 cm), especially in areas close to the bifurcation, projecting to the vessel lumen, hard in consistency, bright and white surface, cross-sectional areas of calcification felt creaking (Fig. 1-B). Cut surfaces were dry and in caseous structure, which was cleaved easily and surrounded by a tough connective tissue to the size varies from lentils to nuts (Fig. 1-A,F). In some areas, upturned edges formed crater-like foci which causes shrinkage of the surface with the star-like connective tissue induration (Fig. 1-A). In cross-sections, caseous foci, easily cleaved and yellowish gray in color were also observed (Fig. 1-F).

Tunica media: The cross section of the affected areas had tough and an irregular surface. Thickening of the medial layer was noticed in areas where parasitic granulomas were observed. In some areas, dark red-brown old hemorrhage

areas hard in consistency were observed along with the long axis. Their cross-sections showed caseous and/or calcified parasite nodules and tunnels (Fig. 1-E,F).

Tunica adventitia: Parasitic nodule formations were observed rarely at the adventitial layer. These nodules bud-like, located sometimes single, sometimes combined with each other in groups of two to three. In this layer the diameter of the granulomas were quite large (the largest of the 4 cm) compared to the intimal layer. Granulomas that overflowing out of the vessel were hard in consistency, and often surrounded by a broad connective tissue (Fig. 1-C). In cross-sections the presence of large caseation and small amount of calcified areas were noticed (Fig. 1-D).

Histopathological Findings

Fourty three of 400 (10.75%) cattle were diagnosed as aortic onchocercosis both macroscopically and microscopically. Histopathologically, 11 of 43 (2.75%) parasitic aortitis were diagnosed with no macroscopic findings. In general, varying degrees of tissue reactions were investigated in the aorta. The majority of parasitic tunnels were located in the intima and media (Fig. 2-B). These tunnels contained dense haemorrhagia without inflammatory cell infiltration (Fig. 2-A). However, calcium deposits were noticed in some regions of these tunnels (Fig. 2-A,B,C). There are round or ellipsoidal cavities containing one or more intact parasitic sections, some with microfilariae. Worms and its structure surrounding thin cuticula appeared to reside within a round or ellipsoidal cavity, with a space between the worm section and the host-derived lining. Some parts of the parasitic cysts were limited by a thin connective tissue without any inflammatory cell infiltration (Fig. 2-B,C,D). A thin halo was found between larvae and the connective tissue capsule (Fig. 2-B,D). Muscle fibers around the parasitic cysts and tunnels were hyalinised. Masson's trichrome stain revealed presence of connective tissue at the cyst capsule. Some areas appeared basophilic with the H&E further stained by von Kossa revealed calcium deposits around and in the parasitic cysts and tunnels.

Details of inflammatory cell infiltration, calcification, sclerosis and parasitic granulomas are given in Table 1.

Inflammatory reactions were examined in 3 stages; acute, subacute, and chronic granulomatous aortitis.

Acute aortitis: Mostly started at adventitial vessels, sporadic eosinophils, neutrophils and macrophage infiltrations were spread in a linear style from intimal vessels to muscle bundles. In some areas, dilated blood vessels, rounded endothelial cells, inflammatory oedema and hyalinization of muscle bundles were detected around the parasitic cysts.

Subacute aortitis: A great number of eosinophils, a few number of macrophages, lymphocytes and plasma cell infiltrations were observed in the surroundings of medial and adventitial layers and that were moving towards to the

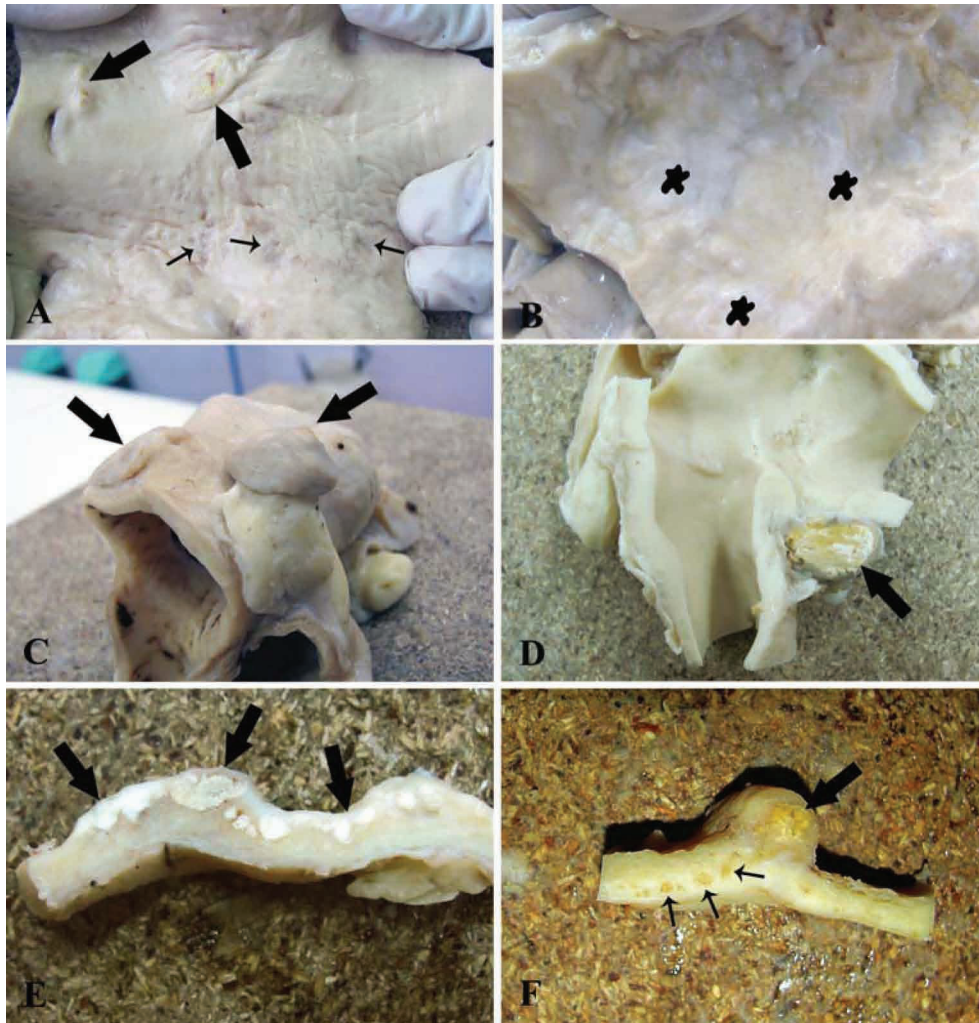


Fig 1. Macroscopic appearance of parasitic aortitis. A- Parasitic nodules in the lumen of the aorta (thick arrows), and areas of scar in appearance of star (thin arrows), B- Irregular and opaque appearance of calcified areas in the lumen of the aorta (stars), C- Parasitic granulomas, surrounded by tough connective tissue, extending outward the aorta serosa (arrows), D- Cross sections of parasitic granuloma in serosal surface (arrow), E- Calcium deposits along with long axis of intimal surface (arrows), F- Parasitic tunnels advancing up to intima, adventitia and serosa (thin arrows), demarcated caseified parasitic granuloma in the serosal surface (thick arrow)

Şekil 1. Parazitik aortitisin makroskopik görünümü. A- Aorta lümenine doğru çıkıntı yapan parazit nodülleri (kalın oklar) ve yıldız görünümündeki sikatriks alanları (ince oklar), B- Aorta lümeninde düzensiz ve opak görünümdeki kalsifikasyon alanları (yıldızlar), C- Aorta serozasından dışa doğru taşan etrafı sert bağ doku ile sarılı paraziter granulomlar (oklar), D- Serozal yüzeydeki paraziter granulomun kesit yüzü (ok), E- İntima yüzeyinde uzun eksen boyunca ilerleyen kalsiyum birikimleri (oklar), F- İntimadan adventisya ve serozaya kadar ilerleyen kazeifiye parazit tünelleri (ince oklar), serozada demarke durumdaki kazeifiye parazit granulomu (kalın ok)

dead and caseous parasitic cysts. This cellular infiltration was generally localised around the old or calcified tunnels (Fig. 2-F). Some areas showed new formation of blood vessels with presence of few fibroblasts.

Chronic granulomatous aortitis: The chronic granulomatous aortitis was the most common type of inflammation. Hyperchromatic and multinucleated giant cells were observed around the dead, degenerated or calcified parasitic cyst walls. A large number of eosinophilic infiltration was present between giant cells. This infiltration was supported by dense macrophages, lymphocytes, and plasma cells (Fig. 2-F). The outermost composed from granulation tissue that including fibroblasts, fibrocytes

and newly formed blood vessels. Muscle bundles were hyalinised (Fig 2-E). In some areas, inflammatory cell infiltration was decreased while sclerosis was increased. In eight cases, arteriosclerotic foci with presence of clefts at their centers composed of lymphocytes, macrophages, fibroblasts, and fibrocytes were found. Some cases did not show any inflammatory cell infiltration around the parasitic cysts (8 cases), some of them were calcified (28 cases) and some others revealed inflammatory cell infiltration (26 cases). On the other hand eosinophils and mononuclear cell infiltration were observed without any sign of parasitic cysts in eight cases. The inflammatory reaction against to cysts and calcified cysts formations was observed at the highest rate in the Diyarbakir province (Fig. 3).

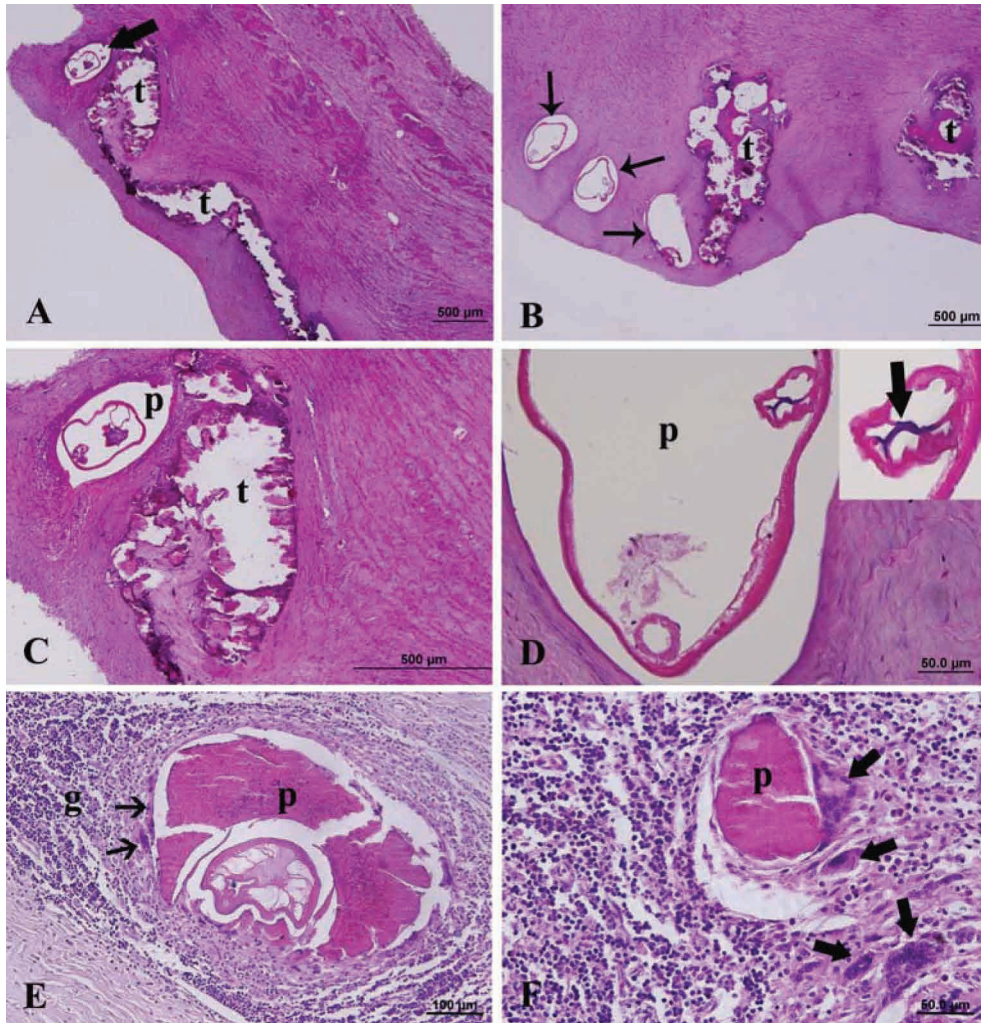


Fig 2. Microscopical aspects of parasitic aortitis. **A-** Calcified tunnels continue along with long axis of intimal surface (t). Parasitic cyst demarcated by a thin connective tissue without any inflammatory reaction (arrow), H&E X 40, **B-** Parasitic cysts surrounded by a thin connective tissue and halo in the intimal layer (arrows) and parasite tunnels (t) containing hyaline tissue and calcium deposits, H&E X 40, **C-** Calcified wide tunnel (t) progressing in different directions towards the layer of media and parasitic cyst (p) demarcated by the connective tissue, H&E X 100, **D-** An active parasitic cysts (p) surrounded by a large halo and a thin connective tissue, H&E X 100, the closer view of the mikroflaria (arrow, enlarged figure), **E-** Central part of parasitic granuloma is partly caseified in the medial layer, (p) surrounded by giant cells (arrows) and connective tissue, H&E X 400, **F-** Completely caseified parasite (p), surrounded by multinucleated giant cells (arrows), eosinophils and mononuclear cell infiltration, H&E X 400

Şekil 2. Parazitik aortitisin mikroskopik görünümü. **A-** İntimada uzun eksen boyunca devam eden kalsifiye olmuş tüneller (t). Etrafı ince bir bağ doku ile sınırlanmış yangı hücreleri bulundurmeyen paraziter kist (ok), H&E X 40, **B-** İntimada etraflarından ince bir bağ doku ve halo ile çevrili paraziter kistler (oklar) içerisinde hyalinize doku ve kalsiyum birikimleri bulunduran parazit tünelleri (t), H&E X 40, **C-** Media katmanına doğru değişik yönlerde ilerleyen kalsifiye olmuş geniş tunel (t) devamında etrafı bağdoku ile sınırlı parazit kisti (p), H&E X 100, **D-** Etrafı ince bir bağ doku ve geniş bir halo ile çevrili aktif parazit kisti (p), H&E X 100, içerisindeki aktif mikroflaria'nın yakından görünümü (ok, büyütülmüş resim), **E-** Media katmanında, merkezi kısmen kazeifiye olmuş, çevresinde dev hücreler (oklar) ve bağdoku ile sarılı paraziter granülom (p), H&E X 400, **F-** Tamamen kazeifiye parazit (p) etrafında çok çekirdekli dev hücreleri (oklar), eozinofil lökositler ve mononükleer hücre infiltrasyonları, H&E X 400

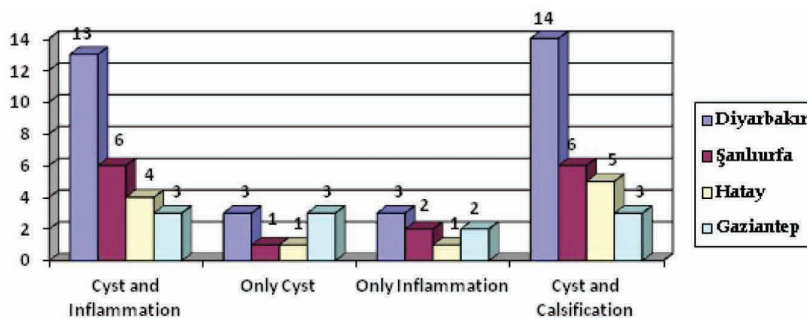


Fig 3. Evaluation of parasitic cysts according to the provinces

Şekil 3. Parazitik kistlerin illere göre değerlendirilmesi

Table 1. Histopathological aspects of parasitic aortitis due to *Oncocerca armillata***Tablo 1.** *Onchocerca armillata*'nın neden olduğu paraziter aortitisin histopatolojik değerlendirmesi

Number	Case Number	Inflammation			Sclerosis			Calcification			Parasitic Cyst		
		I	M	A	I	M	A	I	M	A	I	M	A
1	D-18	-	-	-	+	-	-	-	+	-	-	-	-
2	D-68	MEG	ME	M	+	-	-	+	-	-	+	+	-
3	D-23	M	-	-	+	-	-	+	-	-	+	-	-
4	D-58	-	-	ME	-	-	-	-	-	-	-	-	-
5	D-28	ME	M	ME	-	+	-	-	-	-	-	+	+
6	D-17	-	-	-	+	-	-	+	-	-	+	-	-
7	D-75	ME	ME	ME	+	+	-	+	+	-	+	-	-
8	D-7	-	M	M	-	-	-	-	-	-	-	-	-
9	D-12	-	-	M	-	-	+	+	-	-	-	-	+
10	D-69	-	MD	M	+	+	-	-	-	-	+	+	-
11	D-72	ME	ME	MEG	-	+	-	+	+	+	+	+	+
12	D-86	ME	-	MEG	-	+	+	-	+	+	-	+	+
13	D-67	M	M	-	+	-	-	+	-	-	+	-	-
14	D-4	ME	MG	M	+	+	+	+	+	-	+	+	-
15	D-3	MG	M	M	+	-	+	+	-	-	+	-	-
16	D-56	-	MG	M	-	+	+	-	+	-	-	+	-
17	D-39	-	M	M	-	-	-	-	-	-	-	-	-
18	D-21	-	M	M	+	-	+	+	+	-	+	+	-
19	D-29	ME	MEG	ME	+	+	-	-	+	-	-	+	-
20	D-44	ME	-	ME	+	-	-	+	+	-	+	+	-
21	D-30	-	M	M	-	-	-	-	-	-	-	-	+
22	S-10	-	-	ME	-	-	+	-	-	-	-	-	+
23	S-75	-	ME	ME	-	-	-	-	-	-	-	-	-
24	S-14	M	-	-	-	-	-	+	-	-	+	-	-
25	S-65	-	ME	M	-	-	-	-	-	-	-	-	-
26	S-64	M	MG	MG	-	+	+	-	+	-	-	+	+
27	S-21	MEG	MEG	MEG	+	+	+	+	+	+	+	+	+
28	S-90	MEG	MEG	MEG	+	+	+	+	+	+	+	+	+
29	S-51	M	-	M	+	-	+	+	-	-	+	-	+
30	S-17	-	ME	-	-	+	-	-	-	-	+	-	+
31	GA-52	-	-	-	+	-	-	+	-	-	+	-	-
32	GA-65	-	-	M	-	-	-	-	-	-	-	-	-
33	GA-1	ME	-	-	+	-	-	+	-	-	+	-	-
34	GA-27	-	ME	-	-	-	-	-	-	-	+	+	-
35	GA-78	-	-	-	-	-	-	-	-	-	-	-	+
36	GA-30	ME	ME	-	-	-	-	-	-	-	-	-	-
37	GA-18	ME	ME	-	-	-	-	-	-	-	+	-	-
38	GA-17	-	-	-	-	-	-	+	+	-	+	+	-
39	H-26	ME	ME	MEG	-	+	+	+	+	+	+	+	+
40	H-86	ME	-	-	+	+	-	+	+	-	+	+	-
41	H-36	-	-	-	+	-	-	+	-	-	+	-	-
42	H-23	M	M	-	+	-	-	+	-	-	+	-	-
43	H-48	ME	MEG	MEG	-	+	+	+	+	+	+	+	+

DISCUSSION

Aortic onchocercosis seen in different parts of the world [3,10,15,18]. Previous studies reported high incidence

of onchocercosis in the aorta (50-100%) [1,4,15]. Use of anti-helminthic drugs and pesticides decreased the presence of *Onchocerca* spp. in the both vectors and hosts [13]. In this study, 43 (10.75%) of 400 animals from four different

provinces showed the presence of aortic onchocercosis. This means that antihelminthic drugs were not effectively used in Turkey.

Recent studies showed Onchocercosis have still seen in different parts of the world ^[11,13,14,17]. Alibasoglu et al.^[1] reported that onchocercosis was seen in the southeastern provinces of Turkey in cattle, was not seen in western Anatolia provinces, and it was concluded that the disease was seen in tropical climates. However, according to recent study conducted by Beytut et al.^[6] the parasite was also seen at Kars province where the climate is severely cold than that of southern regions, also some other cases reported from Greece ^[14]. This means that Onchocercosis can also be seen in cold places. It is known that *Onchocerca* spp. is carried by black flies and mosquitoes ^[8,14]. As these flies can be found all over the world it is hard to establish a direct relationship with climate and the incidence of the disease.

Onchocerca armillata is a parasite originated from *Onchocerca* spp. It is known that flarial forms are found in bloodstream and adult forms are reside in around arcus orta. There is any report about parasitic aortitis caused by other *Onchocerca* species which is located around arcus aorta ^[1,8,10]. In this study, we described parasitic aortitis due to *Onchocerca armillata* around arcus aorta

In our country the prevalence and pathology of aortic onchocercosis have been only studied by Alibasoglu et al.^[1] and Beytut et al.^[6]. Alibasoglu et al.^[1] examined the aortas of animals around Cukurova and Western Anatolia regions and they seen intensive infestation in the Cukurova region. Materials of the present study were selected from the cities that are close together and on the same line in the Southeast provinces. Between these cities aortic onchocercosis was mostly seen in Diyarbakir province. This was followed by Sanliurfa, Gaziantep and Hatay provinces. According to Fig. 3; only cysts, the inflammatory reaction against to cysts and calcified cyst formations were occurred at the highest rate in the Diyarbakir province. Only the cyst formation and inflammation were observed in all the provinces with almost equal intensity. The data obtained from the present study indicates that antihelminthic and pesticide drugs are not effectively used in the Diyarbakir and Sanliurfa provinces.

Ogundipe et al.^[10] reported that in the parasitic cysts, live, dead and calcified tissues were located close to each other. In the present study the finding of degenerated or calcified cysts located close to each other were compatible with Ogundipe et al.^[10] and other studies ^[1,10,16]. Similarly, degenerated or calcified cysts were surrounded by the parasitic granulomas that contains giant cells, eosinophils, macrophages, lymphocytes and fibroblasts were also compatible with previous findings ^[10].

The absence of any inflammatory reaction around tunnels, and some parasitic cysts was remarkable. However, in some cases, these parasite tunnels were found to be

calcified. It was previously reported by many researchers that no inflammatory reaction occur against to live and vibrant forms of parasite ^[1,15,16]. Some of the information in the literature show that a substance released from live cuticula of parasite body prevents reaction against to the parasite ^[15,17].

With this study, the prevalence and pathology of *Onchocerca armillata* in cattle was examined in detail in the Southeastern region of Turkey.

REFERENCES

1. Alibasoglu M, Gölesuk K, Erturk E, Güler S: Türkiye'de sığırlarda görülen onchocerciasis olayları (*Onchocerca armillata* Railliet ve Henry 1909). *Ankara Üniv Vet Fak Derg*, 16, 50-60, 1969.
2. Wildenburg G, Kromer M, Buttner DW: Dependence of eosinophil granulocyte infiltration into nodules on the presence of microfilariae producing *Onchocerca volvulus*. *Parasitol Res*, 82, 117-124, 1996.
3. Bhatia BB: *Onchocerca armillata* Railliet and Henry 1909, a study of the infection in Indian sheep with remarks on its bovine hosts. *Ind Vet J*, 37, 394-397, 1960.
4. Chowdhury N, Chakraborty RL: On concurrent and early aortic Onchocerciasis and Spiroerciasis in domestic goats (*Capra hircus*). *Z Parasitenkunde*, 42, 207-212, 1973.
5. Gupta PP, Nagpal SK: Spontaneous aortic atherosclerosis in sheep. *Vet Pathol*, 16, 548-552, 1979.
6. Beytut E, Akca A, Bain O: Teat onchocercosis in cows with reference to prevalence, species involved and pathology. *Res Vet Sci*, 78, 45-51, 2005.
7. Oryan A, Valinezhad A, Bahrami S: Prevalence and pathology of camel filariasis in Iran. *Parasitol Res*, 103, 1125-1131, 2008.
8. Maxie MG, Youssef S: Diseases of the vascular system. In: Maxie MG, Robinson WF (Eds): Jubb, Kennedy and Palmer's Pathology of Domestic Animals. Vol. 3, pp. 91-92, Elsevier Science Publishing, Inc., USA 2007.
9. Neary JM, Trees AJ, Ekale DD, Tanya VN, Hetzel U, Makepeace BL: *Onchocerca armillata* contains the endosymbiotic bacterium *Wolbachia* and elicits a limited inflammatory response. *Vet Parasitol*, 174, 267-276, 2010.
10. Ogundipe GA, Ogunnirade AF, Akpavie SO: The prevalence, gross lesions and histopathology of aortic onchocerciasis in Nigerian cattle. *Vet Q*, 6, 85-89, 1984.
11. Mtei BJ, Sangha HJN: Aortic onchocercosis and elaeophorosis in traditional TSZ-cattle in Tabora (Tanzania): Prevalance and pathology. *Vet Parasitol*, 36, 165-170, 1990.
12. Achukwi MD, Harnett W, Renz A: *Onchocerca ochengi* transmission dynamics and the correlation of *O. ochengi* microfilaria density in cattle with the transmission potential. *Vet Res*, 31, 611-621, 2000.
13. Grillet ME, Villamizar NJ, Frontado HL, Cortez J, Escalona M, Botto C, Basanez MG: Vector competence of *Simulium oyapockense* s.l. and *S. incrustatum* for *Onchocerca volvulus*: Implications for ivermectin-based control in the Amazonian focus of human onchocerciasis, a multi-vector-host system. *Acta Trop*, 107, 80-89, 2008.
14. Otranto D, Torres FD, Papadopoulos E, Petric D, Cupina AI, Bain O: Tracking the vector of *Onchocerca lupi* in a rural area of Greece. *Emerg Infect Dis*, 18, 1196-1200, 2012.
15. Cheema AH, Ivochli B: Bovine onchocerciasis caused by *O. armillata* and *O. gutturosa*. *Vet Pathol*, 15, 495-505, 1978.
16. Specht S, Brattig N, Buttner M, Buttner DW: Criteria for the differentiation between young and old *Onchocerca volvulus* filariae. *Parasitol Res*, 105, 1531-1538, 2009.
17. Ertmann KD, Buttner DW: Immunohistological studies on *Onchocerca volvulus* paramyosin. *Parasitol Res*, 105, 1371-1374, 2009.
18. Zak F: Parasitic aortopathy due to *Onchocerca armillata* in cattle. *Path Microbiol*, 43,150, 1975.

The Effects of Dietary Rosemary (*Rosmarinus officinalis* L.) Oil Supplementation on Performance, Carcass Traits and Some Blood Parameters of Japanese Quail Under Heat Stressed Condition

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Summary

In this study, the effects of rosemary (*Rosmarinus officinalis* L.) oil supplementation to diet were investigated on performance, carcass traits and some blood parameters of Japanese quails exposed to a high ambient temperature of 34°C. A total of 180 fifteen-day-old quails were divided into 6 treatments consisting of 10 birds of 3 replicates. All groups were balanced according to initial live weight and gender. Birds were kept in wire cages in temperature-controlled room at either 22°C for 24 h/d (thermo neutral-TN) or 22°C for 16 h/d and 34°C (heat stress-HS) for 8 h/d (from 9:00 to 17:00) during the study. Trial was conducted as a 2x3 factorial arrangement. Birds were fed either a basal (control) diet (TN and HS) or the basal diet supplemented with 125 or 250 ppm of rosemary oil. The highest final live weight was observed in 250 ppm rosemary oil under the TN condition and the lowest was in control group of HS. HS condition affected negatively on quail live weight ($P<0.05$), but the effect of rosemary oil on this parameter was not significant ($P>0.05$). Live weight gain and feed intake were not significantly different among the treatment groups ($P>0.05$). Feed conversion ratio was better in rosemary oil groups than control groups in both environmental conditions ($P<0.01$). Heat stress deteriorated carcass yield ($P<0.01$). The highest hot and cold carcass yield (g/100 g of body weight) was observed in 250 ppm rosemary oil added group under the TN condition, but this difference did not significant. Birds kept in HS conditions had greater glucose level than hens kept in TN conditions ($P<0.01$). Rosemary oil decreased blood glucose level, especially in 250 ppm group of HS ($P<0.05$). Total, HDL, LDL cholesterol and triglyceride levels were not significantly different among the treatment groups ($P>0.05$). In conclusion, rosemary oil supplementation reduced the negative effects of heat stress. Rosemary oil could be considered as a potential natural feed additive, following further studies.

Keywords: Rosemary oil, Performance, Carcass traits, Blood parameters, Quail, Heat stress

Sıcaklık Stresi Altındaki Japon Bildircinlarında Karma Yeme İlave Edilen Biberiye (*Rosmarinus officinalis* L.) Yağının Performans, Karkas Özellikleri ve Bazı Kan Parametreleri Üzerine Etkileri

Özet

Bu çalışmada, karma yeme ilave edilen biberiye (*Rosmarinus officinalis* L.) yağının yüksek çevre sıcaklığına (34°C) maruz bırakılan Japon bildircinlerinde performans, karkas özellikleri ve bazı kan parametreleri üzerine etkileri araştırılmıştır. Toplam olarak 180 adet 15 günlük bildircin, her tekrerründe 10 bildircinin bulunduğu 3 tekrerrülü 6 gruba ayrılmıştır. Tüm gruplar başlangıç canlı ağırlıkları ve cinsiyet bakımından dengelenmiştir. Bildircinler, çalışma süresince sıcaklık kontrollü odalarda 22°C'de 24 saat/gün (termo nötral-TN) ve 22°C'de 16 saat/gün 34°C'de 8 saat/gün (9:00-17:00) (sıcaklık stresi-HS) tel kafeslerde barındırılmıştır. Deneme 2x3 faktöriyel deneme düzenine göre yapılmıştır. Bildircinler temel yem (kontrol) (TN ve HS) ve temel yeme 125 ve 250 ppm biberiye yağı ilave edilen karma yemler ile beslenmiştir. En yüksek canlı ağırlık TN bölümünde 250 ppm biberiye yağı ile beslenen grupta, en düşük canlı ağırlık ise sıcaklık stresi uygulanan tarafta kontrol grubunda gözlenmiştir. Sıcaklık stresi (HS) bildircinlerin canlı ağırlığını olumsuz yönde etkilemiş ($P<0.05$), biberiye yağının bu parametre üzerine etkisi önemsiz bulunmuştur ($P>0.05$). Canlı ağırlık artışı ve yem tüketimi bakımından araştırma grupları arasında önemli farklılıklar tespit edilmemiştir ($P>0.05$). Yemden yararlanma oranı her iki çevre koşulunda da biberiye yağı ilave edilen gruplarda, kontrol gruplarından daha iyi bulunmuştur ($P<0.01$). Sıcaklık stresi karkas verimini kötü yönde etkilemiştir ($P<0.01$). En yüksek sıcak ve soğuk karkas randımanı (g/100 g canlı ağırlıkta) TN şartlarında 250 ppm biberiye yağı ile beslenen grupta gözlenmiş, fakat bu farklılık istatistiksel olarak önemli çıkmamıştır. Sıcaklık stresi grubundaki bildircinlerin kan glikoz seviyesi termo-nötral gruba göre daha yüksek tespit edilmiştir ($P<0.01$). Sıcaklık stresi uygulanan tarafta özellikle 250 ppm biberiye yağı verilen grupta kan glikoz seviyesi düşmüştür ($P<0.05$). Toplam HDL, LDL kolesterol ve trigliserit düzeylerinde deneme grupları arasında önemli farklılık tespit edilmemiştir ($P>0.05$). Sonuç olarak, biberiye yağı ilavesi sıcaklık stresinin olumsuz etkilerini azaltmıştır. Biberiye yağı müteakip çalışmalardan sonra, potansiyel bir doğal yem katkı maddesi olarak düşünülebilir.

Anahtar sözcükler: Biberiye yağı, Performans, Karkas özellikleri, Kan parametreleri, Bildircin, Sıcaklık stresi



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INTRODUCTION

Heat stress is one of the most important factors adversely affecting overall poultry production in tropical countries [1]. Heat stress has been associated with decreases on weight gain, feed intake, feed efficiency and digestibility of nutrients of birds [2]. The ideal temperature for poultry is 10-22°C for optimum body weight and 15-27°C for feed efficiency.

Different natural agents are used to minimize the harmful effects of heat stress on performance of poultry. For example some vitamins [3,4], minerals [5] and substances with antioxidant character such as tomato powder [6], Turkish propolis [7], different essential oils [8] and oil mix (*Thymus serpyllum*, *Laurus nobilis* L., *Myrtle oil*, *Foeniculum vulgare*, *Salvia officinalis*) [9].

Rosemary, (*Rosmarinus officinalis* L.), of the family Labiatae, is an aromatic shrub with an intense pleasant smell reminiscent of pine wood [10]. The essential oil volatile composition of rosemary has been the subject of considerable research in recent years. The principal volatile compounds in rosemary are camphor and 1,8-cineole, followed by borneol, verbenone, α -pinene and camphene [11-13]. Rosemary's oils from natural populations showed high variations in their antimicrobial and antioxidant activity [14,15].

The aim of this study was to evaluate the effects of dietary Rosemary oil (*Rosmarinus officinalis* L.) supplementation on the performance, carcass traits and some blood parameters in Japanese quail reared in thermo-neutral (TN) condition and exposed to high ambient temperatures (HS) of 22 and 34°C, respectively.

MATERIAL and METHODS

Experimental Design and Diet Regimens

A total of 180 fifteen-day-old Japanese quails (*Coturnix coturnix japonica*) obtained from a commercial company (Deva-Yum Marketing Company, Elazig, Turkey) were used after Firat University Animal Ethical Committee approval (Official form date and number: 20.01.2012 and 2012/06). The experiment was conducted at the Poultry Unit of Veterinary Faculty, Firat University, between February 15 to March 14. The birds which were exposed to two different ambient temperature [thermo-neutral (TN) and heat stress (HS)] and three different concentrations of rosemary oil (0, 125 and 250 mg/kg), were divided into 6 treatments consisting of 10 birds of 3 replicates according to 2x3 factorial order. All groups were balanced according to initial live weight and gender. Birds were kept in wire cages in temperature controlled room at either 22°C for 24 h/d (TN) or 22°C for 16 h/d and 34°C (HS) for 8 h/d (from 9:00 to 17:00 h). At both temperatures, birds were fed either a basal diet or the basal diet supplemented with 125 or 250

ppm rosemary oil. The rosemary oil was mixed in a carrier (zeolite), which was then added at one kg per hundred kg to the basal diet. For the rosemary oil treatments, 125 or 250 mg of commercial rosemary oil were added per kg of feed. The concentrations of the volatile components in rosemary oil were shown Table 1. Diets and fresh water were offered *ad libitum*. Light was provided continuously (24 h) throughout the experiment. Ingredients and chemical composition of the basal diet were shown in Table 2. The basal diets contained 23.87% CP and 2897 kcal/kg of ME.

Table 1. The concentration of the volatile components in rosemary oil
Tablo 1. Biberiye yağının içindeki uçucu bileşenlerin konsantrasyonu

Volatile Components	Concentration (%)
1,8 Cineole	39.31
Camphor	14.69
α -Pinene	13.85
β -Pinene	9.87
Camphene	6.17
Limonene	3.17
P-Cymene	2.58
Borneol	2.33
Myrcene	2.02
α -Terpineol	2.28
Bornyl Acetate	1.46
Others	2.27

Table 2. Ingredients and chemical composition of standard diet
Tablo 2. Standart karma yemin bileşimi ve kimyasal kompozisyonu

Feed Ingredients	%	Nutritional Composition	%
Maize	29.03	Dry matter	88.25
Wheat	25.00	Crude protein	23.87
Soybean meal (48 CP)	34.29	Crude fibre	2.55
Corn Gluten	4.10	Ether extract	4.75
Vegetable oil	2.92	Ash	5.45
Dicalcium phosphate	2.02	Calcium ****	1.00
Ground limestone	0.87	Available phosphorus****	0.79
NaHCO ₃	0.12	Methionine ****	0.40
Salt	0.28	Lysine ****	1.18
DL-Metiyonin	0.02	ME, kcal/kg****	2897
Vitamin mix *	0.25		
Mineral mix**	0.10		
Additive***	1.00		

* Vitamin premix supplied per 2.5 kg; Vitamin A 12.000.000 IU; vitamin D₃ 2.000.000 IU; vitamin E 35.000 mg; vitamin K₃ 4.000 mg; vitamin B₁ 3.000 mg; vitamin B₂ 7.000 mg; Niacine 20.000 mg; Calcium D-pantotenat 10.000 mg; vitamin B₆ 5.000 mg; vitamin B₁₂ 15 mg; Folik Asit 1.000 mg; D-Biotin 45 mg; vitamin C 50.000 mg; Choline chloride 125.000 mg; Canthaxanthin 2.500 mg; Apo Karotenolik Acid Ester 500 mg, ** Mineral premix supplied per kg; Mn 80.000 mg; Fe 60.000 mg; Zn 60.000 mg; Cu 5.000 mg; Co 200 mg; I 1.000 mg; Se 150 mg, *** Group Rosemary 0 (1.000 g zeolit); Group Rosemary 125 (12.5 g rosemary oil+987.5 g zeolit); Group Rosemary 250 (25 g rosemary oil + 975 g zeolit), **** Calculated

Feed intake and BW were determined at weekly intervals. The weight gain and feed conversion of birds were then calculated.

At the end of the study (43th day) six males and females quail from each group with an average body weight near the group average were slaughtered and blood samples were collected. Blood samples were centrifuged at $2260 \times g$ for 5 min, and sera were collected. Following slaughtering, hot and cold carcass characteristics were evaluated according to Institute of Turkish standards rules [16].

Chemical Analysis

Serum cholesterol, triglyceride, and glucose concentrations were measured using a biochemical analyzer (Olympus AU-600) at University of Firat, Faculty of Medicine, Department of Biochemistry. Chemical composition of feed ingredients (dry matter, crude protein, ash and ether extract) were analyzed according to the AOAC [17] procedures and crude fiber was determined by the methods of Crampton and Maynard [18].

Statistical Analysis

Data were subjected to two-way anova by using GLM (General Linear Model) procedure. Significant differences were further subjected to Duncan's multiple range test (SPSS [19]). The results were considered as significant when *P* values were lower than 0.05.

RESULTS

The effects of dietary rosemary oil on performance of quails are given in Table 3. As shown in Table 3, the highest final live weight was observed in 250 ppm rosemary oil under the TN condition, the lowest was obtained in control group of HS condition. Heat stress decreased live weight

of quails ($P < 0.05$), supplementation of rosemary oil on live weight was not significant and dose of rosemary oil was not important among the groups of both TN and HS conditions. Live weight gain and feed intake were not significantly different among the treatment groups both in TN and HS conditions ($P > 0.05$). The worst feed conversion ratio was calculated in control groups of both TN and HS conditions. Feed conversion ratio was improved in rosemary oil groups of 125 and 250 ppm in both conditions ($P < 0.01$). The highest hot and cold carcass yield (g/100 g of body weight) was observed in 250 ppm rosemary oil under the TN condition. The lowest was in control group of HS. Under HS condition, deterioration of carcass yield was found significant ($P < 0.01$).

Heat stress affected blood glucose level ($P < 0.01$). Birds kept in HS conditions had greater glucose level than hens kept in TN conditions (Table 4). Rosemary oil decreased blood glucose level especially in 250 ppm rosemary oil group ($P < 0.05$). Total, HDL, LDL cholesterol and triglyceride levels were not significantly different among the treatments ($P > 0.05$).

DISCUSSION

In the present study, rosemary oil supplementation to diet had significant effects on the measured values under thermo-neutral (TN) and heat stress (HS) conditions in growing Japanese quails; it improved performance, positively. The improved performance of rosemary oil groups could be due to these positive effects of rosemary oil on digestive system. In agreement with these results, HERNANDEZ et al. [20] reported that a supplementation of essential oil extract (EOE) from oregano, cinnamon and pepper improved apparent whole tract and ileac digestibility of the nutrients in broilers. JANG et al. [21] showed that a supplementation of a blend of commercial essential oils

Table 3. Effects of rosemary (*Rosmarinus officinalis* L.) supplementation in diet on performance, hot and cold carcass yield in Japanese quail reared under heat stress

Table 3. Sıcaklık stresi altındaki Japon bıldırcınlarında karma yeme ilave edilen biberiye (*Rosmarinus officinalis* L.) yağının performans, sıcak ve soğuk karkas üzerine etkileri

Traits	Rosemary Oil, ppm						Main Effects of Heat Stress and Feed Additive	
	HS			TN				
	0	125	250	0	125	250	HS	FA
Initial Live Weight, g	58.15±0.85	58.10±0.30	58.10±0.50	58.50±1.00	58.20±0.20	58.10±0.40	NS	NS
Final Live Weight, g	192.67±3.88	200.60±4.05	202.54±3.55	206.20±5.80	205.56±3.92	214.11±3.66	*	NS
Live Weight Gain, g/bird/day	4.64±0.12	4.91±0.40	4.98±0.16	5.09±0.17	5.08±0.13	5.38±0.12	NS	NS
Feed Intake, g/bird/day	17.04±0.06	17.15±0.12	17.28±0.15	18.61±0.10	17.76±0.63	18.60±0.37	NS	NS
Feed Conversion Ratio, g feed/g gain	3.68±0.01 ^a	3.50±0.04 ^b	3.47±0.01 ^b	3.66±0.03 ^A	3.50±0.05 ^B	3.46±0.01 ^B	NS	**
Hot Carcass, g hot carcase wt/100 g live wt.	64.73±0.75	65.97±1.03	66.55±1.92	69.92±1.29	69.31±0.90	70.64±1.15	**	NS
Cold Carcass, g cold carcase wt/100 g live wt.	62.70±0.86	64.32±1.09	64.24±1.96	67.71±1.19	67.85±1.27	69.04±1.04	**	NS

NS: Non significant, * $P < 0.05$, ** $P < 0.01$, ^{a,b,A,B}: Mean values with different superscripts within a column differ significantly, HS: Heat stress, TN: Thermo-Neutral, FA: Feed Additive

Table 4. Effects of rosemary (*Rosmarinus officinalis* L.) supplementation in diet on serum glucose and lipid levels in Japanese quails reared under heat stress**Tablo 4.** Sıcaklık stresi altındaki Japon bıldırcınlarında karma yeme ilave edilen biberiye (*Rosmarinus officinalis* L.) yağının serum glikoz ve lipid seviyelerinin üzerine etkileri

Traits, mg/dl	Rosemary Oil, ppm						Main Effects of Heat Stress and Feed Additive	
	HS			TN				
	0	125	250	0	125	250	HS	FA
Glucose	289.67±5.53 ^a	273.67±7.27 ^{ab}	257.33±7.01 ^b	215.80±8.34 ^B	251.17±8.30 ^A	201.80±5.63 ^B	**	*
Total Cholesterol	209.00±32.75	175.00±21.99	165.33±16.71	152.67±29.01	154.60±25.95	149.83±18.56	NS	NS
Triglyceride	919.50±176.44	913.00±90.93	881.83±231.30	680.20±155.41	741.00±243.22	614.33±97.58	NS	NS
HDL Cholesterol	69.33±9.97	69.67±3.52	70.80±7.04	75.00±4.80	82.80±9.64	80.00±5.29	NS	NS
LDL Cholesterol	95.40±12.08	60.83±13.13	60.33±9.03	54.80±10.19	60.00±7.22	58.83±8.51	NS	NS

NS: Non significant, * $P < 0.05$, ** $P < 0.01$, ^{a,b,A,B}: Mean values with different superscripts within a column differ significantly, HS: Heat stress, TN: Thermo-Neutral, FA: Feed Additive

combined with lactic acid increased trypsin and pancreatic amylase activity in broilers. Rosemary's oils obtained from natural plants showed high variations of their antimicrobial and antioxidant activity [14,15]. Previous studies reported that dietary antioxidants, such as vitamin C, E, flavonoids, and phenolic can reduce oxidative damage in animals which is generated by different stress sources [22,23]. Jamroz and Kamel [24] who observed improvements of 7.7% in feed conversion ratio fed a diet supplemented with a plant extract containing capsaicin, cinnamaldehyde and carvacrol in broilers. Improved feed conversion in groups with supplemented rosemary oil may be due to the combined effects of all these active ingredients in a positive manner.

At the inspection of the carcass characteristics (Table 3), there was significant effect of HS on this parameter. In stressed conditions, elevated concentrations of glucocorticoids exert catabolic effects. This demolition decreases the rate of muscle synthesis and thus results in muscle wasting and retardation in growth [25,26]. Supplementation of rosemary oil to diet did not affect the carcass yields at the present study. However, Simsek et al. [8] reported that adding anise oil in the ration had positive effects on the carcass yield in broilers. Also, Alcicek et al. [27] showed that supplementation essential oil (Herbomix™) in the ration had positive effects on the carcass yield in broilers. Possibility, the doses of rosemary oil could not have enough action on catabolic effects of glucocorticoids via anti-oxidant activity [28].

Effects of rosemary (*Rosmarinus officinalis* L.) oil supplementation in diet on blood levels of lipids and glucose were investigated in the present study (Table 4). We observed that birds kept in HS conditions had greater glucose level than hens kept in TN condition and dietary rosemary oil had a positive effect on blood glucose level. This finding indicates that the rosemary oil might be producing its hypoglycemic activity by a mechanism independent from insulin secretion by the inhibition of endogenous glucose production or the inhibition of intestinal glucose absorption [29,30]. In a previous study, it has been suggested that 50% ethanol extract of *Rosmarinus*

officinalis, in part, due to intestinal α -glycosidase (AGc) inhibitory activity of its active compound might play a role in controlling dietary glucose uptake in the small intestinal track [31]. In agreement with current results, BAKIREL et al. [32] investigated potential effect of ethanolic extract of *Rosmarinus officinalis* leaves on glucose homeostasis in rabbits. Results of that study showed that ethanolic extracts of leaves of *Rosmarinus officinalis* reduced blood glucose level in normoglycemic and glucose-hyperglycemic rabbits.

In conclusion, rosemary oil supplemented to diet especially at a level of 250 ppm level supplemented diet had positive effects on performance and blood glucose level. The rosemary oil could therefore be considered as a potential natural feed additive for growing quails, due to increasing consumer's demand for healthy animal production after further studies carried out in different stress conditions.

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REFERENCES

1. Anwar B, Khan Aslam SA, Maqbool A, Khan AKA: Effects of ascorbic acid and acetylsalicylic acid supplementation on the performance of broiler chicks exposed to heat stress. *Pakistan Vet J*, 24, 109-112, 2004.
2. Donkoh A: Ambient temperature: A factor affecting performance and physiological response of broiler chickens. *Int J Biometeorol*, 33, 259-265, 1989.
3. Çiftçi M, Ertaş ON, Güler T: Effects of vitamin E and vitamin C dietary supplementation on egg production and egg quality of laying hens exposed to a chronic heat stress. *Revue Med Vet*, 156, 107-111, 2005.
4. Önel AG, Daşkıran M, Cengiz Ö, Nazlıgül A, Sarı M: Sıcaklık stresi altındaki erken yumurtlama döneminde olan tavukların rasyonlarına E vitamini ve lizin katkısının performans ve yumurta kabuk özellikleri üzerine etkisi. *Kafkas Univ Vet Fak Derg*, 18 (1): 49-54, 2012.
5. Şahin K, Smith MO, Önderci M, Şahin N, Gürsu MF, Küçük O: Supplementation of zinc from organic or inorganic source improves performance and antioxidant status of heat-distressed quail. *Poult Sci*, 84, 882-887, 2005.
6. Şahin N, Orhan C, Tuzcu M, Şahin K, Küçük O: The effects of tomato

powder supplementation on performance and lipid peroxidation in quail. *Poult Sci*, 87, 276-283, 2008.

7. **Seven PT, Seven I, Yılmaz M, Şimşek UG:** The effects of Turkish propolis on growth and carcass characteristics in broilers under heat stres. *Anim Feed Sci Technol*, 146, 137-148, 2008.
8. **Şimşek UG, Çiftçi M, Dalkılıç B, Güler T, Ertaş ON:** The effects of dietary antibiotic and anise oil supplementation on body weight, carcass characteristics and organoleptic analysis of meat in broilers. *Revue Med Vet*, 158, 514-518, 2007.
9. **Karslı MA, Dönmez HH:** Effects of plant extract on growth performance and villi of the small bowel in heat stressed broiler. *Ataturk Univ Vet Bil Derg*, 2, 143-148, 2007.
10. **Szumny A, Figiel A, Gutierrez-Ortiz A, Carbonell-Barrachina AA:** Composition of rosemary essential oil (*Rosmarinus officinalis*) as affected by drying method. *J Food Eng* 97, 253-260, 2010.
11. **Pino JA, Estarron M, Fuentes V:** Essential oil of rosemary (*Rosmarinus officinalis* L.). *J Essential Oil Res*, 10, 111-112, 1998.
12. **Rao LJ, Singh M, Raghavan B, Abraham KO:** Rosemary (*Rosmarinus officinalis* L.): Impact of drying on its flavor quality. *J Food Quality*, 21, 107-115, 1998.
13. **Diaz-Maroto MC, Sanchez Palomo E, Castro L, Gonzalez Vinas MA, Perez-Coello MS:** Changes produced in the aroma compounds and structural integrity of basil (*Ocimum basilicum* L.) during drying. *J Sci Food Agric*, 84, 2070-2076, 2004.
14. **Janssen AM, Scheffer J, Svendsen A:** Antimicrobial activity of essential oils: 1976-1986 literature review. Aspects of test methods. *Planta Medica*, 53, 395-398, 1987.
15. **Moreno S, Scheyer T, Romano CS, Vojnov AA:** Antioxidant and antibacterial activities of rosemary extracts linked to their polyphenol composition. *Free Radic Res*, 40, 223-231, 2006.
16. **Anonymous:** Turk Standartları-Tavuk Govde Eti Parcalama Kuralları. TSE, 1989.
17. **AOAC:** Official Methods of Analysis Association of AOAC International. 17th ed., (AOAC International Maryland), 2000.
18. **Crampton EW, Maynard LA:** The Relation of cellulose and lignin content to nutritive value of animal feeds. *J Nutr*, 15, 383-395, 1983.
19. **SPSS, Inc.** SPSS for Windows Release 11.5 (6 Sep. 2002), Standard Version, Copyright SPSS Inc., Chicago, 2002.
20. **Hernandez F, Madrid J, Garcia V, Orengo J, Megias MD:** Influence of two plant extract on broiler performance, digestibility, and digestive organ size. *Poult Sci*, 83, 169-174, 2004.
21. **Jang IS, Ko YH, Yang HY, Ha JS, Kim JY, Kang SY, Yool DH, Nam DS, Kim DH, Lee CY:** Influence of essential oil components on growth performance and functional activity of the pancreas and small intestine in broiler chickens. *Asian-Australasian J Anim Sci*, 17, 394-400, 2004.
22. **Bagchi D, Carryl OR, Tran MX, Bagchi M, Garg A, Milnes MM:** Acute and chronic stress-induced oxidative gastrointestinal mucosal injury in rats and protection by bismuth subsalicylate. *Mol Cell Biochem*, 196, 109-116, 1999.
23. **Brisibe EA, Umoren UE, Brisibe F, Magalhães PM, Ferreira JFS, Luthria D, Wu X, Prior RL:** Nutritional characterization and antioxidant capacity of different tissues of *Artemisia annua* L. *Food Chem*, 115, 1240-1246, 2009.
24. **Jamroz D, Kamel C:** Plant extracts enhance broiler performance. In non ruminant nutrition: Antimicrobial agents and plant extracts on immunity, health and performance. *J Anim Sci*, 80, 41 (Abstract), 2002.
25. **Hayashi K, Kayalı AG, Tomita Y:** Reduction of corticosterone-induced growth impairment by testosterone and its mechanism. *J Anim Sci Technol*, 63, 1001-1008, 1992.
26. **Higuchi K, Hayashi K, Shimoozaki Y, Ohtsuka A, Tomita Y:** Calcitonin reduces corticosterone-induced muscle proteolysis. *J Nutr Sci Vitaminol*, 41, 545-552, 1996.
27. **Alçiçek A, Bozkurt M, Çabuk M:** The effect of essential oil combination derived from selected herbs growing wild in Turkey on broiler performance. *South Afr J Anim Sci*, 33, 89-94, 2003.
28. **Bubonja-Sonje M, Giacometti J, Abram M:** Antioxidant and antilisterial activity of olive oil, cocoa and rosemary extract polyphenols. *Food Chem*, 127, 1821-1827, 2011.
29. **Platel K, Srinivasan K:** Plant foods in the management of diabetes mellitus: Vegetables as potential hypoglycaemic agents. *Nahrung*, 2, 68-74, 1997.
30. **Eddouks M, Jouad H, Maghrani M, Lemhadri A, Burcelin R:** Inhibition of endogenous glucose production accounts for hypoglycemic effect of *Spergularia purpurea* in streptozotocin mice. *Phytomedicine*, 6-7, 594-599, 2003.
31. **Koga K, Shibata H, Yoshino K, Nomoto K:** Effect of 50% ethanol extract rosemary (*Rosmarinus officinalis*) on α -glucosidase inhibitory activity and the elevation of plasma glucose level in rats, and its active compound. *J Food Sci*, 71, 507-512, 2006.
32. **Bakirel T, Bakirel U, Keles OU, Ulgen SG, Yardibi H:** In vitro assessment of antidiabetic and antioxidant activities of rosemary (*Rosmarinus officinalis*) in alloxan-diabetic rabbits. *J Ethnopharmacol*, 116, 64-73, 2008.

The Relationships Among Some Udder Traits and Somatic Cell Count in Holstein-Friesian Cows ^[1]

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Summary

This study was carried out to investigate the relationships between somatic cell count (SCC) and some udder traits in 129 head Holstein-Friesian cows (1st and 5th month of lactation) raised at 3 dairy farm in Kamışlar Village in Bozdogan county of Aydin province, Turkey. Farms were visited monthly between July 2011 and June 2012, and about 1108 milk samples and different udder measures were taken in 50±20 days, 90±20 days, 130±20 days of lactation of each cow studied. Mean log₁₀SCC were calculated as 4.9±0.003 (87.923). The overall results showed that dairy farms and month of year groups had a significant effect on log₁₀SCC (P<0.01). Positive correlations among fore teats perimeter and log₁₀SCC (0.11) in farm A and among and log₁₀SCC distance between fore teats (0.12) and distance between rear teats (0.10) in farm B, were found at the level of P<0.01, respectively. The correlation between rear teats perimeter and log₁₀SCC (-0.12) was found to be negative and statistically significant in farm C (P<0.01). Consequently, udder traits relevant to minimization or elevation of SCC should be carefully evaluated in selection studies.

Keywords: Holstein-Friesian, Cow, Somatic Cell Count, Udder Traits

Siyah Alaca İneklerdeki Somatik Hücre Sayısı ile Bazı Meme Özellikleri Arasındaki İlişkiler

Özet

Bu araştırma, Aydın ili Bozdoğan ilçesi Kamışlar Köyü'ndeki 3 süt sığırcılığı işletmesinde yetiştirilen 129 baş Siyah Alaca ineğin (laktasyonun 1. ve 5. ayları arasında) somatik hücre sayısı ile bazı meme özellikleri arasındaki ilişkileri incelemek amacıyla yapılmıştır. İşletmelere Temmuz 2011 ve Haziran 2012 tarihleri arasında, ayda bir gidilerek laktasyonun 50±20 gün, 90±20 gün, 130±20. günlerinde yaklaşık 1108 adet süt örneği ile farklı meme ölçüleri alınmıştır. Ortalama log₁₀SHS 4.9 ± 0.003 (87.923) olarak hesaplanmış, log₁₀SHS üzerine işletme ve ayın istatistiki olarak önemli bir etkisi olduğu tespit edilmiştir (P<0.01). A işletmesinde ön meme başı çapı ile log₁₀SHS arasındaki (0.11) ve B işletmesinde log₁₀SHS ile ön meme başları arası mesafe (0.12) ve arka meme başları arası mesafe (0.10) arasındaki korelasyonlar P<0.01 düzeyinde bulunmuştur. C işletmesinde log₁₀SHS ile arka meme başı çevresi (-0.12) arasındaki korelasyon negatif ve istatistik bakımından önemli çıkmıştır (P<0.01). Çalışmada, SHS'nı azaltan ya da arttıran meme özelliklerinin seleksiyon çalışmalarında dikkatlice değerlendirilmesi sonucuna varılmıştır.

Anahtar sözcükler: Siyah-Alaca, İnek, Somatik hücre sayısı, Meme özellikleri

INTRODUCTION

Milk yield improvement has been well known as the main purpose of most dairy breeders, besides it has also well been documented that single-trait selection for milk production may prone cattle to decreased immunity (resistance against disease) and reproduction efficiency ^[1-3]. Udder diseases, predominantly mastitis as one of the most frequent and economical cow disease, may possess increasing costs on milk producers ^[3-6] and finally lead to obligatory culling ^[7].

Reported heritability for clinical mastitis is very low in general and may vary from 0.02 ^[2] to 0.05 ^[8]. As for this, correlated traits may be used for indirect selection for improvement of mastitis resistance, majorly somatic cell score (SCC - log-transformed somatic cell count), besides within some selected conformation traits ^[3]. In a previous study genetic correlations of the same traits among different lactations were found positive (moderate to high), suggesting that SCC and clinical mastitis may be considered



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as the same traits for different lactations. In addition genetic correlations of clinical mastitis and SCC varied from 0.37 to 0.68^[7], with the average correlation close to 0.70^[2,9], for the first and third lactations, respectively, showing that clinical mastitis and SCC present different features of udder health^[5]. The heritability of SCC is also low, varying from 0.06^[10] to 0.13^[11].

Cell numbers involved in milk are closely related to inflammation and udder health. SCC in raw and heat-treated milk from a healthy mammary gland according to EU Directive (Health and Hygiene Directive 92/46/EEC), is lower than 400.000 cells/ml, SCC in milk and milk products is lower than 500.000 cells/ml^[12].

Somatic cell count, a useful predictor of intramammary infection involving leucocytes^[13], is totally accepted as for the standard measurement of milk quality worldwide. Therefore SCC is readily available in almost dairy farmer at least monthly^[14]. Widely distributed data are now available worldwide on large population of cows regarding factors influencing SCC in milk. A variety of reviews or relevant studies have addressed issues regarding SCC, their variety and probable use for detecting the quality of milk^[14-26]. An elevated SCC in milk has a negative influence on the quality^[21,27-31]. Subclinical mastitis has always been recognized as a result of low milk production, changes related to milk consistency (density), diminished probability of adequate milk processing, lowered protein and higher risk for milk hygiene^[13]. Similar confounding were reported by^[32] indicating a positive relation among SCC and milk yield for the first lactation, and a negative relation for the processing lactations. Clinical mastitis is more likely to be included in a breeding in contrast to SCC, whereas moderate to strong genetic correlations between clinical mastitis and SCC may suggest that selecting diminished SCC may improve mastitis resistance^[33].

The relationships between different udder traits, milk yield and SCC were investigated by one researcher. The genetic correlations among udder height, rear udder height,

cleft depth, udder levelness and SCC were found -0.41, -0.44, 0.20 and 0.21, respectively^[34].

Some authors investigated the farm effects on SCC^[35-40]. Many others reported that the effects of stage of lactation, month and parity on SCC were considered^[5,14,29,31,37,39,41-49].

The objective of this study was to investigate the relationship between some udder traits and SCC in Holstein-Friesian cows raised at selected dairy farms in Bozdogan county, Aydin province.

MATERIAL and METHODS

The survey was conducted with 129 head Holstein-Friesian cows were determined at 1st-5th months of lactation and managed at 3 dairy farms in Kamislar Village in Bozdogan county of Aydin province, Turkey. The general features of the dairy farms are shown in the *Table 1*. Farms were visited monthly from July 2011 to June 2012 and were collected about 1108 milk samples and different udder and teat of each cows were measured into 3 lactation stage (50±20 days, 90±20 days, 130±20 days of lactation). In this study, cows were separated 4 parity groups. The cows having the fourth and larger lactation numbers were included in Parity 4.

Collection of Milk Samples

The milk samples from each cow were withdrawn from each teat into tubes among both the morning and evening milking samples. Morning milking was stored in a cooler box and then was immediately transferred to the laboratory for analyzing on the same day. Evening milk samples were kept in a refrigerator and analyzed the next day immediately. The milk samples were treated according to the directions given previously^[50] and Standard Methods for the Examination of Dairy Products^[51]. SCC in the samples was determined by direct microscopic SCC method^[52]. Milk samples were spread on 2 microscope slide areas, with 5×20 mm² in size. Slides were arranged

Table 1. General features of the dairy farms

Tablo 1. Süt sığırcı işletmelerinin özellikleri

General Features	Farm A	Farm B	Farm C
N cows (N=129)	51	42	36
Barn type	Open Concrete-dirt surface Free stall	Open Concrete-dirt surface Free stall	Open Concrete-dirt surface Free stall
Animal sill	Using/Hay sill	Using/Hay sill	Using/Hay sill
Milking system	Milking parlor (1x10)	Milking parlor (1x9)	Milking parlor (2x5)
Milking machine cleaning	Rightly	Rightly	Rightly
Pre-milking/post-milking udder cleaning and disinfection	Water- Disinfection Using	Water- Disinfection Using	Disinfection Using
Feeding	During milking Roughage + Mixed feed	During milking Roughage + Mixed feed	During milking Roughage + Mixed feed
Touring Area	In barn	In barn	In barn

according to Direct Microscopic Somatic Cell Count (DMSCC) and kept at 37°C in an incubator. Then, milk samples on slides were painted in the manner of the technique of a dye solution involving methylene blue. The dye solution was prepared with combined of 0.6 g of certified methylene blue chloride to 54 mL of 96% ethyl alcohol, 40 mL of tolien and 6 mL glacial acetic acid. The counting was fulfilled in 20 fields under a 100x immersion objective in each slide and averages were calculated. The multiplication of the microscope factor with this average values was corresponded to SCC in per mL of milk^[53].

Udder and Teat Measurement

The distance from fore and rear teats to the floor (DFTF and DRTF, respectively) and vertical length of fore and rear teats (FTL and RTL, respectively), the distance between fore, rear and side teats (FTD, RTD and STD, respectively) and the fore and rear teat perimeter (FTP and RTP, respectively) were measured by using a ruler. The fore and rear teat diameter (FTDM and RTDM, respectively) was measured at the mid-point of each teat by the help of calipers. All measures were performed after and before morning and evening milking.

The SCC values were transformed to \log_{10} for normality and homogeneity of variances in the SCC data. The statistical analyses were performed using least squares method in the General Linear Model (GLM) procedure of Minitab package program^[54]. The means were compared by Duncan's multiple range test^[55]. The statistical model used for the analysis is as follows:

$$Y_{ijklm} = m + s_i + a_j + p_k + lp_l + e_{ijklm}$$

Where; y_{ijklm} : i. farm, j. month, k. parity, l. lactation period, m. cow's logarithmic SCC, μ : population mean, s_i : i. farm's effect (i: 1, 2, 3), a_j : j. month's effect (j: 1, 2, ..., 12), p_k : k. parity's effect (k: 1, 2, 3, 4), lp_l : l. lactation period's effect (1: 50±20 days, 2: 90±20 days, 3: 130±20 days), e_{ijklm} : residual random error.

In this study, the correlation coefficients were calculated between udder measures and \log_{10} SCC. To compute correlations between \log_{10} SCC and udder measures, Pearson's correlation analysis were used. Other statistical analysis was performed using SPSS statistical packet program^[56].

RESULTS

Basic Statistics

The mean values of 305-day milk yield were ranged from 4142 to 4557 kg in farm A, from 3930 to 4847 kg in farm B and from 4344 to 5100 kg in farm C. According to analyses, differences between lactations were non-significant in each farm ($P > 0.05$).

The mean values of udder traits were changed

between 1.98 cm and 1.87 cm for FTDM, 1.82 cm and 1.86 cm for RTDM, 5.86 cm and 6.30 cm in for FTL, 5.24 cm and 5.51 cm for RTL, 8.04 cm and 8.10 cm for FTP, 7.96 cm and 8.15 cm for RTP, 14.71 cm and 15.62 cm for FTD, 7.78 cm and 7.84 cm for RTD, 12.26 cm and 12.75 cm for STD, 52.80 cm and 53.91 cm for DFTF, 53.45 cm and 53.95 cm for DRTF in farm A.

The mean values of udder traits were changed between 2.13 cm and 1.93 cm for FTDM, 1.92 cm and 1.90 cm for RTDM, 5.91 cm and 6.43 cm for FTL, 5.31 cm and 5.79 cm for RTL, 8.12 cm and 7.83 cm for FTP, 8.04 cm and 8.28 cm for RTP, 15.30 cm and 15.42 cm for FTD, 8.69 cm and 8.80 cm for RTD, 12.04 cm and 12.34 cm for STD, 52.23 cm and 50.99 cm for DFTF, 51.74 cm and 51.65 cm for DRTF in farm B.

The mean values of udder traits were 1.88 cm and 1.81 cm for FTDM, 1.79 cm and 1.81 cm for RTDM, 5.63 cm and 6.13 cm for FTL, 5.07 cm and 5.67 cm for RTL, 8.06 cm and 7.24 cm for FTP, 7.80 cm and 7.10 cm for RTP, 13.98 cm and 13.45 cm for FTD, 7.66 cm and 7.40 cm for RTD, 11.72 cm and 11.78 cm for STD, 52.86 cm and 54.23 cm for DFTF, 53.94 cm and 55.66 cm for DRTF in farm C.

The least square means and standard error of means of \log_{10} SCC are shown in Table 2.

The results were shown that \log_{10} SCC was reached the highest value in July and August and also was reached the lowest values between April and June. The values of \log_{10} SCC were decreased from parity 1th to 3rd and were increased to parity 4.

The differences between farms were found statistically significant for \log_{10} SCC ($P < 0.01$) and the analysis revealed that the effects of months were also statistically significant for \log_{10} SCC ($P < 0.01$). The differences between parity and lactation periods were found statistically non-significant for \log_{10} SCC ($P > 0.05$).

Correlations Between SCC and Udder Traits

The correlations between \log_{10} SCC and udder measures are presented in Table 3. The correlation among \log_{10} SCC and udder traits generally tended to negatively and was only found positive and statistically significant correlation between \log_{10} SCC and FTDM (0.11) in farm A ($P < 0.05$). The correlations among \log_{10} SCC-FTD (0.12) and \log_{10} SCC-RTD (0.10) were found positively and statistically significant ($P < 0.01$). The correlations between \log_{10} SCC and udder measures were also found negatively and the highest correlation values was determined to -0.12 (\log_{10} SCC-RTP) and statistically significant in farm C ($P < 0.01$).

DISCUSSION

In the given circumstances (twice a day milking), $\text{SCC} < 100,000$ cell/mL in milks were obtained from udder

Table 2. The least square means and standard error of means of \log_{10} SCC (cell/ml)**Tablo 2.** \log_{10} SHS'nin en küçük kareler ortalamaları ve standart hataları (hücre/ml)

Factors	N	X±Sx	
		\log_{10} SCC	Re-transformation SCC
Farm		**	
A	448	4.90 ^b ±0.003	80.687
B	360	5.09 ^a ±0.007	123.956
C	300	4.94 ^b ±0.004	87.923
Months		**	
1 (January)	142	4.96 ^c ±0.070	92.083
2 (February)	140	4.94 ^c ±0.060	88.919
3 (March)	136	4.94 ^c ±0.005	87.673
4 (April)	106	4.92 ^c ±0.002	83.999
5 (May)	66	4.92 ^c ±0.060	84.123
6 (June)	62	4.92 ^c ±0.004	84.661
7 (July)	46	5.16 ^a ±0.033	146.812
8 (August)	46	5.06 ^b ±0.030	115.401
9 (September)	60	4.99 ^{bc} ±0.019	99.052
10 (October)	82	5.00 ^{bc} ±0.016	100.582
11 (November)	104	5.01 ^{bc} ±0.014	103.137
12 (December)	118	4.94 ^c ±0.005	87.909
Parity		N.S	
1	354	5.00±0.005	93.854
2	328	4.99±0.007	99.583
3	172	4.94±0.006	88.876
4	254	4.96±0.006	91.853
Lactation period		N.S	
1 (50 ± 20)	518	5.00±0.005	100.257
2 (90 ± 20)	386	4.95±0.004	90.783
3 (130 ± 20)	204	4.93±0.003	85.930
The overall mean	1108	4.94±0.003	87.923

N.S: Non-significant, * ($P<0.05$), ** ($P<0.01$), a,b,c: Means with different superscripts in each line are different

lobes that healthy and uninfected was reported [57]. In this study, SCC was determined as 4.94 ± 0.003 (87.923) cell/mL. The vales of mean SCC (4.94 ± 0.003 (87.923) in milk was lower than threshold value (500.000 cell/mL) that was acknowledged according to EU Directive. The mean values of SCC were found lower from findings of some researchers [29,36,37,45,46] and were found higher from findings of some researchers [47]. These circumstances is an indicator of conform to criteria of milk quality in farms in this study.

In the present study, the effect of farm on SCC was determined statistically significant ($P<0.01$). This result was found similar to findings of some other authors [33,35,36,38,39] and was showing differences with findings of some authors [37,44,46]. Also, the effects of month on SCC was

Table 3. The correlations between \log_{10} SCC and udder traits in farms**Tablo 3.** İşletmelerde meme özellikleri ile \log_{10} SHS arasındaki korelasyonlar

Traits	Farms		
	A	B	C
FTDM	0.11*	0.03	-0.08
RTDM	-0.04	-0.02	-0.01
FTL	-0.08	-0.05	-0.03
RTL	-0.04	-0.05	-0.07
FTP	-0.04	-0.01	-0.11
RTP	-0.06	-0.30	-0.12*
FTD	-0.01	0.12**	-0.07
RTD	-0.04	0.10*	0.02
STD	-0.05	0.09	-0.01
DFTF	-0.04	-0.04	-0.01
DRTF	-0.01	0.22	0.09

* ($P<0.05$), ** ($P<0.01$), FTDM: Fore teat diameter, RTDM: Rear teat diameter, FTL: Fore teat length, RTL: Rear teat length, FTP: Fore teat perimeter, RTP: Rear teat perimeter, FTD: Fore teat distance, RTD: Rear teat distance, STD: Side teat distance, DFTF: Distance from fore teat to floor, DRTF: Distance from rear teat to floor

determined statistically significant ($P<0.01$). The SCC values was found higher in summer months than winter months and the results of environmental temperature and hormonal mechanism of cows were reported from some researchers [29,36,40]. In this study, the effect of measuring month on SCC was found statistically significant ($P<0.01$), the highest value of SCC was determined on July-August and the lowest value of SCC on April, May and June. This result was found similar with findings of some researchers [36,38,48] and was found different from some authors [47]. The primary cause of differences in results may be heat stress from calving cow in summer season.

When analyzed to variations of SCC in accordance with parity, the highest value of SCC was determined in first parity and was found similar to values in other parity. The effect of parity on SCC was found statistically non-significant ($P>0.05$). This result was similar with findings of some researchers [46] and was found different from findings of some other authors [5,36,42,45,47].

In the present study, the effect of lactation stage on SCC was found statistically non-significant ($P>0.05$), while the effect of lactation stage on SCC was found statistically significant by some researchers [5,13,29,30,41-47].

In the present study, the positive correlation between SCC and FTP was found statistically significant in Farm A ($P<0.01$). The correlations SCC-FTD and SCC-RTD were found positive and statistically significant in Farm B ($P<0.01$) (respectively 0.12 and 0.10). The correlation between SCC and RTP was found negative and significant ($P<0.01$). These results were found similar with findings of some researchers [9,47]. Finally, correlations between SCC and

other udder traits were found statistically non-significant ($P>0.05$), especially relationships between SCC and FTL were found similar with findings of some researchers^[58].

The results from this study indicate that some important findings about the correlations between SCC and some udder traits were obtained in farms. The values of mean SCC (4.94 ± 0.003 (87.923) cell/mL) in milk was lower than threshold value (500.000 cell/mL) that was acknowledged according to EU Directive. The dairy cows are approved to physiologically for SCC in farms. In the sense of udder healthy optimum management programs should be carried to seasonal. Hereby, udder traits relevant to minimization or elevation of SCC should be carefully evaluated in selection studies.

REFERENCES

- Dematawewa CMB, Berger PJ:** Genetic and phenotypic parameters for 305-day yield, fertility and survival in Holsteins. *J Dairy Sci*, 81, 2700-2709, 1998.
- Negussie E, Stradén I, Mäntysaari EA:** Genetic association of clinical mastitis with test-day somatic cell score and milk yield during first lactation of Finnish Ayrshire cows. *J Dairy Sci*, 91, 1189-1197, 2008.
- Ptak E, Jagusiak W, Zarnecki A:** Relationship between test day somatic cell score and conformation traits in Polish Holstein cattle. *59th Annual Meeting of the European Association for Animal Production*, Vilnius, Lithuania - August 24-27, 2008.
- Kadarmideen HN:** Genetic correlations among body condition score, somatic cell score, milk production, fertility and conformation traits in dairy cows. *Animal Science*, 79, 191-201, 2004.
- Şeker İ, Rişvanlı A, Kul S, Bayraktar M, Kaygusuzoğlu E:** İsviçre Esmeri ineklerde meme özellikleri ve süt verimi ile CMT skoru arasındaki ilişkiler. *Lalahan Hay Araşt Enst Derg*, 40 (1): 29-38, 2000.
- Kul E, Erdem H, Atasever S:** Süt sığırlarında farklı meme özelliklerinin mastitis ve süt somatik hücre sayısı üzerine etkileri. *OMÜ Zir Fak Dergisi*, 21 (3): 350-356, 2006.
- Pösö J, Mäntysaari EA:** Relationships between clinical mastitis, somatic cell score, and production for the first three lactations of Finnish Ayrshire. *J Dairy Sci*, 79, 1284-1291, 1996.
- Lund MS, Jensen J, Peterson PH:** Estimation of genetic and phenotypic parameters for clinical mastitis, somatic cell production deviance, and protein yield in dairy cattle using Gibbs sampling. *J Dairy Sci* 82, 1045-1051, 1999.
- Rupp R, Boichard D:** Genetic parameters for clinical mastitis, somatic cell score, production, udder type traits, and milking ease in first lactation Holsteins. *J Dairy Sci*, 82, 2198-2204, 1999.
- Dal Zotto R, De Marchi M, Dalvit C, Cassandro M, Gallo L, Carnier P, Bittante G:** Heritabilities and genetic correlations of body conformation score and calving interval with yield, somatic cell score, and linear type traits in Brown Swiss cattle. *J Dairy Sci*, 90, 5737-5743, 2007.
- De Haas Y, Ouweltjes W, Ten Napel J, Windig JJ, De Jong G:** Alternative somatic cell count traits as mastitis indicators for genetic selection. *J Dairy Sci*, 91, 2501-2511, 2008.
- Anonymous:** Europa, EU. Animal Health and Welfare, http://europa.eu/eur-lex/en/consleg/pdf/1992/en_1992L0046_do_001.pdf, 1992.
- Sharma N, Singh NK, Bhadwal MS:** Relationship of somatic cell count and mastitis: An overview. *Asian-Aust J Anim Sci*, 24 (3): 429-438, 2011.
- Harmon RJ:** Somatic cell counts: A primer. In, *Proc. National Mastitis Council Annual Meeting*, Reno, NV, USA, pp.3-9, 2001.
- Bodoh GW, Battista WJ, Schultze LH, Johnston RP:** Variation in somatic cell counts in dairy herd improvement milk samples. *J Dairy Sci*, 59, 1119, 1976.
- Brolund L:** Individual cow somatic cell counting: Diagnostic significance and applicability. *Kieler Milchwirtschaftliche Forschungsberichte*, 37, 286, 1985.
- Dohoo IR, Meek AH:** Somatic cell counts in bovine milk. *Can Vet J*, 23, 119, 1982.
- Eberhart RJ, Hutchinson LJ, Spencer SB:** Relationships of bulk tank somatic cell counts to prevalence of intramammary infection and to indices of herd production. *J Food Protect*, 45, 1125, 1982.
- Harmon RJ:** Physiology of mastitis and factors affecting somatic cell counts. *J Dairy Sci*, 77, 2103, 1994.
- Miller RH, Paape MJ:** Relationship between milk somatic cell count and milk yield. *Proc Ann Mtg Natl Mastitis Counc*, Arlington VA, USA, p.60, 1985.
- Raubertas RF, Shook GE:** Relationship between lactation measures of somatic cell concentration and milk yield. *J Dairy Sci*, 65, 419, 1982.
- Reneau JK:** Using DHI somatic cell counts. *Proc Ann Mtg Natl Mastitis Counc*, Arlington, Virginia, USA, p.73, 1985.
- Reneau JK:** Effective use of dairy herd improvement somatic cell counts in mastitis control. *J Dairy Sci*, 69, 1708, 1986.
- Reneau JK, Packard VS:** Monitoring mastitis, milk quality and economic losses in dairy fields. *Dairy Food Environ, Sanitation*, 11 (1): 4-11, 1991.
- Schultz LH:** Somatic cells in milk - physiological aspects and relationship to amount and composition of milk. *J Food Protect*, 40, 125, 1977.
- Sheldrake RF, Hoare RJT, McGregor GD:** Lactation stage, parity and infection affecting somatic cells, electrical conductivity and serum albumin in milk. *J Dairy Sci*, 66, 542, 1983.
- Jones GM, Pearson RE, Clabaugh GA, Heald CM:** Relationship between somatic cell counts and milk production. *J Dairy Sci*, 67, 1823-1831, 1984.
- Jones GM:** Qualifying milk under reduced somatic cell count limit. *Dairy Pipeline*, Virginia Cooperative Extension, 1999.
- Eyduran E, Özdemir T, Yazgan K, Keskin S:** Siyah Alaca inek sütündeki somatik hücre sayısına laktasyon sırası ve dönemin etkisi. *YYÜ Vet Fak Derg*, 16 (1): 61-65, 2005.
- Juozaitiene V, Juozaitis A, Mickeviciene R:** Relationship between somatic cell count and milk production or morphological traits of udder in Black and White cows. *Turk J Vet Anim Sci*, 30, 47-51, 2006.
- O'Brien B, Berry DP, Kelly Meaney WJ, O'Callaghan EJ:** A study of the somatic cell count (SCC) of Irish milk from herd management and environmental perspectives, <http://www.teagasc.ie/research/reports/foodprocessing/5399/eopr-5399.pdf>, 2009.
- Miura S:** Genetic correlations of milk production and type traits with somatic cell score from Holstein cows in Hokkaido. *Animal Science Journal*, 76 (4): 401-406, 2005.
- Coffey EM, Vinson WE, Pearson RE:** Potential of somatic cell concentration in milk as a sire selection criterion to reduce mastitis in dairy cattle. *J Dairy Sci*, 69 (8): 2163-2172, 1986.
- Rogers GW:** Index selection using milk yield, somatic cell score, udder depth, teat placement, and foot angle. *J Dairy Sci*, 76, 664-670, 1993.
- Barkema HW, Van Der Ploeg JD, Schukken YH, Lam TJGM, Benedictus G, Branda A:** Management style and its association with bulk milk somatic cell count and incidence rate of clinical mastitis. *J Dairy Sci*, 82, 1655-1663, 1999.
- Uzmay C, Kaya A, Kaya İ, Akbaş Y:** İzmir ili Holstein Damızlık Süt Sığırı Yetiştiriciliği Birliği işletmelerinde mastitisin yaygınlık düzeyi ve etkileyen etmenler üzerine araştırmalar, 2. Yönetim uygulamaları ile subklinik mastitis arası ilişkiler. *Ege Üniv Ziraat Fak Derg*, 38 (2-3): 71-78, 2001.
- Göncü S, Özkütük K:** Adana entansif süt sığırcılığı işletmelerinde yetiştirilen saf ve melez Siyah Alaca inek sütlerinde somatik hücre sayısına etki eden faktörler ve mastitis ile ilişkisi. *Hayvansal Üretim*, 43 (2): 44-53, 2002.
- Topaloğlu N, Güneş H:** İngiltere'de yetiştirilen Siyah-Alaca sığırların

süt verimi özellikleri üzerinde araştırmalar. *İstanbul Üniv Vet Fak Derg*, 31 (1): 149-164, 2010.

39. Özdede F: Ankara ili Damızlık Süt Sığırını Yetiştiriciliği Birliğine üye süt sığırcılığı işletmelerinde üretilen sütlerin somatik Hücre sayıları. *Doktora Tezi*, Ankara Üniv. Fen Bil. Enst., 2009.

40. Bytqi H, Zaugg U, Sherifi K, Hamidi A, Gjonbalaj M, Muji S, Mehmeti H: Influence of management and physiological factors on somatic cell count in raw cow milk in Kosovo. *Veterinaski Arhiv*, 80 (2): 173-183, 2010.

41. Busato A, Trachsel P, Schallibaum M, Blum JW: Udder health and risk factors for subclinical mastitis in organic dairy farms in Switzerland. *Preventive Vet Med*, 44 (3-4): 205-220, 2000.

42. De Haas Y: Somatic cell count patterns. Improvement of udder health by genetics and management, *PhD Thesis*, Wageningen University, 2003.

43. De Haas Y, Barkema HW, Schukken YH, Veerkamo RF: Genetic parameters for clinical mastitis and traits for somatic cell count based on its lactation curve. *7th World Congress on Genetic Applied to Livestock Production*, August 19-23. Montpellier, France, 2002.

44. Koivula M, Negussie E, Mantysaari EA: Genetic parameters for test-day somatic cell count at different stages of lactation in Finnish Ayrshire cattle. *7th World Congress on Genetic Applied to Liv Pro*, August 19-23, Montpellier, France, 2002.

45. Koç A: Aydın'da yetiştirilen Siyah-Alaca ve Esmer ırkı sığırlarda sütteki somatik hücre sayısının değişimi. *4. Ulusal Zootečni Kongresi*, 1-3 Eylül, SDÜ Ziraat Fakültesi Zootečni Bölümü, Isparta, 2004.

46. Koç A: Aydın ilinde yetiştirilen Siyah-Alaca ve Esmer ırkı sığırların laktasyon süt verimleri ve somatik hücre sayıları. *Hayvansal Üretim*, 47 (2): 1-8, 2006.

47. Coban O, Sabuncuoglu N, Tuzemen N: Siyah Alaca ve Esmer ineklerde somatik hücre sayısına çeşitli faktörlerin etkisi. *Lalahan Hay Arast Enst Derg*, 47 (1): 15-20, 2007.

48. Gökçe G: Çukurova Bölgesi Entansif Süt Sığırını İşletmelerindeki ilkinde doğuran Siyah Alacalarda somatik hücre sayısına etki eden bazı tip, sağım ve amenajman özellikleri arası ilişkiler. *Doktora Tezi*, Çukurova Üniv. Fen Bil. Enst., 2011.

49. Temelli S, Şerbetçioğlu T: Bir süt işletmesinde işlenen inek sütlerinde somatik hücre sayısının dört yıllık periyottaki değişiminin incelenmesi. *Uludağ Üniv J Fac Vet Med*, 30 (1): 1-7, 2010.

50. Breed RS: The microscopic appearance of unpasteurized market milk and cream. *New York State, Agricultural Experiment Station*, Geneva, Bull, 566, 1929.

51. American Public Health Association: Standard methods for the examination of dairy products. *Amer Public Health Assoc*, 1939.

52. Packard VS, Tatini VS, Fugua R, Heady J, Gilman C: Direct microscopic methods for bacteria or somatic cells. *16th American Public Health Association*, Washington, DC, USA, pp.309-321, 1992.

53. Packard VS, Ginn RE: Standard methods for the examination of dairy products. *American Public Health Association*, Washington, DC, USA, pp.219-237, 1985.

54. Anonymous: Minitab Release 16.2 Version for Windows. Copyright © 2010, Minitab Inc., 2010.

55. Duncan DB: Multiple range and multiple F test. *Biometrics*, 11, 42, 1995.

56. Anonymous: SPSS Statistics Release 17.0.0. Copyright 1993-2007 Polar Engineering and Consulting, <http://www.winwrap.com>, 2008.

57. Kaya İ, Uzmay C, Ayılmaz T, Ünlü HB: Ege Üniversitesi Ziraat Fakültesi Menemen Araştırma ve Uygulama Çiftliğinde yetiştirilen Siyah Alaca ineklerde somatik hücre ölçümüne dayalı olarak meme sağlığı. *Ege Üniv Ziraat Fak Derg*, 48 (3): 229-239, 2011.

58. Şeker, İ, Rişvanlı A, Bayraktar M, Kul S, Kaygusuzoğlu E: İsviçre Esmeri ineklerde bazı linear tip özellikleri ile mastitis arasındaki ilişkiler üzerine bir çalışma. *Vet Bil Derg*, 16 (2): 111-117, 2000.

Effects of Acrylamide Treatment on Oxidant and Antioxidant Levels in Rats

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Summary

The aim of this study was to investigate serum total antioxidant status (TAS), total oxidant status (TOS) and ischemia-modified albumin (IMA) levels in long term acrylamide (ACR) given rats, compared to control rats. In total, 25 male and 25 female Wistar rats were involved in this experiment. Animals in each sex were segregated into three groups. Two of them were treatment groups and one of them was control group. Each treatment group consisted of ten animals and each control group consisted of five animals. ACR was administered to the treatment groups at 2 and 5 mg/kg/day via drinking water for 90 days. In the end of the experiment, serum samples were analyzed for IMA, TAS, TOS and albumin levels with the spectrophotometric method. Serum IMA and adjusted IMA levels were significantly higher at concentrations of 2 mg/kg and 5 mg/kg in the male rats when compared with those of the control male rats. Serum TAS levels significantly decreased at concentrations of 5 mg/kg in the male rats when compared with those of the control rats. We also observed a significant increase in the levels of serum TOS at concentrations of 5 mg/kg in the male rats. There were no significant differences between serum IMA, TAS, TOS and albumin levels at concentrations of 2 mg/kg and 5 mg/kg in the female rats. Our findings show that long term treatment with 2 mg/kg and 5 mg/kg doses of ACR led to a significant depletion of serum TAS levels and overproduction of serum TOS and IMA levels, consequently, to an increase in oxidative stress.

Keywords: Acrylamide, Total antioxidant status, Total oxidant status, Ischemia-modified albumin, Oxidative stress

Ratlarda Akrilamid Kullanımının Antioksidan ve Oksidan Değerleri Üzerine Etkisi

Özet

Bu çalışmada, uzun süre akrilamid verilen sıçanlar üzerinde total antioksidan durum (TAS), total oksidan durum (TOS) ve iskemi modifiye albumin'in (IMA) serum düzeylerinin nasıl değiştiğinin araştırılması amaçlanmıştır. Çalışmada 65-75 g ağırlığında ve yaşları 3-4 haftalık 25 erkek ve 25 dişi Wistar cinsi sıçanlar kullanılmıştır. Hayvanlar 90 gün boyunca standart sıçan yemi ile beslenmişlerdir. Bununla beraber, günlük tüketecekleri içme suyuna 2 mg/kg/gün ve 5 mg/kg/gün dozunda akrilamid ilave edilmiştir. Akrilamid uygulaması sonrası hayvanlar anestezi altında servikal dislokasyonla öldürülmüş ve serumlarında IMA, TAS, TOS ve albumin düzeyleri spektrofotometrik yöntem ile ölçülmüştür. 2 mg/kg ve 5mg/kg akrilamid verilen erkek sıçanlara ait serum IMA düzeyleri kontrol grubuna göre önemli derecede yüksek bulunmuştur. Ayrıca, 5mg/kg akrilamid verilen erkek sıçanlara ait serum TAS düzeyleri kontrol grubuna göre önemli derecede düşük ve serum TOS değerleri önemli derecede yüksek bulunmuştur. 2 mg/kg ve 5mg/kg akrilamid verilen dişi sıçanlara ve kontrol grubuna ait serum IMA, TAS, TOS ve albumin düzeyleri arasında istatistiki açıdan önemli bir fark bulunamamıştır. Bu sonuçlara bağlı olarak bulgularımız, akrilamidin oksidatif stresi artırdığını göstermektedir.

Anahtar sözcükler: Akrilamid, Total antioksidan durum, Total oksidan durum, İskemi modifiye albumin, Oksidatif stres

INTRODUCTION

Acrylamide (C₃H₅NO) (ACR) is a small hydrophilic molecule that polymerizes readily in the presence of an initiator because of the double bond between the first and second C-atoms, which makes it a versatile industrial

chemical ^[1]. Polymers of ACR (polyacrylamide) are extensively used in modern chemical technology for a variety of purposes ^[2]. In the last years, relevant amounts of ACR have been identified in heat-treated carbohydrate-rich foods



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such as fried potatoes, cookies, bread, cold breakfast cereal, biscuits and crackers^[3,4]. It was shown that ACR might be formed through the Maillard reaction from amino acids (e.g. asparagine) and reducing sugars (e.g. glucose)^[3]. Maillard reaction is proposed the major pathway for the formation of ACR in food^[5,6]. Several different reaction routes have been proposed for the further reaction to ACR such as decarboxylation of the Schiff base^[6-8], decarboxylated Amadori product^[6,7], formation of a 3-Aminopropionamide^[8,9], Strecker degradation of asparagine^[5].

The biological consequences of ACR exposure have chiefly centered on neurotoxicity ever since this effect was observed in humans occupationally exposed to this compound. Subsequently, experimental exposure of rodents to ACR has also revealed a carcinogenic mode of action for this chemical^[10]. It has been reported that most investigations of ACR toxicity have mainly focused on their genotoxic and carcinogenic properties; however, accumulating evidences seem to indicate that ACR also possesses cytotoxic properties by affecting the redox status of the cells^[11].

Oxidative stress results from an imbalance between the production of free radical and antioxidant activity, leads to damage of biological macromolecules and disruption of normal metabolism and physiology^[12]. Oxidative stress in the cells or tissues refers to enhanced generation of reactive oxygen species (ROS) and/or depletion of antioxidant defense system. ROS can attack the polyunsaturated fatty acid in the biological membranes and induce free radical chain reactions, leading to the enhancement of lipid peroxidation^[13]. The methods used for the determination of oxidative stress are generally of board range. An often used and easily detectable parameter for the serum anti-oxidative and oxidant properties are the total antioxidant status (TAS) and total oxidant status (TOS)^[14,15]. It has been showed that the high concentrations of ACR exert deteriorating effects on antioxidant enzyme activities, lipid peroxidation process and haemolysis in human erythrocytes *in vitro*^[16].

Ischemia-modified albumin (IMA) is a novel marker of tissue ischemia^[17]. IMA is serum albumin in which the N-terminus has been chemically modified^[18]. The diagnostic albumin Co²⁺ binding test is based for IMA on the observation that the affinity of serum albumin for Co²⁺ is reduced after N-terminus modifications^[18]. Proposed mechanisms for the conversion of serum albumin to IMA include hypoxia, acidosis, superoxide-radical injury, energy-dependent membrane disruption and exposure to free iron and copper^[19]. Nowadays, IMA is accepted as a marker of oxidative stress. It has been proposed that ROS such as superoxide (O₂⁻) and hydroxyl (·OH) radicals generated during ischaemia modify the N-terminus of serum albumin resulting in IMA formation^[18].

Studies describing the role of oxidative stress in ACR

toxicity are limited, although the main pathways for metabolism of ACR occur either through conjugation with reduced glutathione (GSH) or via epoxidation to glycidamide^[16]. Also, to our knowledge, the effects of intermediate doses of ACR on serum IMA, TAS and TOS levels in rats have not been investigated yet. Therefore, the present experiment was carried out to investigate the effects of 2 and 5 mg/kg/day doses of ACR on serum IMA, TAS and TOS levels in rats.

MATERIAL and METHODS

Chemicals

ACR (Cas 79-06-1) was received from Sigma Aldrich Chemical Company. The material was a white odourless crystalline solid, with a chemical purity of >99% (for electrophoresis). The test material was prepared weakly and stored at room temperature. All other chemicals used were of analytical grade.

Animals and Husbandry

Male and female weaned Wistar rats, weighing 65-75 g and aged 3-4 weeks old, were obtained from Selcuk University Experimental Medicine Research and Application Centre (Konya, Turkey). The rats were housed in wire-topped opaque polycarbonate cages and maintained under constant environmental conditions with a 12 h light/dark schedule. The environmental temperature was 20°C and humidity was 50%. Commercial food pellets and drinking water were provided *ad libitum*. The protocols of the animal experiments were approved by the internal ethical committee of the Selcuk University (number: 2010-98).

Dosing

We choosed the doses of ACR applied as 2 and 5 mg/kg/day in our study since not to occure a characteristic sign of ACR-induced neurotoxicity such as hindlimb foot splaying^[20,21].

Experimental Design

In total, 25 male and 25 female Wistar rats were involved in this experiment. Animals in each sex were segregated into three groups. Two of them were treatment groups and one of them was control group. Each treatment group consisted of ten animals and each control group consisted of five animals. ACR was administered to the treatment groups at 2 and 5 mg/kg/day via drinking water for 90 days. Tap water was administered to five animals of control group in the same manner as in the treatment groups. After 24 h from the last application all animals were anaesthetized with ketamine in combination with a sedative. Blood samples were collected from the cardiac puncture of all rats under anaesthesia. After collecting blood samples rats were sacrificed by cervical dislocation.

Blood samples were obtained after suitable centrifugation and samples were stored frozen at -80°C until the day of analysis.

Measurement of Total Antioxidant Status

TAS of serum were determined using an automated measurement method which was based on the bleaching of the characteristic color of a more stable 2,2'-azino-bis (3-ethylbenz-thiazoline-6-sulfonic acid) (ABTS) radical cation by antioxidants^[14]. The results were expressed in mmol Trolox equivalents/L.

Measurement of Total Oxidant Status

TOS of serum were determined using a novel automated measurement method.¹⁵ Oxidants present in the sample oxidize the ferrous ion-o-dianisidine complex to ferric ion. The oxidation reaction is enhanced by glycerol molecules, which are abundantly present in the reaction medium. The ferric ion makes a colored complex with xylenol orange in an acidic medium. The color intensity, which can be measured spectrophotometrically, is related to the total amount of oxidant molecules present in the sample. The assay is calibrated with hydrogen peroxide and the results are expressed in terms of micromolar H₂O₂ equivalent per liter (μmol H₂O₂ equiv./L).

Measurement of Ischemia-Modified Albumin Levels

IMA level was measured by a colorimetric assay developed by Bar-Or et al.^[17] based on measurement of unbound cobalt after incubation with patient serum. Increased amounts of IMA results in less cobalt binding and more residual unbound cobalt available for complex with a chromogen [dithiothreitol (DTT)], which can be measured photometrically. The procedure was as follows:

50 μL of 0.1% cobalt chloride was added to 200 μL of serum, gently mixed, and waited 10 min for adequate cobalt-albumin binding. Fifty microliters of DTT, at a

concentration of 1.5 mg/ml, was added as a colorizing agent and the reaction was stopped 2 min later by adding 1.0 mL of 0.9% NaCl. The colored product was measured at 470 nm and compared to a serum-cobalt blank without DTT and reported in absorbance units (ABSU).

Adjusted IMA was calculated as (individual serum albumin concentration/median serum albumin concentration of the population)×IMA ABSU value. This formula was applied to correct IMA values for serum albumin (Median serum albumin concentration of each group of the subjects were used separately)^[22].

Measurement of Albumin Levels

Serum albumin levels were measured by commercially available kits based on routine methods on the Synchron LX System (Beckman Coulter, Fullerton CA).

Statistical Analysis

All data are expressed as mean±standart deviations (SD). Statistical analyses were done using SPSS v. 16.0 (SPSS Inc.,IL, USA). Groups of data were compared with an analysis of variance (ANOVA) followed by Tukey's multiple comparison tests. Differences were considered significant at a probability level of P<0.05.

RESULTS

Serum IMA, adjusted IMA, TAS, TOS and albumin levels of the groups are presented in Table 1. As seen from the table, serum IMA and adjusted IMA levels were significantly higher at concentrations of 5 mg/kg (P<0.001 for IMA and P<0.01 for adjusted IMA) and 2 mg/kg (P<0.01) in the male rats when compared with those of the control male rats. However, there were no significant differences between the same parameters at concentrations of 5 mg/kg and 2 mg/kg in the female rats and the control female rats (Fig. 1). Serum TAS levels significantly decreased at concentrations

Table 1. Serum IMA, adjusted IMA, TAS, TOS and albumin levels of the groups (All values are mean±standart deviations)

Tablo 1. Grupların serum IMA, ayarlanmış IMA, TAS, TOS ve albumin düzeyleri (Bütün değerler ortalama±standart sapma)

Parameter	Group	Control	ACR 2 mg/kg	ACR 5 mg/kg
IMA (ABSU)	Male	0.53±0.09	0.71±0.07 ^b	0.72±0.07 ^a
	Female	0.64±0.09	0.69±0.06	0.73±0.03
Adjusted IMA (ABSU)	Male	0.54±0.09	0.71±0.07 ^b	0.73±0.07 ^b
	Female	0.63±0.1	0.70±0.08	0.73 ± 0.02
TAS (mmol Trolox equivalents/L)	Male	3.65±1.2	3.34 ± 1.2	1.77±1.1 ^c
	Female	3.69±1.5	3.67±1.1	3.31±1.0
TOS (μmol H ₂ O ₂ Eq/L)	Male	8.88±2.3	12.67±2.0	14.72±5.0 ^c
	Female	9.95±1.9	11.18±2.5	14.28±4.1
Albumin (g/dL)	Male	2.72±0.1	2.66±0.2	2.63±0.1
	Female	3.17±0.1	3.06±0.4	2.88±0.3

IMA- ischemia-modified albumin; TAS- total antioxidant status; TOS- total oxidant status; ^aP<0.001, ^bP<0.01, ^cP<0.05 compared with control group

of 5 mg/kg ($P<0.01$) in the male rats when compared with those of the control rats. But, this difference was not found at concentrations of 5 mg/kg in the female rats. On the other hands, there were no significant differences between serum TAS levels at concentrations of 2 mg/kg in the male and female rats and the control rats (Fig. 2). We also observed a significant increase in the levels of serum TOS at concentrations of 5 mg/kg ($P<0.05$) in the male rats. No differences were found in serum TOS levels between at concentrations of 5 mg/kg in the female rats and control rats. Also, no differences were found in serum TOS levels between at concentrations of 2 mg/kg in the male and female rats and control rats (Fig. 3). There were no significant differences between serum albumin levels of the groups.

In addition, no sign of sickness and decreased activity of the rats or mortality was observed during the study.

DISCUSSION

ROS are produced in metabolic and physiological processes, and harmful oxidative reactions may occur in organisms, which remove them via enzymatic and non-enzymatic antioxidative mechanisms. Under some conditions, the increase in oxidants and decrease in anti-oxidants cannot be prevented, and the oxidative/anti-oxidative balance shifts toward the oxidative status^[23]. Several reports have reported that ACR promotes the generation of ROS and the depletion of anti-oxidants^[13,24].

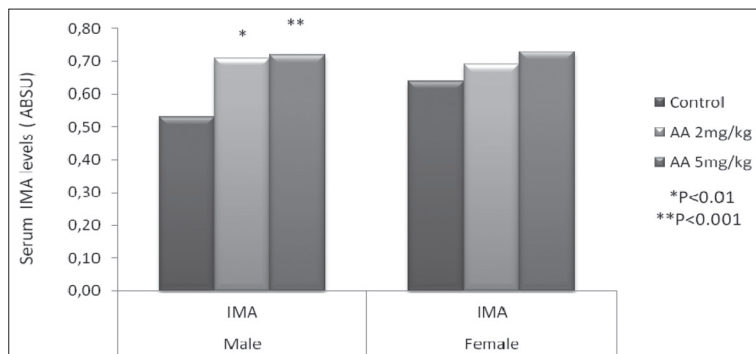


Fig 1. Serum ischemia-modified albumin (IMA) levels in all groups

Şekil 1. Bütün gruplarda serum iskemi-modifiye albumin (IMA) düzeyleri

Fig 2. Serum total antioxidant status (TAS) levels in all groups
Şekil 2. Bütün gruplarda serum total antioksidan durum (TAS) düzeyleri

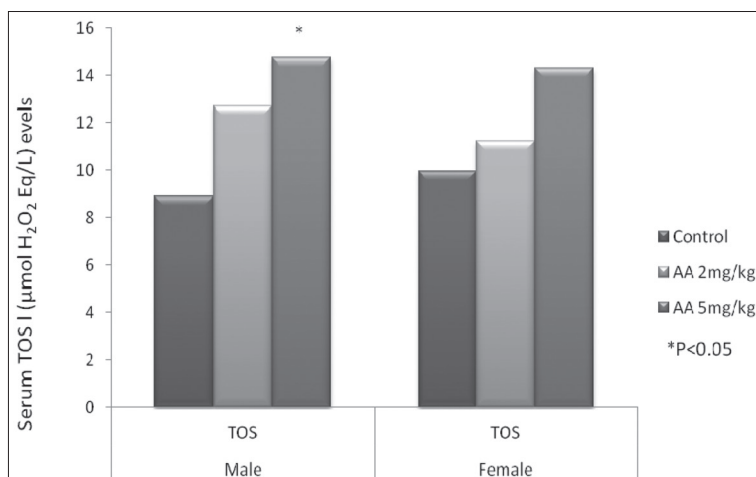
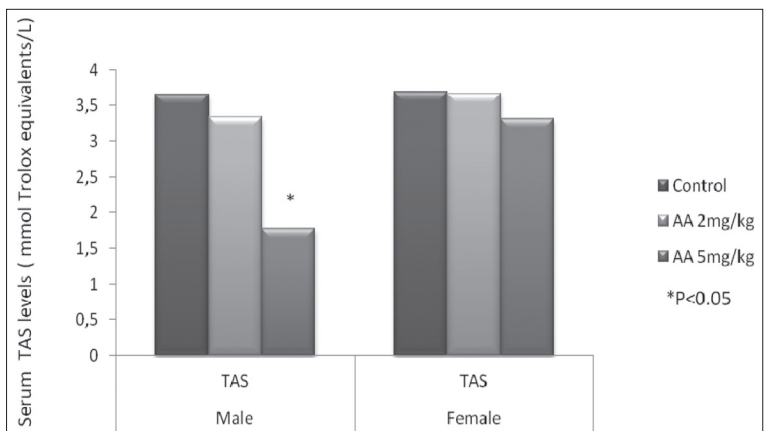


Fig 3. Serum total oxidant status (TOS) levels in all groups

Şekil 3. Bütün gruplarda total antioksidan (TOS) durum düzeyleri

The present study shows that ACR given via drinking water to male rats induced to increase in TOS and IMA levels and decrease in TAS levels.

Conjugation with GSH catalyzed by glutathione-S-transferase is the main pathway of ACR metabolism and detoxification [25]. GSH is one of the essential compounds for the maintenance of the cell integrity because of its reducing properties and participation in the cell metabolism [26]. ACR is capable of interacting with vital cellular nucleophiles possessing $-SH$, $-NH_2$ or $-OH$. Therefore, it reacts with glutathione in a similar manner and forms glutathione S-conjugates, which is the initial step in the biotransformation of electrophiles into mercapturic acids [27]. Decreasing GSH content in the tissues with the increase of ACR concentration could be due to increased formation of S-conjugates between ACR and GSH [28]. ACR can be reactive unlike the above mentioned way. Firstly, ACR can undergo radical-mediated polymerization. Secondly, it can be metabolized to an epoxide derivate, glycidamide, presumably by cytochrome P450E1, being readily reactive toward DNA and other macromolecules [16].

In the literature, our finding supported in studies which made with different oxidative stress marker after ACR-treatment. For example, Allam et al. [29] have reported that ACR administered either prenatally or perinatally has been shown to induce significant increase of thiobarbituric acid reactive substances (TBARS) and oxidative stress (significant reductions in total thiols, superoxide dismutase (SOD) and peroxidase activities) in the developing cerebellum of rats [29]. Alturfan et al. [30] have found that 8-OHdG levels as an indicator of oxidative DNA damage, significantly increased in ACR group. In addition, they demonstrated that in ACR-treated rats, GSH levels decreased significantly while the malondialdehyde (MDA) levels, myeloperoxidase activity and collagen content increased in the tissues suggesting oxidative organ damage. Yousef et al. [28] have found that the concentration of TBARS, and the activities of glutathione S-transferase and SOD in plasma, liver, testis, brain, and kidney were increased in ACR-treated male rats [28]. Catalgol et al. [16] have demonstrated that *in vitro* treatment of ACR results in haemolysis and induction of lipid peroxidation and changes the activities of antioxidant enzymes at high doses in human erythrocytes.

The results of the present study indicate that serum IMA levels increase in ACR-treated rats, compared to control rats. Albumin is the most abundant serum protein, and is a powerful extracellular antioxidant. The biochemical mechanism modifying the N-terminal region of albumin during ischemia is unclear, but reperfusion after an ischemic event may damage serum albumin as much as, if not more than, ischemia itself [31]. These modifications to albumin may involve hypoxia, acidosis, or free radical damage, most of which occur within minutes [23]. According to our hypothesis, IMA is increased in conditions that produce free radicals, such as ACR-treatment. Naruszewicz

et al. [32] have reported that the first time that chronic ingestion of dietary ACR might induce oxidative stress in humans through leukocyte activation and increased production of ROS [32]. In addition, they have reported that a significant increase in the oxidized low-density lipoprotein, interleukin-6, and high sensitivity C-reactive protein concentrations occurred, with all these factors recognized as capable of enhancing atherosclerosis progression [32]. According to their hypothesis, long-term ingestion of high ACR doses with food may cause chronic inflammation and contribute to the development of early atherosclerosis as well as increase the risk of coronary artery disease. IMA was considered to be related as a potential cardiovascular risk factor [18]. Previous studies have found that IMA levels increased in acute coronary syndromes, during percutaneous coronary intervention, and myocardial ischemia [17,33,34]. Our finding supports Naruszewicz et al.'s [32] hypothesis. Because, long term intermediate doses ACR-treatment, which increased serum IMA and TOS levels, may be risk for cardiovascular diseases.

On the other hand, a significant increase serum IMA and TOS levels and decrease TAS levels did not observed in female rats. We think that estrogen may be cause this situation. The hormone estrogen, (estradiol) provides natural antioxidant properties because it increases the expression of antioxidant enzymes and decreases troublesome NADPH oxidase enzyme activity and superoxide production. Estrogens have a phenolic hydroxyl group at position 3 and a methyl group at position 13. The presence of this phenol group gives estrogen its antioxidant property by neutralizing oxygen free radicals. The estrogens estriol and estradiol confer significant antioxidant activity [35].

In our study, ACR administered rats grew normally and gained body weight over a 12-week period, and there were no significant differences in the average body weight compared with control rats. This finding is in accordance with that of Park et al. [36]. However, previous studies indicated that rats exposed to ACR showed a decrease in body weight [37,38]. According to our hypothesis, there were no changes in body weight in intermediate doses.

As a result, our findings show that long-term treatment with intermediate-ACR doses led to a significant depletion of serum TAS levels and overproduction of serum TOS and IMA levels especially in male rats, and cause oxidative stress.

REFERENCES

1. Hogervorst JG, Schouten LJ, Konings EJ, Goldbohm RA, van den Brandt PA: Dietary acrylamide intake and the risk of renal cell, bladder, and prostate cancer. *Am J Clin Nutr*, 87 (5): 1428-1438, 2008.
2. Dearfield KL, Abernathy CO, Ottley MS, Brantner JH, Hayes PF: Acrylamide: Its metabolism, developmental and reproductive effects, genotoxicity, and carcinogenicity. *Mutat Res*, 195 (1): 45-77, 1988.
3. Tareke E, Rydberg P, Karlsson P, Eriksson S, Törnqvist M: Analysis of acrylamide, a carcinogen formed in heated foodstuffs. *J Agric Food Chem*,

50 (17): 4998-5006, 2002.

4. Becalski A, Lau BP, Lewis D, Seaman SW, Hayward S, Sahagian M, Ramesh M, Leclerc Y: Acrylamide in French fries: Influence of free amino acids and sugars. *J Agric Food Chem*, 52 (12): 3801-3806, 2004.

5. Mottram DS, Wedzicha BL, Dodson AT: Acrylamide is formed in the Maillard reaction. *Nature*, 419, 448-449, 2002.

6. Stadler RH, Robert F, Riediker S, Varga N, Davidek T, Devaud S, Goldmann T, Hau J, Blank I: In-depth mechanistic study on the formation of acrylamide and other vinyllogous compounds by the Maillard reaction. *J Agr Food Chem*, 52, 5550-5558, 2004.

7. Yaylayan VA, Wnorowski A, Locas CP: Why asparagine needs carbohydrates to generate acrylamide. *J Agr Food Chem*, 51, 1753-1757, 2003.

8. Zyzak DV, Sanders RA, Stojanovic M, Tallmadge DH, Eberhart BL, Ewald DK, Gruber DC, Morsch TR, Strothers MA, Rizzi GP, Villagran MD: Acrylamide formation mechanism in heated foods. *J Agr Food Chem*, 51, 4782-4787, 2003.

9. Granvogl M, Jezussek M, Koehler P, Schieberle P: Quantitation of 3-aminopropionamide in potatoes - A minor but potent precursor in acrylamide formation. *J Agr Food Chem*, 52, 4751-4757, 2004.

10. Besaratinia A, Pfeifer GP: A review of mechanisms of acrylamide carcinogenicity. *Carcinogenesis*, 28 (3): 519-528, 2007.

11. Rodríguez-Ramiro I, Martín MÁ, Ramos S, Bravo L, Goya L: Olive oil hydroxytyrosol reduces toxicity evoked by acrylamide in human Caco-2 cells by preventing oxidative stress. *Toxicology*, 288 (1-3): 43-48, 2011.

12. Trevisan M, Browne R, Ram M, Muti P, Freudenheim J, Carosella AM, Armstrong D: Correlates of markers of oxidative status in the general population. *Am J Epidemiol*, 154 (4): 348-356, 2001.

13. Zhu YJ, Zeng T, Zhu YB, Yu SF, Wang QS, Zhang LP, Guo X, Xie KQ: Effects of acrylamide on the nervous tissue antioxidant system and sciatic nerve electrophysiology in the rat. *Neurochem Res*, 33 (11): 2310-2317, 2008.

14. Erel O: A novel automated method to measure total antioxidant response against potent free radical reactions. *Clin Biochem*, 37 (2): 112-119, 2004.

15. Erel O: A new automated colorimetric method for measuring total oxidant status. *Clin Biochem*, 38 (12): 1103-1111, 2005.

16. Catalgol B, Ozhan G, Alpertunga B: Acrylamide-induced oxidative stress in human erythrocytes. *Hum Exp Toxicol*, 28 (10): 611-617, 2009.

17. Bar-Or D, Lau E, Winkler JV: A novel assay for cobalt-albumin binding and its potential as marker its potential as a marker for myocardial ischemia-A preliminary report. *J Emer Med*, 19, 311-315, 2000.

18. Roy D, Quiles J, Gaze DC, Collinson P, Kaski JC, Baxter GF: Role of reactive oxygen species on the formation of the novel diagnostic marker ischaemia modified albumin. *Heart*, 92 (1): 113-114, 2006.

19. Sbarouni E, Georgiadou P, Voudris V: Ischemia modified albumin changes - review and clinical implications. *Clin Chem Lab Med*, 49, 177-184, 2011.

20. Burek JD, Albee RR, Beyer JE, Bell TJ, Carreon RM, Morden DC, Wade CE, Hermann EA, Gorzinski SJ: Subchronic toxicity of acrylamide administered to rats in drinking water followed by up to 144 days of recovery. *J Environ Pathol Toxicol*, 4, 157-182, 1980.

21. Smith MK, Zenick H, Preston RJ, George EL, Long RE: Dominant lethal effects of subchronic acrylamide administration in the male Long-Evans rat. *Mutat Res*, 173, 273-277, 1986.

22. Lippi G, Montagnana M, Salvagno GL, Guidi GC: Standardization of ischaemia-modified albumin testing: Adjustment for serum albumin. *Clin Chem Lab Med*, 45 (2): 261-262, 2007.

23. Gutteridge JM, Halliwell B: Free radicals and antioxidants in the year 2000. A historical look to the future. *Ann N Y Acad Sci*, 899, 136-147, 2000.

24. Jiang L, Cao J, An Y, Geng C, Qu S, Jiang L, Zhong L: Genotoxicity of acrylamide in human hepatoma G2 (HepG2) cells. *Toxicol In Vitro*, 21 (8): 1486-1492, 2007.

25. Odland L, Romert L, Clemenson C, Walum E: Glutathione content, glutathione transferase activity and lipid peroxidation in acrylamide-treated neuroblastoma N1E 115 cells. *Toxicol In Vitro*, 8 (2): 263-267, 1994.

26. Conklin KA: Dietary antioxidants during cancer chemotherapy: Impact on chemotherapeutic effectiveness and development of side effects. *Nutr Cancer*, 37 (1): 1-18, 2000.

27. Awad ME, Abdel-Rahman MS, Hassan SA: Acrylamide toxicity in isolated rat hepatocytes. *Toxicol In Vitro*, 12 (6): 699-704, 1998.

28. Yousef MI, El-Demerdash FM: Acrylamide-induced oxidative stress and biochemical perturbations in rats. *Toxicology*, 219 (1-3): 133-141, 2006.

29. Allam A, El-Ghareeb AA, Abdul-Hamid M, Baikry A, Sabri MI: Prenatal and perinatal acrylamide disrupts the development of cerebellum in rat: Biochemical and morphological studies. *Toxicol Ind Health*, 27 (4): 291-306, 2011.

30. Alturfan AA, Tozan-Beceran A, Sehirli AO, Demiralp E, Sener G, Omurtag GZ: Resveratrol ameliorates oxidative DNA damage and protects against acrylamide-induced oxidative stress in rats. *Mol Biol Rep*, 39 (4): 4589-4596, 2011;

31. Edwards SW, Hallett MB, Campbell AK: Oxygen-radical production during inflammation may be limited by oxygen concentration. *Biochem J*, 217, 851-854, 1984.

32. Naruszewicz M, Zapolska-Downar D, Kośmider A, Nowicka G, Kozłowska-Wojciechowska M, Vikström AS, Törnqvist M: Chronic intake of potato chips in humans increases the production of reactive oxygen radicals by leukocytes and increases plasma C-reactive protein: a pilot study. *Am J Clin Nutr*, 89 (3): 773-777, 2009.

33. Sinha MK, Gaze DC, Tippins JR, Collinson PO, Kaski JC: Ischemia-modified albumin is a sensitive marker of myocardial ischemia after percutaneous coronary intervention. *Circulation*, 107, 2403-2405, 2003.

34. Sinha MK, Roy D, Gaze DC, Collinson PO, Kaski JC: Role of "ischemia-modified albumin" a new biochemical marker of myocardial ischemia, in the early diagnosis of acute coronary syndromes. *Emerg Med J*, 21, 29-34, 2004.

35. Pialoux V, Brown AD, Leigh R, Friedenreich CM, Poulin MJ: Effect of cardiorespiratory fitness on vascular regulation and oxidative stress in postmenopausal women. *Hypertension*, 54 (5): 1014-1020, 2009.

36. Park HR, Kim MS, Kim SJ, Park M, Kong KH, Kim HS, Kwack SJ, Kang TS, Kim SH, Kim HS, Lee J: Acrylamide induces cell death in neural progenitor cells and impairs hippocampal neurogenesis. *Toxicol Lett*, 193 (1): 86-93, 2010.

37. Bowyer JF, Latendresse JR, Delongchamp RR, Warbritton AR, Thomas M, Divine B, Doerge DR: The mRNA expression and histological integrity in rat forebrain motor and sensory regions are minimally affected by acrylamide exposure through drinking water. *Toxicol Appl Pharmacol*, 240 (3): 401-411, 2009.

38. Raju J, Sondagar C, Roberts J, Aziz SA, Caldwell D, Vavasour E, Mehta R: Dietary acrylamide does not increase colon aberrant crypt foci formation in male F344 rats. *Food Chem Toxicol*, 49 (6): 1373-1380, 2011.

Effect of Hyaluronic Acid/Carboxymethylcellulose and Flunixin Meglumine Combination on the Prevention of Postoperative Intraabdominal Adhesions: An Experimental Study in Rabbits ^[1]

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Summary

Despite the development in the knowledge concerning pathophysiology of abdominal cavity and improvement in surgical techniques, intraabdominal adhesions are one of the most common complications in surgery. The aim of this study was to determine the effects of hyaluronic acid/carboxymethylcellulose (HA/CMC) and flunixin meglumine combination on the prevention of postoperative intraabdominal adhesions in rabbits. All the surgery were made under aseptic condition under general anesthesia. A 2x2 cm area of parietal peritoneum was resected on the abdominal wall after median laparotomy. Twenty-four adult, male rabbits were divided into four groups equally. Groups were evaluated as C (control), S (HA/CMC), F (flunixin meglumine), and SF (HA/CMC and flunixin meglumine combination). No medication was given to the rabbits in group C. In S and SF groups, 3x3 cm of HA/CMC applied directly to peritoneal defect. In F and SF groups, flunixin meglumine was administered 1.1 mg/kg for 5 days intraperitoneally. Relaparotomy was performed 10 days after surgery. As a result of macroscopical and histopathological evaluations, the scores of adhesion in the S, F, and SF groups were significantly lower than those of the control group ($P<0.05$). Adhesion scores in SF group were lower as compared to S and F groups ($P<0.05$). It is observed that HA/CMC and flunixin meglumine combination can be used more effective on the prevention of postoperative intraabdominal adhesions in rabbits.

Keywords: Intraabdominal adhesion, Hyaluronic acid/carboxymethylcellulose, Flunixin meglumine, Rabbit


Postoperatif İntraabdominal Adezyonların Önlenmesinde Hyaluronik Asit/Karboksimetilselüloz ve Fluniksin Meglumin Kombinasyonu'nun Etkisi: Tavşanlarda Deneysel Bir Çalışma

Özet

Abdominal boşluğun patofizyolojisi konusunda bilgilerin artmasına ve cerrahi tekniklerin gelişmesine rağmen intraabdominal adezyonlar, cerrahideki en önemli sorunlardan biridir. Bu çalışmada, tavşanlarda postoperatif intraabdominal adezyonların önlenmesinde hyalüronik asit/karboksimetilselüloz (HA/KMS) ve fluniksin meglumine kombinasyonunun etkisinin değerlendirilmesi amaçlandı. Tüm operasyonlar genel anestezi eşliğinde aseptik koşullar altında yapıldı. Median laparotomiden sonra abdominal duvar üzerinden 2x2 cm'lik bir parietal periton alanı rezeke edildi. Çalışmada 24 adet yetişkin erkek tavşan 4 gruba eşit olarak ayrıldı. Gruplar; C (kontrol), S (HA/KMS), F (fluniksin meglumine), ve SF (HA/KMS ve fluniksin meglumine kombinasyonu) olarak değerlendirildi. Kontrol grubundaki olgulara herhangi bir tedavi uygulanmadı. S ve SF gruplarındaki olguların defektli alanları 3x3 cm'lik HA/KMS ile örtüldü. F ve SF gruplarına, fluniksin meglumine 1.1 mg/kg dozunda intraperitoneal olarak 5 gün süre ile verildi. Postoperatif 10. günde tekrar laparotomi yapıldı. Makroskopik ve histopatolojik değerlendirmeler sonucunda; S, F, and SF gruplarındaki adezyon skorları kontrol grubundakine oranla önemli derecede düşük bulundu ($P<0.05$). SF grubundaki adezyon skorları, S ve F grupları ile karşılaştırıldığında daha düşük olduğu saptandı ($P<0.05$). Tavşanlarda postoperatif intraabdominal adezyonların önlenmesinde HA/KMS ve fluniksin meglumine kombinasyonun daha etkili olduğu gözlemlendi.

Anahtar sözcükler: İntraabdominal adezyon, Hyalüronik asit/karboksimetilselüloz, Fluniksin meglumine, Tavşan

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INTRODUCTION

Adhesions are fibrous bands that cause separate organs to adhere to one another and can be classified into congenital, inflammatory, and postoperative. Postoperative adhesions are considered to be an inevitable result of surgery. The incidence of postoperative adhesion ranges from 66 to 93% after operations [1-3]. Surgical trauma, tissue ischemia, hemorrhage, infection, and foreign materials play an important role in etiology of postoperative adhesions. Postoperative intraabdominal adhesions cause chronic abdominal/pelvic pain, female infertility, and small bowel obstruction. Also, fibrous adhesions make prolong operation time, operation difficulty, increased bleeding, intestinal rupture, and morbidity in repeated operations [1-5].

When there is an injury in mesothelial surface of the peritoneum, it triggers an inflammatory reaction characterized by increased neutrophil leucocytes, fibroblasts, macrophages, and mesothelial cells. There is increase in vascular permeability and release of fibrin-rich exudate. Peritoneal mesothelial cells have a critical duty in maintaining the intraperitoneal balance between fibrinolysis and formation. Plasminogen is an important protein in the process of fibrinolysis. Early fibrinolysis, within 5 days, encourages healing of the peritoneum without adhesion formation. Inadequate fibrinolysis processes lead to persistent fibrin structures that subsequently mature into fibrous tissue, followed by organization into rigid, persistent fibrous adhesions containing blood vessels and nerve fibers [1-6].

Many different techniques, agents, and materials have been tried to reduce adhesions [1-3,6-9]. Several pharmacological drugs including corticosteroid, non-steroid antiinflammatory drugs (NSAIDs), melatonin, progesterone, aprotinin, methylen blue, heparin, mitomycin C, vitamin E, and tPA have been used to prevent adhesions [1,2,8,10,11]. NSAIDs inhibit prostaglandin and thromboxane synthesis by changing cyclooxygenase activities. Flunixin meglumine (non-selective COX inhibitors) are commonly used to treat animals suffering from various diseases and has been proved to prevent the formation of peritoneal adhesions in studies [1,2,7,9-12]. Mechanical barriers such as carboxymethylcellulose, hyaluronic acid, icodextrin, polyethylene glycol, fibrin glue, oxidized regenerated cellulose, expanded polytetrafluoro ethylene, and hyaluronic acid/carboxymethylcellulose (HA/CMC) have been applied to prevent adhesion by mechanically minimizing the development of fibrin between serosal surfaces [1,2,6,13]. HA/CMC is a biosynthetic material that have been widely used in human and animals. In developed countries, HA/CMC is accepted as an important antiadhesive barrier. Unfortunately, none of these treatments is ideal [1-3,5,8,9,14-16].

Although several substances have been tried to eliminate adhesions, these have had a limited success in the

intraabdominal adhesions. A current interest to overcome adhesions have been focused on various combination of drugs [2,4,8,9,17-23]. Therefore, it was to investigate the effect of HA/CMC, flunixin meglumine and their combination on the prevention of postoperative intraabdominal adhesions in rabbits.

MATERIAL and METHODS

Animals

Twenty-four rabbits weighing approximately 2.8-3.6 kg were used in this study. The rabbits were hospitalized for 2 weeks with a deprivation of food for 12 h immediately before surgery. The animals were kept under standart conditions and had free access to water and pellets. The study protocol was approved by Animal Ethic Committee of the Veterinary Faculty, Firat University (2006/7).

Anesthesia

The animals were anesthetized with 10 mg/kg of xylazine HCl (Rompun®, Bayer, İstanbul, Turkey) and 40 mg/kg of ketamine HCl (Ketalar®, Eczacıbaşı, İstanbul, Turkey) intramuscularly.

Surgical Procedure

All the surgery were performed under aseptic conditions, including an iodine scrub, drapping and use of a steril technique. Powder-free gloves were used for the operation. Following complete aseptic preparations, a 5-6 cm of incision was performed for midline laparotomy. A 2x2 cm area of parietal peritoneum was removed on the right abdominal wall. The animals were divided into four groups, each containing 6 rabbits: group C (control), group S (HA/CMC), group F (flunixin meglumine), and group SF (HA/CMC and flunixin meglumine combination). No medication was applied to the control group. HA/CMC (Seprafilm®, Genzyme, USA) 3 cm x 3 cm were placed on the deperitonized area of abdominal wall in S and SF groups. Flunixin meglumine (Finadyn®, Eczacıbaşı, İstanbul) was given 1.1 ml/kg for 5 days in F and SF groups intraperitoneally. The abdomen was closed with continuous sutures with 2/0, and skin was closed with interrupted 2/0 polypropylene sutures (Monofilament polyglytone, Caprosyn™, USA).

All animals were sacrificed on the 10th days after operation. After performing a U-type incision to the anterior abdominal wall, the evaluation was macroscopically classified according to adhesion scoring system described by the Majuzi and colleagues [24]. The degree of adhesion formation was evaluated with the following adhesion scores: 0=no adhesion, 1=light adhesions can be easily removed through blunt dissection, 2=moderate adhesions which need sharp dissections (lower than 50%) for the separation, 3=severe adhesions which need

sharp dissections (more than 50%) for the separation, 4=presence of serosal injury, 5=presence of full-thickness injury.

Histopathological Evaluations

Peritoneal tissue samples containing adhesion were excised and the samples were fixed in 10% formaldehyde solution. After dehydration, the specimens were embedded in paraffin blocks. Then, 5 µm sections in thickness were prepared using a microtome and stained with Hematoxylin and Eosin (H&E). Histopathological evaluations were performed under a light microscope by one pathologist who was blind to groups according to semiquantitative scoring system described by Hooker and colleagues [25]. The histopathological adhesions were categorized as Grades 0–III based on the presence and extent of fibrosis and inflammation. Grade 0 was defined as no fibrosis, grade I as mild fibrosis, grade II as moderate fibrosis, and grade III as severe fibrosis. The grade of inflammation was assessed according to grade I on this scale represents a mild inflammatory reaction with giant cells, occasionally scattered lymphocytes, and plasma cells, grade II represents moderate reaction with giant cells and increased admixed

lymphocytes, neutrophils, eosinophils and plasma cells, and grade III represents a severe inflammatory reaction with microabscesses present.

Statistical Analysis

The group data were analyzed using the Kruskal-Wallis nonparametric test. Differences among the groups were evaluated using Mann-Whitney U test. Differences with a value of $P < 0.05$ were considered to be statistically significant. Data were analyzed by SPSS software program (Version 14.0; SPSS Inc, Chicago IL).

RESULTS

Most of the animals tolerated the surgery well. No death was observed throughout the study period. There was no evidence of any of complication in the animals. The results of macroscopic and histopathologic evaluation are summarized in Table 1.

There were no adhesions in Grade 0, 1 or 2 in the control group. The adhesions in the control group were mostly Grade 3 or 4. In one case in control group, adhesion

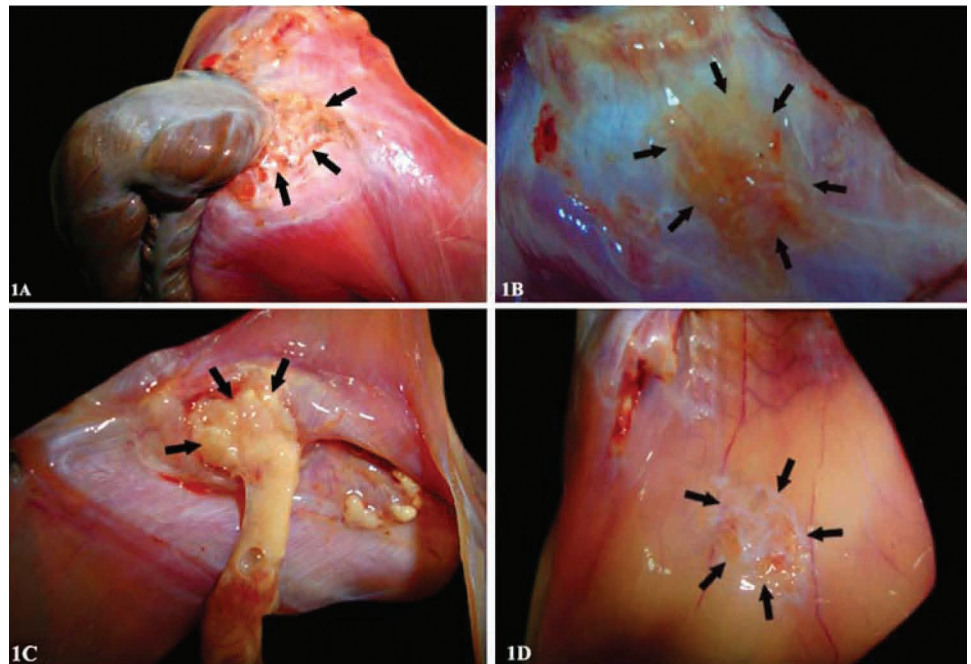
Table 1. The statistical results of adhesion, fibrosis, and inflammation scores

Tablo 1. Adezyon, fibrozis ve yangı skorlarının istatistiksel sonuçları

Parameters	Groups				P value
	C	S	F	SF	
Macroscopic	3.56±0.89 ^a	1.83±0.75 ^b	2.00±0.89 ^b	1.05±0.89 ^c	0.002
Fibrosis	2.50±0.55 ^a	1.50±0.55 ^b	1.33±0.52 ^b	1.18±0.00 ^b	0.003
Inflammation	2.50±0.55 ^a	1.52±0.55 ^{bc}	1.23±0.52 ^b	0.93±0.41 ^c	0.002

Superscripts a-c indicate that means in the same line without a common superscript differ significantly ($P < 0.05$)

Fig 1. Macroscopical evaluations: A) Presence of full-thickness injury and large area of fibrosis (arrows) with score 5 in C group, B) Slight adhesion (arrows) with score 1 in S group, C) Severe adhesion which need sharp dissections (more than 50%) for the separation with score 3 in F group, D) No adhesion occurrence with score 0 in SF group



Şekil 1. Makroskobik değerlendirmeler. A) C grubundaki 5. derece adezyonlu geniş fibrozis (oklar) ve tam kat adezyon varlığı, B) S grubundaki 1. dereceli hafif adezyon (oklar) görünümü, C) F grubundaki 3. derecedeki ayrılma için keskin diseksiyona (%50 den daha fazla) ihtiyaç gösteren ciddi adezyonun görünümü, D) SF grubundaki adezyonun yokluğu (0 derece)

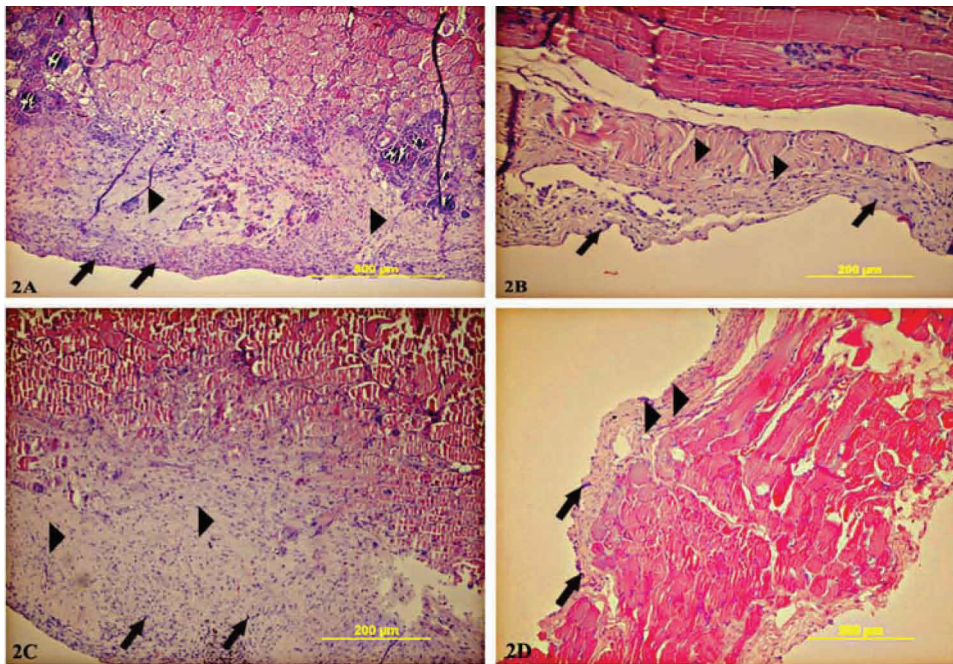


Fig 2. Histopathological evaluations. A) Severe fibrosis (arrow heads) and inflammatory changes (arrows) in C group, Grade III, B) Mild fibrosis (arrow heads) and mild inflammatory changes (arrows) S group, Grade 1, C) Severe fibrosis (arrow heads) and inflammatory changes (arrows) in F group, Grade III, D) Mild fibrosis (arrow heads) and no inflammatory changes (arrows) in SF group

Şekil 2. Histopatolojik değerlendirmeler. A) C grubundaki şiddetli fibrozis (ok başı) ve yangısal değişiklikler (ok), B) S grubundaki hafif fibrozis (ok başı) ve yangısal değişiklikler (ok), C) F grubundaki şiddetli fibrozis (ok başı) ve yangısal değişiklikler (ok), D) SF grubundaki hafif derecedeki fibrozis (ok başı) ve yangısal bulgu yokluğu (ok)

in Grade 5 was recorded (Fig. 1-A). On the other hand, there were no adhesions in Grade 4 or 5 in the S, F, and SF groups. Most adhesion scores in the S and F groups were grade 1, 2, and 3 (Fig. 1-B-C). Adhesion scores in the SF group were seen 2 cases in Grade 1 and 2. No adhesion was noted in 2 cases in the SF group (Fig. 1-D). Statistical analysis indicated that adhesion scores were significantly lower in S, F, and SF groups when compared to the control group ($P < 0.05$). A statistically significant difference was not observed between groups S and F ($P > 0.05$). Group SF showed the lowest incidence of adhesions as compared to S and F groups macroscopically ($P < 0.05$).

Severe fibrosis and inflammatory changes were seen in control group (Fig. 2-A). Minimal fibrosis and low degree of inflammation were observed in the S, F, and SF groups (Fig. 2 B-C). Statistical evaluation of fibrosis scores showed significant difference in S, F, and SF groups when compared to the control group ($P < 0.05$). On the other hand, there was not significant difference between scores of fibrosis in SF group compared to that of S and F groups ($P > 0.05$). However, there was a significant difference between the degrees of adhesion and inflammatory changes in SF group compared to S group ($P < 0.05$), (Fig. 2-D).

DISCUSSION

Despite the development in the knowledge concerning pathophysiology of abdominal cavity and improvement in surgical techniques, postoperative intraabdominal adhesions are significant health problem with major adverse effects on quality of life, use of health care resources, and financial costs after laparotomy [1-4,7-9,18,21].

There are many experimental models for the

development of inflammatory and postoperative adhesions including the local peritoneal model, the damaged uterine horn, introduction of foreign objects in the abdominal and pelvic cavity, the direct mechanical intestinal damage, the ileal transection, the thermal damage, the large bowel anastomosis, the bacterial contamination, and the scraping models [16,17,19,20,22]. In this study, the parietal peritoneal model was chosen due to the similarities of the physiopathology after abdominal surgery. Since, it was observed that all cases in the control group occurred different degrees of adhesion.

Various studies have pointed out that the adhesion-forming process and mesothelization are completed after 7 to 14 days following surgery [10,17,23,26]. Therefore, it was chosen to wait 10 days before performing relaparotomy because other studies had shown high adhesion scores at the period of this time [4,5,27]. There have many adhesion scoring systems described by researchers in experimental models [4-6,16,17,22]. Macroscopical and histopathological evaluations were made according to semiquantitative scoring systems that had been described by Mazuji et al. [24] and Hooker et al. [25]. To date mentioned evaluations which have been used by many reseachers [4,22,23] is simple and suitable scoring systems for use in experimental studies.

The peritoneal healing process differs from skin healing process, whose differences actually came from the reepithelization, and consequences of fibrin depositions. Intraabdominal adhesions are formed as a result of tissue inflammation, fibrin deposition, fibrin organization, collagen formation and maturation [2,3,6,7]. Surgical trauma initiates a widespread inflammatory response in the peritoneum that is associated with the recruitment and activation of inflammatory cells, and the secretion of proinflammatory

mediators. Peritoneal inflammation leads to the formation of an inflammatory exudate. The increase of vessel permeability produces an outpouring of serosanguineous exudate rich in inflammatory cells. These cells promote the formation of a fibrin-rich matrix at the sites of peritoneal injury that leads to the formation of fibrinous adhesions and eventually permanent adhesions [1,3,7,22].

It is important to use minimal invasive surgery techniques and various agents to diminish the post-operative adhesions. These techniques and equipments have focused mainly on limiting tissue injury, like keeping the tissues moist, removing of foreign materials, avoiding of desiccation/ischemia, and using of laparoscopy. The roles of pharmacological drugs are diminishing inflammatory response, enhancing fibrinolysis, preventing collagen deposition and maturation [1-4,6]. NSAIDs decrease vascular permeability, platelet aggregation, plasmin inhibitor and coagulation, and enhance macrophage function [1,2,7]. Some researchers [7,10,11,22] have demonstrated that NSAIDs have been used to prevent the adhesion formations by blocking the initial stage of inflammatory response. An optimal barrier should be non-toxic, biodegradable, easily applicable, without compromising wound healing, remain active in the presence of body fluids, and dissolved after 1 to 2 weeks. HA/CMC has been used to reduce adhesion formation after the surgery. It becomes a hydrophilic gel within approximately 24 hours after placement. It is absorbed within 7 days and cleared completely from the body within 28 days [1-4,14-16]. In our study, it was used the effect of flunixin meglumine 1.1 mg/kg on the postoperative adhesions and HA/CMC covered on the peritoneal defect. These results, as expected under guidance of previous result of the studies [7,10,11,14-18], have supported that both HA/CMC and flunixin meglumine decreases adhesion formation, as evaluated by the scoring of adhesion, fibrosis, and inflammatory compared with the control group ($P<0.05$), (Table 1).

The physical barriers should focus on more widespread adhesion prevention without compromising peritoneal fibrinolytic activity. HA/CMC is accepted as one of the most effective barrier to prevent intraabdominal adhesions. However, their efficacy may be limited to the site of application. HA/CMC does not address the complex nature of adhesion formation and has hydroflotation and slonization effect on the applicated area [6-8,9,21,28]. An current approach to increase the efficacy of barriers or drugs could be improved the synergistic effect of various combinations without causing significant complications [4,9,19-23,26,28]. Lim et al. [8] found that the coadministration of HA/CMC with neurokinin 1 receptor antagonist not only increases the efficacy of the barrier at the site of application, but also significantly reduces the adhesions formation at distal unprotected sites. Attar et al. [18] have reported that combined use of HA/CMC and melatonin were found to be effective reducing the adhesion formation in a

rat uterine horn model. In this study, adhesion scores of the HA/CMC and melatonin groups were similar, and significantly lower than those of the control group. Also, adhesion scores in the combination group were lower than those in the control, HA/CMC, and melatonin groups. Altun et al. [23] recently showed that low molecular weight heparin and octreotide were almost have the same effect for prevention of adhesion formation. In this work, it was observed that the scores of adhesion and inflammation in SF group were significantly lower than that in S and F groups ($P<0.05$), (Table 1). So that based on this results, it was concluded that they were effective in combination rather than the separate using. The result of HA/CMC and flunixin meglumine combination in this study was parallel to those of other combination [8,9,18,23,26].

Numerous articles on the combination of some agents used adhesion prevention have been published but knowledge on this subject is limited and contradictory [17,18,29-31]. Şahin and Sağlam [19] researched heparin to the carboxymethylcellulose at laparotomy, whereas Başbuğ et al. [20] added hyaluronic acid plus heparin, Tayyar et al. [26] tried heparin to the amniotic membrane to cover injured rabbit uterine horns. These combinations seem to be more effective in reducing adhesion formation rather than the use of heparin, hyaluronic acid, carboxymethylcellulose and amniotic membrane alone. Conversely, some researchers [17,27,29-32] revealed that there was no difference between the groups in the combined application of various drugs (HA/CMC+atorvastatin, dimethyl sulfoxide+synovial fluid, low molecular weight heparin+hyperbaric oxygen, medroxyprogesterone acetate+heparin, HA/CMC+vitamin E, vitamin E+selenium) on the prevention of intra-abdominal adhesions.

In conclusion, HA/CMC and flunixin meglumine reduce significantly the prevalence of adhesion development separately. However, HA/CMC and flunixin meglumine combination have reduced more effectively the formation of postoperative intraabdominal adhesions. Further and more detailed investigations could be useful and necessary using additional examinations.

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REFERENCES

1. Arung W, Meurisse M, Detry O: Pathophysiology and prevention of postoperative peritoneal adhesions. *World J Gastroenterol*, 17 (41): 4545-4553, 2011.
2. Ergül E, Korukluoğlu B: Peritoneal adhesions: Facing the enemy. *Int J Surg*, 6, 253-260, 2008.
3. Rizo A, Spedicato M, Mutinati M, Minoia G, Angioni S, Jirillo F, Pantaleo M, Sciorci RL: Peritoneal adhesions in human and veterinary

medicine: From pathogenesis to therapy. A review. *Immunopharm Immunot*, 32 (3): 481-494, 2010.

4. İrkorucu O, Ferahköşe Z, Memiş L, Ekinci Ö, Akın M: Reduction of postsurgical adhesions in a rat model: A comparative study. *Clinics*, 64 (2): 143-148, 2009.

5. Emre A, Akın M, Işıkgönül İ, Yüksel O, Anadol AZ, Çifter Ç: Comparison of intraperitoneal honey and sodium hyaluronate-carboxymethylcellulose (Seprafilm™) for the prevention of postoperative intra-abdominal adhesions. *Clinics*, 64 (4): 363-368, 2009.

6. Tarhan ÖR, Eroğlu A, Çetin R, İnce A, Bülbül M, Altuntaş İ: Effects of seprafilm on peritoneal fibrinolytic system. *ANZ J Surg*, 75 (8): 690-692, 2005.

7. Ezberçi F, Bülbüloğlu E, Çıraçıl P, Gül M, Kurutaş EB, Bozkurt S, Kale İT: Intraperitoneal tenoxicam to prevent abdominal adhesion formation in a rat peritonitis model. *Surg Today*, 36 (4): 361-366, 2006.

8. Lim R, Stucchi AF, Morrill JM, Reed KL, Lunch R, Becker JM: The efficacy of a hyaluronate-carboxymethylcellulose bioresorbable membrane that reduces postoperative adhesions is increased by the intra-operative co-administration of a neurokinin 1 receptor antagonist in a rat model. *Surgery*, 148 (5): 991-999, 2010.

9. Buckenmaier CC, Pusateri AE, Harris RA, Hetz SP: Comparison of antiadhesive treatments using an objective rat model. *Am Surgeon*, 65 (3): 274-282, 1999.

10. Alkan F, Koç Y, Çelik İ, Erol M, Aydın MF: An experimental study evaluating on the effects of flunixin meglumine and mepiramine maleate in the prevention of intraabdominal adhesions in rabbits. *Vet Bil Derg*, 23 (1): 41-46, 2007.

11. Köklü S: Postoperatif intraabdominal adezyonların önlenmesinde non-steroid antienflamatuar grubu bazı ilaçların etkilerinin değerlendirilmesi. *Yüksek Lisans Tezi*, Adnan Menderes Üniv. Sağlık Bil. Enst., 2008.

12. Özyayın İ, Okumuş Z, Maraşlı Ş, Cihan M, Acarer N, Çelebi F: Clinical and biochemical effects of local flunixin meglumine on the treatment of arthritis, bursitis and tenosynovitis in cattle. *Kafkas Üniv Vet Fak Derg*, 1 (1-2): 68-74, 1995.

13. Belge A, Gülbahar MY, Bakır B: The effects of fibrin glue on the prevention of adhesion formation, leakage and wound healing in the dog gaster model. *Indian Vet J*, 82, 36-39, 2005.

14. Beck DE: The role of Seprafilm bioresorbable membrane in adhesion prevention. *Eur J Surg Suppl*, 577, 49-55, 1997.

15. Günerhan Y, Çağlayan K, Sümer A, Köksal N, Altınlı E, Onur E, Güneş P, Gümrükçü G: The efficacy of carboxymethylcellulose for prevention adhesion formation after thyroid region surgery. *Kafkas Üniv Vet Fak Derg*, 15 (5): 785-789, 2009.

16. Durmuş AS: Effect of seprafilm on prevention of intraabdominal adhesions in rats. *Indian Vet J*, 87, 816-817, 2010.

17. Lalountas MA, Ballas KD, Skouras C, Asteriou C, Kontoulis T, Pissas D, Triantafyllou A, Sakantamis AK: Preventing intraperitoneal adhesions with atorvastatin and sodium hyaluronate/carboxymethylcellulose: A comparative study in rats. *Am J Surg*, 200 (1): 118-123, 2010.

18. Attar R, Yıldırım G, Kumbak B, Ficioğlu C, Demirbağ S, Yeşildağlar

N: Efficacy of melatonin and hyaluronate/carboxymethylcellulose membrane in preventing adhesion reformation following adhesiolysis in the rat uterine model. *J Obstet Gynaecol Res*, 37 (2): 125-131, 2011.

19. Şahin Y, Sağlam A: Synergistic effects of carboxymethylcellulose and low molecular weight heparin in reducing adhesion formation in the rat uterine horn model. *Acta Obstet Gyn Scan*, 73 (1): 70-73, 1994.

20. Başbuğ M, Aygen E, Tayyar M, Kaya E, Narin F, Öktem Ö: Hyaluronic acid plus heparin for improved efficacy in prevention of adhesion formation in rat uterine horn model. *Eur J Obstet Gyn R B*, 78, 109-112, 1998.

21. Ward BC, Panitch A: Abdominal adhesions: Current and novel therapies. *J Surg Res*, 165, 91-111, 2011.

22. Avşar FM, Şahin M, Aksoy F, Avşar AF, Aköz M, Hengirmen S, Bilici S: Effects of diphenhydramine HCl and methylprednisolone in the prevention of abdominal adhesions. *Am J Surg*, 181 (6): 512-515, 2001.

23. Altun Ö, Tayyar AT, Tayyar A, Özgün T, Öztürk F, Tayyar M: The effect of low molecular weight heparin and octreotide in an experimental uterine horn adhesion model. *Cumhuriyet Med J*, 34, 307-313, 2012.

24. Mazuji MK, Kalambaheti K, Pawar B: Prevention of adhesion formation with polyvinylpropiondone. Preliminary report. *Arch Surg*, 54, 1011-1015, 1964.

25. Hooker GD, Taylor BM, Driman DK: Prevention of adhesion formation with use of sodium hyaluronate-based bioresorbable membrane in a rat model of ventral hernia repair with polypropylene mesh. A randomized, controlled study. *Surgery*, 125, 211-216, 1999.

26. Tayyar M, Turan R, Ayata D: The use of amniotic membrane plus heparin to prevent postoperative adhesions in the rabbit. *Tokai J Exp Clin Med*, 18 (1-2): 57-60, 1993.

27. Kılıç K, Kılıç N, Kılıç E, Yayla S, Ermutlu CŞ, Yayla S, Özyayın İ, Dağ S: A comparison of the efficacy of dimethyl sulfoxide (DMSO) and synovial fluid in the prevention of peritoneal adhesions (experimental rabbit model). *Kafkas Üniv Vet Fak Derg*, 19 (Supp-A): 27-32, 2013.

28. Lim R, Morrill JM, Lynch RC, Reed KL, Gower AC, Leeman SE, Stucchi AF, Becker JM: Practical limitations of bioresorbable membranes in the prevention of intra-abdominal adhesions. *J Gastrointest Surg*, 13, 35-42, 2009.

29. Arıkan S, Adaş G, Barut G, Toklu AS, Kocakuşak A, Uzun H, Kemik Ö, Daduk Y, Aydın S, Purisa S: An evaluation of low molecular weight heparin and hyperbaric oxygen treatment in the prevention of intra-abdominal adhesions and wound healing. *Am J Surg*, 189 (2): 155-160, 2005.

30. Sağıl S, Özşener S, Dinçer O, Yılmaz H, Karadadaş N: The effect of medroxyprogesterone acetate and heparin in the prevention of postsurgical adhesion formation in the rat uterine model. *J Obstet Gynaecol Res*, 25 (4): 287-293, 1999.

31. Correles F, Corrales M, Sachirmer CC: Preventing intraperitoneal adhesions with vitamin E and sodium hyaluronate/carboxymethylcellulose. A comparative study in rats. *Acta Cir Bras*, 23 (1): 36-41, 2008.

32. Durmuş AS, Yıldız H, Yaman I, Şimşek H: Efficacy of vitamin E and selenium for the prevention of intra-abdominal adhesions in rats: uterine horn models. *Clinics*, 66 (7): 1247-1251, 2011.

Risk Factors Associated with Passive Immunity, Health, Birth Weight and Growth Performance in Lambs: II. Effects of Passive Immunity and Some Risk Factors on Growth Performance During the First 12 Weeks of Life ^[1]

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Summary

The primary objective of this study was to evaluate the relationship between passive immunity and growth performance in lambs in neonatal and post-neonatal periods. The study also evaluated the effect of gender, type of birth, birth weight and health of lambs, lambing season and dam's age on growth performance. This study included two crossbred Akkaraman flocks (n=301) in Kars, Turkey. The results disclosed that serum IgG concentration determined at 24th of birth (SlgG-24) was significantly associated with growth performance in both periods (P<0.001) but this relationship was poor (R²=0.148 to 0.198). Neonatal growth performance was influenced only by birth weight, health status, SlgG-24 and type of birth (R²=0.527 to 0.721 P<0.001) on multiple stepwise regression analyses. The study also disclosed that post-neonatal growth performance was affected by all variables (P<0.001 R²=0.471 to 0.472). This study suggested that passive transfer cause a significant amount of variation in growth performance for lambs from birth to 12 weeks of life, a conclusion that has not previously been identified. It also disclosed positive association among growth performance, birth weight, type of birth, gender, health status of lambs, dam's age and lambing season. The growth performance was rendered by dam's age (≤ 2 years old), twin lambs, female lambs, winter lambing, illness, and birth weight of ≤ 3 kg. These factors should be taken into consideration and dealt with in order to increase growth performance so that which farm productivity and profitability is maintained.

Keywords: Lamb, Passive immunity, Growth performance, Risk factors

Kuzularda Pasif İmmünite, Sağlık, Doğum Ağırlığı ve Büyüme Performansı İle İlişkili Risk Faktörleri: II- Pasif İmmünite ve Bazı Risk Faktörlerinin İlk 12 Haftalık Büyüme Performansı Üzerine Etkisi

Özet

Bu çalışmanın temel amacı kuzularda neonatal ve sonraki periyotta pasif immünite ve büyüme performansı arasındaki ilişkinin belirlenmesidir. Çalışmada ayrıca kuzunun cinsiyeti, doğum tipi, doğum ağırlığı ve sağlığı ile birlikte kuzulama sezonu ve anne yaşının büyüme performansı üzerine etkisi değerlendirildi. Çalışma Türkiye'de Kars'ta iki Akkaraman melezi sürüde (n=298) yürütüldü. Sonuçlar doğumdan sonra 24. saat serum IgG konsantrasyonu ve her iki dönem büyüme performansı arasında önemli (P<0.001) fakat zayıf (R²=0.148-0.198) bir ilişki olduğunu göstermiştir. Çoklu adımsal regresyon modeli ile neonatal büyüme performansının sadece doğum ağırlığı, sağlık durumu, SlgG-24 ve doğum tipinden etkilendiği (P<0.001 R²=0.527-0.721) belirlendi. Aynı metot ayrıca post-neonatal büyüme performansının tüm değişkenlerden etkilendiği göstermektedir (P<0.001 R²=0.471-0.472). Bu çalışma kuzularda pasif immunitenin ilk 12 haftalık dönem büyüme performansı ile önemli oranda varyasyon gösterdiğini işaret etmektedir. Bu sonuç daha önce değerlendirilmemiştir. Çalışmada ayrıca doğum ağırlığı, doğum tipi, cinsiyet, sağlık durumu, anne yaşı ve kuzulama sezonunun da büyüme performansı üzerine etkili olduğu belirlendi. Kuzularda anne yaşının ≤ 2 olması, ikiz, dişi ve kış sezonunda doğması, hastalığa maruz kalması ve doğum ağırlığının ≤ 3 kg olmasının büyüme performansını azalttığı saptandı. Bu faktörler büyüme performansını artırmak için dikkatle ele alınması gerekmektedir. Böylece çiftlik verimliliği ve karlılığı artırılabilir.

Anahtar sözcükler: Kuzu, Pasif immünite, Büyüme performansı, Risk faktörleri



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INTRODUCTION

Most of the outputs from livestock consist of food production in the form of meat, milk and other products. The profitability of sheep farm is significantly limited by considerable growth performance, low market weight and ill-health^[1-5]. Therefore, it is important for the sheep industry to be well informed about the external, controllable factors that affect live weight. For this reason, much work has been done on the effects of genetic and non-genetic factors on growth performance. The growth rate of lambs, particularly during the early stages, is influenced by several factors including breed, the ewe's diet, milk yield, parity or age of the ewe, type of birth, gender, birth weight and environmental factors such as the lambing season^[1,2,4,6]. Thus, there is a need for evaluating the effects of animal-related and environmental factors on growth traits if adequate level of growth performance is to be attained. Nonetheless, not enough research has been carried out in Turkey to determine the relationship between risk factors and growth performance in lambs^[7-9].

Recent studies indicate that passive immunity may be a significant factor that affects growth performance^[10]. Lambs are born agammaglobulinemic at birth due to syndesmochorial placentation. Therefore, immunity during the neonatal period depends on the passive transfer of maternal IgG through the colostrum. This process is referred as passive transfer of immunity (PTI), determined by measuring serum IgG concentration (SIgGC)^[10-13]. If neonatal lambs are unable to obtain and absorb a sufficient amount and quality of colostral IgG in the first 12 h of birth, secondary immunodeficiency disorder in other words Failure of Passive Transfer (FPT) develops. FPT results in hypogammaglobulinemia and predisposes neonates to diseases^[10-16]. There exists sufficient amount of knowledge on the association between PTI and the risk of illness and death in neonatal lambs^[10,12,14,15], but not enough has been reported about the potential long-term implications of passive transfer. PTI appears to help predict health and productivity in juvenile ruminants, both before and after the neonatal period^[10,11]. Studies to date have indicated that there appear a significant linear correlation between passive immunity and growth performance in different periods during the first 6 months of life in beef^[17], dairy heifer^[18] and buffalo^[19] calves, as well as in dairy lambs^[10,20] and dairy goat kids^[21,22]. Furthermore, an association between PTI and first lactation, milk yield and milk fat content in dairy heifer calves^[23] and pre- and post-weaning growth performance in crossbreed calves have been found^[24,25]. Nevertheless, very little information is available on how passive transfer affects growth performance in lambs, especially after the neonatal period.

To the authors' knowledge, no reports have been published on the predictive value of passive transfer for

the growth performance of lambs in long term period. Therefore, the primary objectives of this study was to determine the relationship between passive immunity and growth performance in different periods of the first 12 weeks of life and the secondary purpose of this study was to evaluate the effect of gender, type of birth, birth weight, health of lambs, lambing season and dam's age on growth performance in lambs of two crossbred Akkaraman flocks in Kars, North-Eastern Anatolia, Turkey.

MATERIAL and METHODS

Animals, Data Collection and Farm Management

This study was carried out on two sheep farms located in the central of Kars province in North-Eastern Anatolia, Turkey in 2009. All ewes and lambs were kept under identical feeding and management conditions. Management was typical of North-Eastern Anatolian flocks with lambs being born in winter (December to February) or spring (March to May), and being raised intensively. Other than deworming agents and routine vaccines against clostridial diseases, no drugs and other compounds administered during gestation or parturition. Only lambs resulting from observed parturitions were included in the study. Plastic ear tags were attached to both ears of study lambs soon after their birth and the following data were recorded for each lamb: an individual identification number, gender, date of birth, age of dam, ear tag number of dam and type of birth. The lambs were weighed at birth (before colostrum intake) using a scale [CASIA DB2-150 kg (± 30 g)]. After this procedure, lambs were allowed to naturally suckle their dams. The newborn lambs were kept with their dams during their first week of life. After this period, the lambs were transferred to a separate pen and allowed to suckle twice a day (in the morning and evening) for 3 months. Lambs had access to hay after the first week of neonatal life, and to straw and commercial growth feed (Bayramoglu AS, Turkey) from the third week of life. This feeding regime lasted for three months. Subsequently, lambs were turned on pasture and supplemented with hay and commercial feed when they were brought in for the night. No vaccines, drugs or other compounds were administered to lambs during the neonatal period. Deworming agents and routine vaccines against clostridial diseases were administered to the lambs after the neonatal period. The lambs were also weighed on day 28 and day 84 using the same scale.

Blood Sampling and IgG Measurement

Blood samples were collected from lambs by jugular venipuncture at 24 ± 1 h. Serum was harvested by centrifugation and stored at -20°C until analysis. SIgG-24 was measured using a commercial ELISA kit (Bio-X Competitive ELISA Kit For Ovine Blood Serum IgG Assay-BIO K 350, Bio-X Diagnostics, Belgium).

Clinical Examination

Clinical examination was performed as previously defined by the authors^[14]. The health status of the lambs was monitored by visiting farms daily during the neonatal period and every two days in the period covering 5 to 12 weeks of life referred as post-neonatal period in the text.

Statistical Analysis

The present study was conducted on 301 Akkaraman crossbreeds and 347 lambs born to these ewes. Those lambs whose birth weight, body weight on 28 and 84 days of life, gender, type of birth, health status, serum IgG concentration, birth date and dam's age were recorded and those survived neonatal or post-neonatal period were included in statistical analysis. Therefore, only 298 and 280 lambs were used to evaluate growth performance during neonatal and post-neonatal periods, respectively. Mean \pm SD (Range) values for SIgG-24, birth weight, neonatal (day 28) and post-neonatal (day 28 to day 84) weight, neonatal (birth to day 28) and post-neonatal (from 28 to 84 days) Average Daily Gain (ADG) were calculated. The lambs were categorized as healthy or ill based on their clinical examination results. Birth weights were categorized as low (≤ 3 kg), medium (>3 to ≤ 4 kg) or high (>4 kg). Dam's age was categorized as ≤ 2 , 3, 4 or ≥ 5 years. SIgG-24 was also categorized as <1000 mg/dL, 1001-2000 mg/dL or >2000 mg/dL. The Tukey HSD test was used to identify differences in growth performance in lambs grouped according to SIgG-24, birth weight and dam's age. Independent Samples T test was used to identify variations in growth performance according to gender, type of birth, birth date and health of the lambs. Multivariable stepwise linear regression analysis (MSRA) was used to evaluate the association between SIgG-24, birth weight, age of dams (considered as continuous independent variables), type of birth, gender and health status of lambs, lambing season (considered as categorical independent variables) and the growth performance outcomes, including ADG and body weight. The linear regression model with the all potential independent variables considered in the study defined as follows: $Y = \alpha + \beta_1 X_1 + \beta_2 X_2 + \beta_3 X_3 + \beta_4 X_4 + \beta_5 X_5 + \beta_6 X_6 + \beta_7 X_7 + \epsilon$. Where Y denotes the weight or ADG, X_1 is the birth weight, X_2 is the type of birth (twin=1 and single=0), X_3 is the health of the lambs (healthy=0 and ill=1), X_4 is the passive immune status (SIgG-24), X_5 is gender (male=0 and female=1), X_6 is age of the dam and X_7 is the lambing season [Spring (March, April, May)=0 and Winter (December, January, February)=1], α is the y-intercept, and $\beta_1, \beta_2, \beta_3, \beta_4, \beta_5, \beta_6, \beta_7$ are the regression coefficients and ϵ , indicates the random part of the theoretical model. To identify the best models in the stepwise technique, the coefficients of determination (R^2) were used. The coefficient of determination was multiplied by 100 and expressed as a percentage to indicate the total variation in Y explained by the selected independent variables. The relationship between SIgG-24 and growth performance (weight or ADG) was explored by simple regression analysis. The methods of multivariable

and simple regression have been described previously in detail^[10,11]. The computer program SPSS was used for all analyses and values of $P < 0.05$ were considered to be significant. The program Origin 6 was used to obtain scatter diagram illustrations (Origin 6 Copyright® 1991-1999 Microcol TM, Software, Inc) in regression analysis.

RESULTS

The study evaluated 298 lambs and mean SIgG-24 was 22.1 ± 10.74 (2.71-53.02) mg/mL. Birth weight ranged from 2.260 to 5.900 g (mean \pm SD, 4.075 ± 631 g). Changes in growth performance based on birth weight, gender, type of birth, SIgG-24 and health status in lambs, age of dam and lambing season have been given in Table 1. In addition, the effects of these factors on neonatal and post-neonatal growth performance (ADG or body weight) have been presented in Table 2 and Table 3, respectively.

Effect of Passive Immunity and Other Some Variables on Neonatal Growth Performance

Body weight at 28 days of life ranged from 4.364 to 14.015 g (8.852 ± 1.930 g). Average daily gain from birth to day 28 ranged from 16 to 339 g/d (mean \pm SD, 170 ± 53 g/day). Lambs with SIgG-24 levels below 1000 mg/dL had significantly lower neonatal growth performance (ADG or body weight) than lambs with SIgG-24 in the range of 1001 to 2000 ($P < 0.001$) and above 2000 mg/dL ($P < 0.001$). Lambs having SIgG-24 in the range of 1001 to 2000 mg/dL had significantly lower growth performance when compared to lambs with SIgG-24 above 2000 mg/dL. With regard to birth weight, the neonatal growth performance in the low weight category was significantly lower than those of medium ($P < 0.01$) and high birth weight category ($P < 0.001$). Similarly, lambs with medium birth weight had significantly ($P < 0.001$) lower weight or ADG than those with high birth weight. Neonatal growth performance was significantly lower in twins ($P < 0.001$) and diseased ($P < 0.001$) lambs. Body weight of day 28 was significantly ($P < 0.05$) higher in male lambs when compare to female lambs. However, ADG from birth to 28 day was not significantly different between male and female lambs ($P > 0.05$). There was no significant difference in growth performance between lambs born in spring and winter seasons ($P > 0.05$). Dam's age did not significantly effect on neonatal growth performance.

The simple regression models disclosed that SIgG-24 was significantly ($P < 0.001$) associated with both weight determined on day 28 ($R^2 = 0.148$; Fig. 1A) and ADG determined for the period of birth to 28 days ($R^2 = 0.151$; Fig. 1B). Each 1 mg/mL increase in SIgG-24 was associated with a 1.92 g/d increase in ADG and a 69.2-g increase in body weight of day 28.

On multiple regressions analysis, type of birth, SIgG-

Table 1. Changes in growth performance of lambs according to various factors**Tablo 1.** Kuzularda bazı faktörlere bağlı büyüme performansındaki değişiklikler

Factor	Group	Period					
		Neonatal Growth Performance (g) (up to 4 Weeks)			Post-Neonatal Growth Performance (g) (5 to 12 Weeks)		
		n	WG1	ADG1	n	WG2	ADG2
SlgG-24 (mg/dL)	≤1000	25	7029±19513 ^a	120±54 ^a	22	8793±2066 ^a	156±37 ^a
	1001-2000	142	8631±2026 ^b	165±55 ^b	135	11077±2844 ^b	197±51 ^b
	>2000	131	9439±1530 ^c	186±43 ^c	123	12700±2313 ^c	226±41 ^c
Type of Birth	Twin	80	6794±1178 ^{***}	118±34 ^{***}	74	9639±2486 ^{***}	172±44 ^{***}
	Single	218	9606±1569	190±45	206	12320±2561	220±46
Gender	Male	160	9109±2011 [*]	175±57	150	12069±2849 ^{**}	215±51 ^{**}
	Female	138	8554±1794	165±48	130	11084±2656	197±47
Dam Age (Years)	≤2	45	8545±1923 ^a	168±58 ^a	41	10696±2958 ^a	190±53 ^a
	3	134	8766±1880 ^a	169±49 ^a	126	11232±2787 ^{ac}	200±50 ^{ac}
	4	82	9087±1795 ^a	176±51 ^a	78	12109±2588 ^{bc}	216±46 ^{bc}
	≥5	37	9017±2370 ^a	168±65 ^a	35	12946±2520 ^b	231±45 ^b
Health Status	Ill	40	7411±2125 ^{***}	129±60 ^{***}	81	10313±2426 ^{***}	184±43 ^{***}
	Healthy	258	9075±1802	177±49	199	12141±2774	216±50
Lambing Season	Winter	91	8980±1835	176±52	87	10671±2615 ^{***}	190±47 ^{***}
	Spring	207	8796±1792	168±54	193	12036±2783	214±50
Birth Weight (g)	Low	17	6266±1175 ^a	124±40 ^a	16	8723±3074 ^a	155±55 ^a
	Medium	117	7622±1606 ^b	143±51 ^b	109	10512±2595 ^b	187±44 ^b
	High	164	9997±1308 ^c	195±41 ^b	155	12684±2799 ^c	226±43 ^c

*** $P < 0.001$, ** $P < 0.01$, * $P < 0.05$, **WG1** = weight on day 28, **ADG1** = [(day 28 weight-birth weight)/28x1000], **WG2** = (weight on day 84-weight at day 28), **ADG2** = [(WG2/56)x1000]

Table 2. Regression models between independent variables and neonatal growth performance in lambs**Tablo 2.** Kuzularda neonatal büyüme performansı ve bağımsız değişkenler arasındaki regresyon modelleri

	Healthy Status	Formulas	R ²	P
	General (n=298)	W=7328.9 + 69.2 x IgG ADG=128.2 + 1.92 x IgG	0.148 0.151	<0.001 <0.001
Simple Regression Analysis *	Healthy (n=258)	W=7805.8 + 55.3 x IgG ADG =141.8 + 1.53 x IgG	0.105 0.109	<0.001 <0.001
	Ill (n=40)	W=5837.9 + 98.2 x IgG ADG=86.27 + 2.67 x IgG	0.215 0.197	<0.01 <0.01
Multiple Regression Analysis **	Model	Formulas	R ²	P
	Onset	W=2.4 x BW - 949.7	0.622	<0.001
	Final	W=1602 + 1.7 x BW - 1186 x TB - 796 x HS + IgG x 21.1	0.721	<0.001
	Onset	ADG=190.3-72*TB	0.364	<0.001
	Final	ADG=57.4 - 42 x TB + 0.28 x BW - 28.48 x HS + 0.75 x IgG	0.527	<0.001

W = Weight (g), ADG = Average daily gain (g), BW = Birth weight (g), TB = Type of birth, HS = Health status of lambs, IgG = Serum IgG concentration at 24 h after birth, AD = Age of dam, LS = Lambing season, * Independent variable: IgG and dependent variable: (W or ADG), ** Independent variables: BW, TB, HS, IgG, Gender, AD and dependent variable: (W or ADG)

24, birth weight and health of lambs were significantly associated with body weight at day 28 and ADG from birth to 28 days. The variation in weight at day 28 and ADG from birth to day 28 was 72% ($R^2=0.721$) and 53% ($R^2=0.527$) respectively (Table 2).

Effect of Passive Immunity and Other Some Variables on Post-neonatal Growth Performance

Total body weight from day 28 to day 84 (post neonatal period weight) ranged from 3.400 to 18.650 g (mean ± SD:

Table 3. Regression models between independent variables and post-neonatal growth performance in lambs**Tablo 3.** Kuzularda post-neonatal büyüme performansı ve bağımsız değişkenler arasındaki regresyon modelleri

	Healthy Status	Formulas	R ²	P
Simple Regression Analysis*	General (n=280)	$W = 9010.9 + 118.5 \times \lg G$	0.198	<0.001
		$ADG = 160.4 + 2.1 \times \lg G$	0.198	<0.001
	Healthy (n=199)	$W = 9642.2 + 105.9 \times \lg G$	0.167	<0.001
		$ADG = 171.7 + 1.89 \times \lg G$	0.168	<0.001
	Ill (n=81)	$W = 8452.2 + 103.9 \times \lg G$	0.145	<0.001
		$ADG = 150.3 + 1.86 \times \lg G$	0.146	<0.001
	Model	Formulas	R ²	P
Multiple Regression Analysis**	Onset	$W = 2165.7 + 2.32 \times BW$	0.273	<0.001
	Final	$W = 4918.5 + 1.2 \times BW + 68.3 \times \lg G - 956.9 \times LS - 1097.2 \times HS - 1088.4 \times TB + 440.2 \times DA - 639.9 \times \text{Gender}$	0.471	<0.001
	Onset	$ADG = 38.2 + 0.041 \times BW$	0.273	<0.001
	Final	$ADG = 87.29 + 0.021 \times BW + 1.2 \times \lg G - 17.1 \times LS - 19.5 \times HS - 19.4 \times TB + 7.89 \times AD - 11.5 \times \text{Gender}$	0.472	<0.001

W = Weight (g), *ADG* = Average daily gain (g), *BW* = Birth weight (g), *TB* = Type of birth, *HS* = Health status of lambs, *lgG* = Serum IgG concentration at 24 h after birth, *AD* = Age of dam, *LS* = Lambing season, * Independent variable: *lgG* and dependent variable: (*W* or *ADG*), ** Independent variables: *BW*, *TB*, *HS*, *lgG*, *Gender*, *AD* and dependent variable: (*W* or *ADG*)

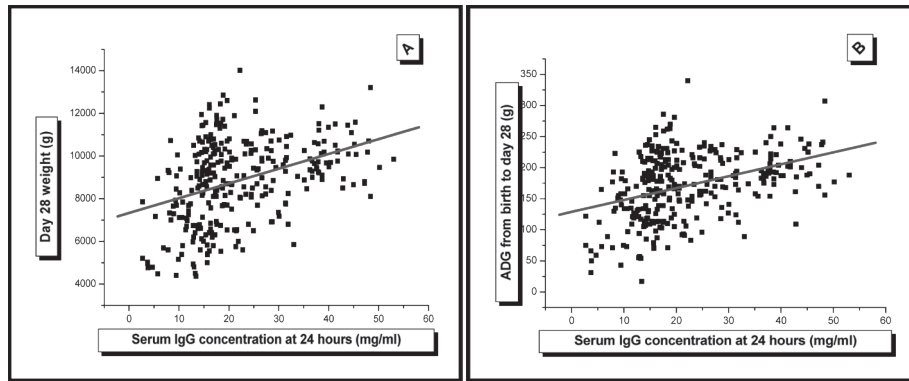


Fig 1. Scatterplots of serum IgG concentration (mg/mL) 24 h after parturition in 298 lambs versus neonatal growth performance [body weight (g) at day 28 (A), ADG (g/d) from birth to day 28 (B)]. In each graph, the solid line represents the best fit for the data, as determined by means of simple linear regression

Şekil 1. 298 kuzuda doğumdan sonra 24. saat serum IgG konsantrasyonu ve neonatal büyüme performansı [28. gün vücut ağırlığı (g) (A), doğumdan 28. güne kadar olan ADG (g/d) (B)] arasındaki ilişkiyi gösteren dağılım grafiğinin görünümü. Basit doğrusal regresyon ile belirlenen grafikte, koyu çizgi, iki değişken arasındaki en iyi uyum noktasını göstermektedir

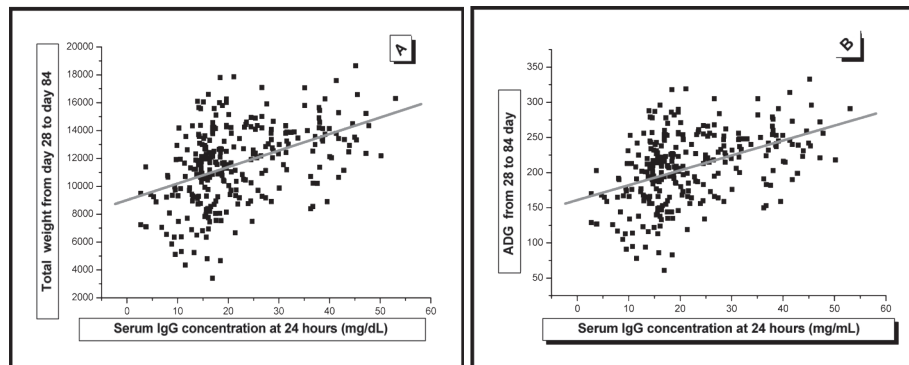


Fig 2. Scatterplots of serum IgG concentration 24 h after parturition in 298 lambs versus post-neonatal growth performance [total gain (g) (C) and ADG (g/d) (D) between day 28 and day 84]. In each graph, the solid line represents the best fit for the data, as determined by means of simple linear regression

Şekil 2. 280 kuzuda doğumdan sonra 24. saat serum IgG konsantrasyonu ve post-neonatal büyüme performansı [28 ve 84. günler arasındaki gün vücut ağırlığı (g) (A), doğumdan 28. güne kadar olan ADG (g/d) (B)] arasındaki ilişkiyi gösteren dağılım grafiğinin görünümü. Basit doğrusal regresyon ile belirlenen grafikte, koyu çizgi, iki değişken arasındaki en iyi uyum noktasını göstermektedir

11.612±2.799 g). ADG during this period ranged from 60 to 333 g/d (mean ± SD: 207±50 g/d). Lambs with SlgG-24 levels below 1000 mg/dL had significantly lower post-neonatal growth performance than lambs with SlgG-24 in the range of 1001 to 2000 ($P<0.001$) and SlgG-24 above 2000 mg/dL ($P<0.001$). As the SlgG-24 increased the growth performance was enhanced (Table 2). Lambs in low birth weight had significantly lower growth performance when compare to those born with medium ($P<0.001$) and high birth weight ($P<0.001$) in post-neonatal period. Lambs born from dams aged 4 ($P<0.05$) and ≥5 years old ($P<0.01$) had significantly better growth performance during the post-neonatal period when compare to other categories. Lambs born in the winter season, twin and female lambs and those with exposure to any disease had significantly lower growth performance from day 28 to day 84 than their contemporaries born in the spring season ($P<0.001$), singleton ($P<0.01$), male lambs ($P<0.001$) and those that remained healthy ($P<0.001$).

The simple regression models showed that SlgG-24 was significantly ($P<0.001$) associated with body weight gain ($R^2 = 0.198$; Fig. 1C) and ADG from day 28 to day 84 ($R^2=0.198$ Fig. 1D). Each 1 mg/mL increase in SlgG-24 was associated with a 2.1 g/d increase in ADG and a 118.5 g increase in weight during the post neonatal period.

Birth weight, gender, type of birth, SlgG-24 and health of lambs, age of dam and lambing season had significant relationship with body weight and ADG from day 28 to day 84 on multiple stepwise regression analysis. The final model showed that 47% ($R^2=0.471$) of the variance in total weight gain and 47% ($R^2=0.472$) in ADG from day 28 to day 84 was explained by the independent variables.

DISCUSSION

Recent studies have shown that passive immunity in newborn ruminants [17-22] not only helps to prevent disease but also enhances growth performance. However, the relationship between growth performance and passive immunity in lambs was studied for only healthy and small number of lambs (generally <40) and the period generally restricted to neonatal period (first 28 days of life) [10,15,20]. Therefore this study explained the relationship between passive immunity and other parameters and growth performance beyond the neonatal period for the first time.

Passive Immunity and Growth Performance

The results of our study indicated that passive transfer, was a significant source of variation in both neonatal and post-neonatal periods with regard to growth performance in lambs that were allowed to suckle naturally. Similar results were reported for dairy heifer calves [18,23,25], dairy healthy goats [21,22], beef calves [17,24] and healthy buffalo calves [19]. Our study determined a significant positive

linear correlation between SlgG-24 and body weight at day 28 and ADG from birth to day 28 as reported by a single previous study [10] but this study was conducted only 20 healthy lambs. However, no study has yet investigated the relationship between growth performance and SlgG-24 in lambs during the post-neonatal period. The present study also revealed a significant linear relationship between SlgG-24 and post-neonatal growth performance as SlgG-24 increased, the growth performance also improved significantly in both periods of life.

The present study did not explicitly disclose the biological basis for relationship between SlgG-24 and growth performance. However, this relationship can be explained by a number of theories in the light of earlier studies [10,19,21]. First of all, there may be a direct cause-effect relationship between SlgG-24 and growth performance as passive transfer of immunity might have enhanced efficient metabolic systems and rendered disease occurrence which in turn contributes to better growth performance [10,12,21,26,27]. The positive linear relationship among SlgG-24, health status and growth performance in this study added credence to above assumptions as low serum IgG concentrations at 24 to 72 h associated with reduced performance of surviving lambs through increased susceptibility to diseases [10,12,21,27]. Secondly, Massimini et al. [10], suggested that lambs receives and absorbs not only IgG but also nonimmunoglobulin factors present in colostrums which may contribute to growth performance as it is well known that colostrum also contains nutrients, minerals, trace elements, vitamins, enzymes, hormones, growth factors, antibodies and other immunologically active component such as cells, cytokines and acute-phase proteins which are necessary for normal development [10-13,28-31]. Neonatal ruminants are pseudomonogastrics and maturation of their gastrointestinal tract is modulated by growth factors through absorption of colostrums [30,31]. In recent years, considerable number of researchers examined the relationships between the development of neonatal calves and various growth factors, such as insulin-like growth factor-I and the growth hormone [30,31] and yet this aspects remains to be explored in lambs. Studies also revealed the effect of non-immunoglobulin components such as neutrophils, serum opsonic activity, colostrum cytokines, complement proteins and lactoferrin on improvement of weight gain by protecting against diseases [10,21,28-32] through boosting the immune and metabolic system. However, much is required to be performed with regard to the effect of non-immunoglobulin colostrum content on neonatal and post-neonatal health and growth [10].

In contrast, many researchers also reported no correlation between passive immunity and growth performance in ruminants [15,20,26,33,34]. However, these studies had many different aspects to the present study such as small number of subjects ($n<40$) [15,20] and species of ruminant (goat, calves) studied [21,26,33,34], method of colostrum feeding (heat

treated colostrums, bottle fed)^[26,34], and measurement of Ig concentration (zinc sulfate turbidity test)^[33,34], and length of study^[20,21].

Other Factors that Affect Growth Performance

The present study revealed that the variation attributable to SlgG-24 accounted on average for 15% of the total variation in ADG and weight from birth to day 28 and 20% of the total variation in weight and ADG from day 28 to day 84. Thus, a large proportion of variation in growth performance was not explained by SlgG-24. Multi-variable stepwise linear regression analysis determined a considerable variation in growth performance by health status, gender, type of birth, birth weight, dam's age and lambing season. Of these factors, birth weight, type of birth and health status were considerable source of variation for growth performance in neonatal period, while post-neonatal growth performance was influenced by all variables.

Health Status

Health of the lambs was positively associated with growth performance and in both periods as healthy lambs had a better growth rate than lambs exposed to disease and disease caused 796 g and 1097 g decrease in body weight for each lamb that became ill in neonatal and post-neonatal period, respectively. It is well known that the severity and duration of the disease results in decreased growth performance^[26,27]. This is attributed to the fact that diseased animal cannot fully take advantage of the nutrients for growth and development, since a portion of the ingested nutrients must be committed to attacking and eliminating pathogens as well as repairing any damage caused^[18,35].

Birth Weight

A positive and strong linear relationship between birth weight and growth performance in both periods was evident in our study. As birth weight increased, growth performance also increased considerably. Similar results were reported by previous studies^[2,4,36,37]. Several explanations have been put forward. Of these, lambs born with acceptable birth weight (generally >3 kg) are able to ingest sufficient amounts of colostrum earlier because they are physically strong and viable. This in turn not only increases their resistance to diseases, it also ensures that they ingest many components of the colostrum that affect growth performance^[10,38]. On the other hand, lambs with lower birth weight are physically weak, to stand at birth and to suckle sufficient amount of colostrum and this consequently results frequent exposure to disease as was the case in our study where lambs with low birth weight were more prone to disease development and poor growth performance^[35,38-40]. It is already known that lambs with a birth weight of ≤ 3 kg usually suffer failure of passive transfer and that most of them die during the

neonatal period, or even if they survive, they do not exhibit good growth performance^[5,12,35,39-41]. Other explanation may be that lambs with low birth weights are less mature than those with high birth weights with regard to the development of their metabolic and endocrine systems that improve lambs ability to generate energy from amino acids and sustain gluconeogenesis immediately after parturition^[42,43].

Type of Birth

Type of birth had a significant effect on the weight of the lamb as single-born lambs were heavier and had higher ADG in the neonatal and post-neonatal periods as compared to twins in this study. Twin lambs were approximately 1088 g lighter than singleton lambs in the neonatal period and 1186 g in the post-neonatal period and ADG of twin lambs was 42 g lower than single lambs during the neonatal period and 19 g lower during the post-neonatal period. Single lambs have already been reported to attain better growth performance in comparison to twin lambs^[2,4,7,44-48]. This has been attributed to competition between the siblings for dam's milk supply, inadequate milk to rear two lambs, lower birth weight of twins, mismothering, hypothermia, or the inability to obtain sufficient milk or colostrum due to physical weakness^[1,6,8,12,35,41].

Gender

This study revealed no significant association between gender and growth performance in the first month of life on multiple regression analysis as reported by others^[2,4,47]. On the contrary, studies reported that gender had a significant effect on ADG and weight in this period^[36,45,48]. On the other hand, male lambs recorded a higher ADG and were heavier than female lambs from day 28 to day 84, and gender was a significant source of variation in post-neonatal growth performance in this study. Similar results have previously been reported^[4,6,7,8,45]. The faster growth of male lambs could be due to hormonal differences in their endocrinological and physiological functions which play role in enhancement of growth and their superior weight at birth compared to females^[4,6,44]. On the contrary studies also reported no significant effect of gender on ADG^[1,2,36,44,47] and body weight^[1,2,48] from birth to three months of life.

Age of Dam

Dam's age had no significant effect on growth performance in the neonatal period as earlier studies reported^[2,4,44,45] but some studies found the effect of parity or dam's age on weight at day 28 or on ADG from birth to day 28^[36,47]. On the other hand multiple regression analysis revealed a significant positive effect of dam's age on growth performance in post-neonatal period. This effect might be explained by the difference in milk production and maternal care since maiden or first parity ewes are less

mature physiologically and consequently produce less milk, and are less experienced in taking care of their lamb ^[2,4,6,9,44,47]. Additionally, younger ewes produce lighter lambs due to competition between fetus growth and maternal growth, which has a negative effect on birth weight and subsequent growth performance ^[1,38,41,49-51]. However, contradicting results suggesting no effect of parity on growth performance from birth to three months of life ^[1,2,4,44,45].

Lambing Season

Lambing season had no effect on growth performance in the neonatal period in this study as reported by Taye et al.^[4]. In contrast, earlier studies reported that lambing season had a significant effect on neonatal growth performance ^[2,36,47]. However, a significant relationship between lambing season and growth performance was determined in the post-neonatal period on multiple stepwise regression analysis. Other studies have also reported positive association between lambing season and growth performance during the first three months of its life ^[6,9,36,47,51]. Studies suggested that lambing season could cause changes in growth performance due to temperature during the birth season, morbidity rates, birth weight due to the dam's nutrition in late pregnancy and milk yield due to feeding regime of dams in post-parturient period ^[6,8,35,44,46,51].

This study revealed association between growth performance and birth weight, type of birth, health status and SlgG-24 in neonatal period and additionally lambing season, dam's age and gender in post-neonatal period. Statistical models used in this study also suggested that variation in growth performance remained to be effected by some other factors noted previously ^[5,10,38,43,50]. These factors include, managemental interventions such as feeding regime and other animal-related factors such as body condition score, breed, placental development and function, dam and lamb behavior at birth, maternal ability, gastrointestinal absorptive ability of the lambs ^[2,6,7,38,43,50]. The amount of variation attributable to farm management procedures, the production system and the ewe's prenatal nutrition and breed was minimized in this study as all lambs were taken from two farms that had similar management practices and reared Akkaraman crossbreeds.

In conclusion, this study indicated that passive transfer causes a significant amount of variation in growth performance for lambs from birth to 12 weeks of life, a conclusion that was noted for the first time. It is possible for passive transfer monitoring programs to identify lambs suffering from FPT, but there are many factors that cause FPT ^[10,11,21,41]. That is to say that programs designed only to identify affected lambs will not be as effective as comprehensive flock management programs as our study has found that aside from passive immunity, growth performance is affected by birth weight, type of birth, health status of lambs, dam's age and lambing season. These factors may be improved by managemental inter-

vention in order to increase growth performance and thus farm income and welfare.

REFERENCES

1. **Yiheiyis A, Tegegn F, H/Meleket M, Taye M:** Pre-weaning growth performance of Sekota sheep breed in Waghimra Zone, Ethiopia. *Online J Anim Feed Res*, 2, 340-343, 2012.
2. **Talore DG:** On-farm performance evaluation of indigenous sheep and goats in Alaba, Southern Ethiopia. A thesis, Department of Animal and Range Sciences, College of Agriculture, School of Graduate studies, Hawassa University, Awassa, Ethiopia. 2009.
3. **Gökçe E, Erdoğan HM:** Pneumonia in neonatal lambs: Frequency and some associated risk factors. *Kafkas Univ Vet Fak Derg*, 14 (2): 223-228, 2008.
4. **Taye M, Abebe G, Gizaw S, Lemma S, Mekoya A, Tibbo M:** Growth performances of Washera sheep under smallholder management systems in Yilmanadensa and Quarit districts, Ethiopia. *Trop Anim Health Prod*, 42, 659-667, 2010.
5. **Mukasa-Mugerwa E, Lahlou-Kassi A, Anindo D, Rege JEO, Tembely S, Tibbo M, Baker RL:** Between and within breed variation in lamb survival and the risk factors associated with major causes of mortality in indigenous Horro and Menz sheep in Ethiopia. *Small Rumin Res*, 37, 1-12, 2000.
6. **Tibbo M:** Productivity and health of indigenous sheep breeds and crossbreds in the central Ethiopian highlands. *PhD Dissertation*, Department of Animal Breeding and Genetics, Faculty for Veterinary Medicine and Animal Sciences, Swedish University of Agricultural Sciences (SLU), Uppsala, Sweden. 2006.
7. **Sezgin E, Kopuzlu S, Yüksel S, Esenbuğa N, Bilgin ÖC:** Determination of growth traits and heritabilities of growth characteristics of Hemşin Sheep reared in Artvin. *Kafkas Univ Vet Fak Derg*, 18 (6): 899-905, 2012.
8. **Yilmaz O, Denk H, Bayram D:** Effects of lambing season, sex and birth type on growth performance in Norduz lambs. *Small Rum Res*, 68, 336-339, 2007.
9. **Thieme O, Karazeybek M, Özbayat Hİ, Sözmen R:** Performance of village sheep flocks in Central Anatolia I. Growth of lambs. *Turk J Vet Anim Sci*, 23, 467-474, 1999.
10. **Massimini G, Britti D, Peli A, Cinotti S:** Effect of passive transfer status on preweaning growth performance in dairy lambs. *J Am Vet Med Assoc*, 229, 111-115, 2006.
11. **Britti D, Massimini G, Peli A, Luciani A, Boari A:** Evaluation of serum enzyme activities as predictors of passive transfer status in lambs. *J Am Vet Med Assoc*, 226, 951-955, 2005.
12. **Christley RM, Morgan KL, Parkin TD, French NP:** Factors related to the risk of neonatal mortality, birth-weight and serum immunoglobulin concentration in lambs in the UK. *Prev Vet Med*, 57, 209-226, 2003.
13. **Maden M, Altunok V, Birdane V, Nizamlioğlu M:** Blood and colostrum/milk serum γ -glutamyltransferase activity as a predictor of passive transfer status in lambs. *J Vet Med B Infect Dis Vet Public Health*, 50, 128-131, 2003.
14. **Gökçe E, Erdoğan HM:** An epidemiological study on neonatal lamb health. *Kafkas Univ Vet Fak Derg*, 15 (2): 225-236, 2009.
15. **Gilbert RP, Gaskins CT, Hillers JK, Parker CF, McGuire TC:** Genetic and environmental factors affecting immunoglobulin G₁ concentrations in ewe colostrum and lamb serum. *J Anim Sci*, 66, 855-863, 1988.
16. **Gökçe E, Ünver A, Erdoğan HM:** İshalli neonatal kuzularda enterik patojenlerin belirlenmesi. *Kafkas Univ Vet Fak Derg*, 16 (5): 717-722, 2010.
17. **Dewell RD, Hungerford LL, Keen JE, Laegreid WW, Griffin DD, Rupp GP, Grotelueschen DM:** Association of neonatal serum immunoglobulin G1 concentration with health and performance in beef calves. *J Am Vet Med Assoc*, 228, 914-921, 2006.
18. **Robison JD, Stott GH, DeNise SK:** Effects of passive immunity on growth and survival in the dairy heifer. *J Dairy Sci*, 71, 1283-1287, 1988.

19. **Mastellone V, Massimini G, Pero ME, Cortese L, Piantedosi D, Lombardi P, Britti D, Avallone L:** Effects of passive transfer status on growth performance in buffalo calves. *Asian-Aust J Anim Sci*, 24, 952-956, 2011.
20. **Kaymaz AA, Bakirel U, Cağtay P, Tan H:** The effects of serum IgG and trace elements-copper and zinc on the development of kıvrık lambs following colostrum intake. *Istanbul Univ Vet Fak Derg*, 26, 475-481, 2000.
21. **Massimini G, Mastellone V, Britti D, Lombardi P, Avallone L:** Effect of passive transfer status on preweaning growth performance in dairy goat kids. *J Am Vet Med Assoc*, 231, 1873-1877, 2007.
22. **Yalçın E, Temizel EM:** Sütten kesme öncesi dönemde oğlakların büyüme performansına pasif transfer durumunun etkisi. *Uludağ Univ J Fac Vet Med*, 29 (1): 23-26, 2010.
23. **DeNise SK, Robison JD, Stott GH, Armstrong DV:** Effects of passive immunity on subsequent production in dairy heifers. *J Dairy Sci*, 72, 552-554, 1989.
24. **Wittum TE, Perino LJ:** Passive immune status at postpartum hour 24 and long-term health and performance of calves. *Am J Vet Res*, 56, 1149-1154, 1995.
25. **Virtala AMK, Mechor GD, Grohn YT, Erb HN:** The effect of calving diseases on growth of female dairy calves during the first 3 months of life in New York State. *J Dairy Sci*, 6, 1040-1049, 1996.
26. **Quigley JD, Martin KR, Bemis DA: Potgieter LND, Reinemeyer CR, Rohrbach BW, Dowlen HH, Lamar KC:** Effects of housing and colostrum feeding on serum immunoglobulins, growth, and fecal scores of Jersey calves. *J Dairy Sci*, 78, 893-901, 1995.
27. **Pare J, Thurmond MC, Gardner IA, Picanso JP:** Effect of birth weight, total protein, serum IgG and packed cell volume on risk of neonatal diarrhea in calves on two California dairies. *Can J Vet Res*, 57, 241-246, 1993.
28. **Dawes ME, Tyler JW, Marsh AE, Larson RL, Steevens BJ, Lakritz JD:** In vitro effects of lactoferrin on lipopolysaccharide-induced proliferation, gene expression, and prostanoid production by bovine peripheral blood mononuclear cells. *Am J Vet Res*, 69, 1164-1170, 2008.
29. **Loste A, Ramos JJ, Fernández A, Ferrer LM, Lacasta D Verde MT, Marca MC, Ortín A:** Effect of colostrum treated by heat on immunological parameters in newborn lambs. *Livest Sci*, 117, 176-183, 2008.
30. **Blum JW:** Nutritional physiology of neonatal calves. *J Anim Physiol Anim Nutr*, 90:1-11, 2006.
31. **Pakkanen R, Aalto J:** Growth factors and antimicrobial factors of bovine colostrum. *Int Dairy J*, 7, 285-297, 1997.
32. **Prgomet C, Prenner ML, Schwarz FJ, Pfaffl MW:** Effect of lactoferrin on selected immune system parameters and the gastrointestinal morphology in growing calves. *J Anim Physiol Anim Nutr (Berl)*, 91, 109-119, 2007.
33. **Moraes MP, Weiblen R, Rebelatto MC, Moraes da Silva A:** Relationship between passive immunity and morbidity and weight gain in dairy cattle. *Ciência Rural*, 30, 299-304, 2000.
34. **O'Brien JP, Sherman DM:** Field methods for estimating serum immunoglobulin concentrations in newborn kids. *Small Rumin Res*, 11, 79-84, 1993.
35. **Gökçe E, Kırmızıgül AH, Çitil M, Erdoğan HM:** Risk factors associated with passive immunity, health, birth weight and growth performance in lambs: I. Effect of parity, dam's health, birth weight, gender, type of birth and lambing season on morbidity and mortality. *Kafkas Univ Vet Fak Derg*, 19 (Suppl-A): A153-A160, 2013, DOI: 10.9775/kvfd.2012.8440.
36. **Awgichew K:** Comparative performance evaluation of Horro and Menz sheep of Ethiopia under grazing and intensive feeding conditions. *PhD Dissertation*, Humboldt University, Berlin, Germany, 2000.
37. **Sušić V, Pavić V, Mioč B, Štoković I, Kabalin AE:** Seasonal variations in lamb birth weight and mortality. *Vet Arhiv*, 75, 375-381, 2005.
38. **Dwyer CM:** The welfare of the neonatal lamb. *Small Rumin Res*, 76, 31-41, 2008.
39. **Ahmad RA, Khan MT, Javed I, Hussein M:** Immunoglobulins and neonatal lambs mortality in Pak-Karakul sheep. *Vet Arhiv*, 70, 129-139, 2000.
40. **Khan A, Sultana MA, Jalvib MA, Hussain I:** Risk factors of lamb mortality in Pakistan. *Anim Res*, 55, 301-311, 2006.
41. **Gökçe E, Atakişi O, Kırmızıgül AH, Erdoğan HM:** Risk factors associated with passive immunity, health, birth weight and growth performance in lambs: III. The relationship among passive immunity, birth weight gender, birth type, parity, dam's health and lambing season. *Kafkas Univ Vet Fak Derg*, 2013 (In Press).
42. **Greenwool PL, Hunt AS, Slepatis RM, Finnerty KD, Alston C, Beermann DH, Bell AW:** Effects of birth weight and postnatal nutrition on neonatal sheep: III. Regulation of energy metabolism. *J Anim Sci*, 80, 2850-2861, 2002.
43. **Dwyer CM, Calvert SK, Farish M, Donbavand J, Pickup HE:** Breed, litter and parity differences in the morphology of the ovine placenta and developmental consequences for the lamb. *Theriogenology*, 63, 1092-1110, 2005.
44. **Abbas SF, Abd Allah M, Allam FM, Aboul-Ella AA:** Growth performance of Rahmani and Chios Lambs weaned at different ages. *Aust J Basic Appl Sci*, 4, 1583-1589, 2010.
45. **Abegaz S, Hegde BP, and Taye M:** Growth and physical body characteristics of gumuz sheep under traditional management systems in Amhara Regional State, Ethiopia. *Livestock Research for Rural Development*, 23 (5): 2011.
46. **Teleb DF, Saifelnasr EOH, El-Sayed EH:** Factors affecting performance and survivability of Saidi lambs from lambing to weaning. *Egypt J Sheep Goat Sci*, 4, 55-74, 2009.
47. **Gbangboche AB, Youssao AKI, Senou M, Adamou-Ndiaye M, Ahissou A, Farnir F, Michaux C, Abiola FA, Leroy PL:** Examination of non-genetic factors affecting the growth performance of djallonke sheep in soudanian zone at the Okpara breeding farm of Benin. *Trop Anim Health Prod*, 38, 55-64, 2006.
48. **Özcan M, Altinel A, Yılmaz AG, Yğüneş H:** Studies on the possibility of improving lamb production by two-way and three way crossbreeding with German Black Headed Mutton Kıvrık and Chios sheep breeds 1. Fertility lamb survival and growth of lambs. *Türk J Vet Anim Sci*, 25, 687-694, 2001.
49. **Yıldız N, H Denk:** Van Bölgesinde halk elinde yetiştirilen Akkaraman Koyunların çeşitli verim özelliklerinin araştırılması II. Kirli yapağı verimleri, lüle uzunlukları, beden ölçüleri, kuzuların doğum ağırlıkları ve yaşama güçleri. *Fırat Univ Sağlık Bil Derg*, 20, 29-37, 2006.
50. **Mousa-Balabel TM:** The relationship between sheep management and lamb mortality, World Academy of Science, *Engineering and Technology*, 41, 1201-1206, 2010.
51. **Hassen Y, Sölkner J, Fuerst-Walt B:** Body weight of Awassi and indigenous Ethiopian sheep and their crosses. *Small Rumin Res*, 55, 51-56, 2004.

Effects of FGA Sponge and Ovsynch Based Protocols on Reproductive Performance of Fat-tailed Ewes During the Breeding Season

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Summary

The effects of different synchronisation protocols on the reproductive parameters of fat-tailed ewes during the breeding season were studied herein. Seventy-one multiparous fat-tailed ewes were randomly divided into four treatment groups, as follows; Group 1 (long-term FGA): for a total of 20 ewes, 20 mg FGA sponges inserted intravaginally for 14 d, plus an i.m. injection of 400 IU PMSG upon the sponge withdrawal, Group 2 (short-term FGA): for 18 ewes, the insertion of sponges for 8 d, plus the PMSG injection on day 8 upon the sponge withdrawal, Group 3 (Ovsynch): for 17 ewes, an i.m. injection of 0.004 mg GnRH on day zero, followed by the injection of 125 µg PGF_{2α} on day 7 and the second GnRH injection on day 9, and Group 4 (short-term FGA plus Ovsynch): for 16 ewes, the FGA sponge inserted along with the first GnRH injection (as day zero), PGF_{2α} injection on day 7 upon the sponge withdrawal, and the second GnRH injection on day 9. The results showed that; time to onset of oestrus ($P<0.001$), duration of oestrus ($P<0.05$), the rates of pregnancy ($P<0.01$) and lambing ($P\leq 0.001$) differed between the synchronisation groups. Overall, findings of onset/duration of oestrus and pregnancy/lambing rates suggest that; the standard Ovsynch synchronisation programme was markedly inferior to the long- and short-term FGA sponge protocols in fat-tailed ewes during the breeding season.

Keywords: Fat-tailed ewe, Oestrus synchronisation, FGA, Ovsynch

Aşım Mevsimindeki Yağlı Kuyruklu Koyunlarda FGA ve Ovsynch Temelli Senkronizasyon Protokollerinin Üreme Performansı Üzerine Etkileri

Özet

Sunulan çalışmada, aşım sezonundaki yağlı-kuyruklu koyunlarda farklı senkronizasyon protokollerinin reproduktif parametreler üzerine etkileri araştırıldı. Bu amaçla, toplam 71 baş birden fazla doğum yapmış yağlı-kuyruklu koyun rasgele 4 deneme grubuna ayrıldı: Birinci gruptaki koyunlara (n=20, uzun süreli FGA), 20 mg FGA içeren süngerler intravaginal olarak yerleştirildikten sonraki 14. günde uzaklaştırılarak, 400 IU PMSG intramuskuler olarak enjekte edildi. İkinci gruptaki koyunlara (n=18, kısa-süreli FGA), 20 mg FGA içeren süngerler intravaginal olarak yerleştirildikten sonraki 8. günde uzaklaştırılarak, 400 IU PMSG intramuskuler olarak enjekte edildi. Üçüncü gruptaki koyunlara (n=17, Ovsynch grubu), Ovsynch protokolü (0. günde 0.004 mg GnRH, 7. günde 125 µg PGF_{2α} ve 9. günde 0.004 mg GnRH) uygulandı. Dördüncü gruptaki koyunlara (n=16; kısa-süreli FGA + Ovsynch) ise, 20 mg FGA içeren süngerlerin intravaginal olarak yerleştirilmesiyle birlikte 0.004 mg GnRH enjekte edildi. Süngerlerin 7 gün sonra uzaklaştırılmasını takiben 125 µg PGF_{2α} ve 9. günde 0.004 mg GnRH uygulandı. Senkronizasyon grupları arasında, son uygulama-östrus aralığı ($P<0.001$), östrus süresi ($P<0.05$), gebelik ($P<0.01$) ve doğum oranları ($P\leq 0.001$) açısından önemli farklar olduğu gözlemlendi. Sonuç olarak; standart Ovsynch protokolünün, üreme sezonundaki yağlı-kuyruklu koyunlarda klasik kısa ve uzun süreli FGA içeren vajinal sünger uygulamalarına göre gebelik ve doğum oranlarında önemli ölçüde azalmaya yol açtığı kanısına varıldı.

Anahtar sözcükler: Yağlı kuyruklu koyun, Östrus senkronizasyonu, FGA, Ovsynch



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INTRODUCTION

Numerous methods have been used to for improving the reproductive performance in sheep. For this, some researchers have focused on animal husbandry practices (e.g. flushing or the ram effect), while others adopted the administration of certain agents ^[1-3].

Oestrus synchronisation has been practised in sheep industry for almost a half century ^[4]. The methods are based on the control of the lifespan of corpus luteum (by using prostaglandin or progestagen). The most common techniques are progestagen devices, inserted for 12-14 d, followed by an administration of eCG ^[5]. Administrations of intravaginal progestagen (FGA or CIDR), for 10-16 d, followed by intramuscular (i.m.) injection of eCG, have been successfully used ^[6-8]. Progestagens appear to be the most practical hormones of choice. However, a prolonged time of administration could result in low conception rates ^[9-11], as likely to be attributable to the impaired sperm transport *in vivo* ^[12]. Meanwhile, short-term protocols possibly allow for facilitating the managerial tasks, minimising the vaginal discharge and infection risks, and thus increasing the fertility rates. Indeed, short periods of sponge administration, for as short as 5-7 d, have been successful in sheep regardless of breeding season ^[10,13].

Alternatively, a GnRH-based PGF_{2α} protocol has also been reported to be effective in ewes during the breeding season ^[13,14]. Furthermore, their combination with the Ovsynch protocol has also been used in cattle ^[15]. Likewise, this protocol was also used recently in ewes ^[16]. Physiological aspects of Ovsynch protocol may be described as follows: on day zero, the ovulation occurs followed by the progression of corpus luteum or, alternatively, intrafollicular luteinisation takes place after the first GnRH injection. On day 5, the injection of PGF_{2α} induces luteolysis, while the injection of second GnRH on day 7 evokes the ovulation. Indeed, Deligiannis et al.^[16] reported that the majority of animals eventually became pregnant following the artificial insemination performed 36-62 h after the second GnRH. Furthermore, Holtz et al.^[17] comparing progesterone sponge and Ovsynch protocols in goats concluded that the latter protocol could also be an alternative choice with reasonable outcome.

In the literature, there exist only a little information on the effect of Ovsynch protocol or its combination with FGA-impregnated intravaginal sponges in fat-tailed ewes. Therefore, the present study was performed to investigate the reproductive efficiency of Ovsynch and or FGA protocols for oestrus synchronisation during the natural breeding season in fat-tailed Akkaraman and Awassi ewes.

MATERIAL and METHODS

Locations and Animals Used

This study was conducted during the breeding season at the Experimental Research and Practice Farm, Faculty of Veterinary Science, Ankara University, Ankara, Turkey. Ewes were in dry period (not lactating) and clinically healthy.

A total number of 71 fat-tailed ewes (40 Akkaraman and 31 Awassi breed) were used. The animals were 2-5 years old with body weight between 45-55 kg and body condition score of 2.5-3.0 based on the method described by Ucar et al.^[18]. The ewes were kept in open pen (2.5 meter square per ewe) with adequate watering, and offered alfalfa hay *ad libitum* together with a commercial concentrate supplement (400-500 g/head).

Oestrus Synchronisation Protocols

The ewes were randomly divided into four treatment groups, as follows:

Group 1 (Long-term FGA sponge plus PMSG injection): For a total of 20 ewes (11 Akkaraman and 9 Awassi), 20 mg FGA, Fluorogestone acetate, sponges (Chronogest® CR, Intervet, Istanbul, Turkey), inserted intravaginally for 14 d, followed by i.m. injection of 400 IU PMSG, pregnant mare serum gonadotropin (Chronogest/PMSG, Intervet) upon the sponge withdrawal.

Group 2 (short-term FGA sponge plus PMSG injection): For a total of 18 ewes (10 Akkaraman and 8 Awassi), 20 mg FGA sponges inserted intravaginally for 8 d, followed by i.m. injection of 400 IU PMSG upon the sponge withdrawal.

Group 3 (Ovsynch): For a total of 17 ewes (10 Akkaraman and 7 Awassi), i.m. injection of 0.004 mg GnRH analogue, Buserelin (Receptal®, Intervet) on day zero followed by an i.m. injection of 125 µg PGF_{2α} analogue, Cloprostenol (Estrumate®, DIF; Istanbul, Turkey) 7 d later, and finally a second injection of 0.004 mg Buserelin on day 9.

Group 4 (short-term FGA plus Ovsynch): For a total of 16 ewes (9 Akkaraman and 7 Awassi), we implemented the FGA sponge inserted along with the injection of 0.004 mg Buserelin (as day zero), followed by an injection of 125 µg Cloprostenol on day 7 upon the sponge withdrawal and, finally the second injection of Buserelin on day 9.

Oestrus Detection, Mating and Fertility

Oestrus signs were monitored four times a day for 15 min each time as described by Ucar et al.^[8]. The signs were observed following each protocol for 18-72 h with intervals of 6 h. Teaser rams were used for detection of ewes on heat.

Once the ewes were detected in oestrus, they were then separated from the rest of flock and hand-mated with fertility-proven rams ($n=4$, for each breed) used rotationally. Duration of oestrus was recorded as the time starting from the onset of oestrus signs until the rejection of mounting ram, as the end of receptivity. The pregnancy rates as well as the rates of lambing and twinning were recorded after parturition. Ongoing pregnancies were determined by considering those ewes that have previously showed oestrus, mated, and non-returned (no oestrus/no mating again) within the subsequent oestrus cycle (between day 14-21 post-mating/synchronisation).

Statistical Analyses

The present data (mean \pm SEM) from the onset of oestrus signs and duration of oestrus were analysed by one-way analysis of variance (ANOVA). The analyses of oestrus responses, pregnancy rate as well as lambing and twinning rates were performed using Chi square test. A 95% confidence interval was used. Differences between the synchronisation groups were considered statistically different, using the least significant difference ($P<0.05$). Minitab statistical software programme (MINITAB, Version 11.2; Minitab Inc., Pennsylvania, USA) was used for all statistical analyses.

RESULTS

According to our preliminary statistical analyses (ANOVA), considering the breed as co-factor (Akkaraman vs. Awassi), there were no significant differences in any of the reproductive parameters studied. Therefore, all the animals from the two breeds were considered as 'fat-tailed ewes'.

Considering the effect of different oestrus synchronisation groups (Table 1), the onset of oestrus signs, duration of oestrus, pregnancy and lambing rate of ewes were differed significantly between the groups.

The onset of oestrus signs was significantly shorter in the long-term FGA-PMSG group (Group 1) as compared to those in other groups. Also, the duration of oestrus was significantly shorter in the Ovsynch group (Group 3), as compared to those in others.

Additionally, the rates of pregnancy and lambing were significantly lower in the Ovsynch group as compared to those in the long-term FGA-PMSG (Group 1) and short-term FGA-PMSG groups (Group 2). The concerned rates of Ovsynch and short-term FGA plus Ovsynch groups (Group 4) both were also significantly lower than those in the long-term and short-term FGA groups

Finally, for the twinning rates, the values in the Ovsynch protocol were numerically lower as compared to those in others.

DISCUSSION

Our findings indicated that the standard Ovsynch programme was inferior to the conventional FGA-PMSG based on the classical reproductive traits in ewes during the breeding season.

The oestrus rate was relatively higher (87.5%) in the short FGA plus Ovsynch-treated ewes than those (75-82.4%) in other groups. So far, a high oestrus response (75-100%) has been recorded with the FGA-PMSG [19-21]. Indeed, using the FGA-PMSG (control) herein, the oestrus rate was 75%, similar to those (73-77%) in Menze and Dorset ewes [19,22]. A higher (up to 100%) rate was also reported in other breeds [21,23]. Different results might be related to differences in the PMSG dose, the source of progesterone, breed type, etc. The oestrus rate with the short FGA-PMSG was 77.8%, similar to 83.3-88% in Awassi [24] and Ossimi ewes [25]. However, Martemucci and D'Alessandro [26], using the Ovsynch in cross-bred Altamurana ewes in non-breeding season reported a rather lower oestrus rate

Table 1. The effects of different oestrus synchronisation protocols upon the reproductive traits in fat-tailed ewes

Table 1. Yağlı kuyruklu koyunlarda farklı senkronizasyon protokollerinin reproduktif parametreler üzerine etkileri

Parameters Studied	Synchronisation Groups*				Statistics		
	Group 1 (n=20)	Group 2 (n=18)	Group 3 (n=17)	Group 4 (n=16)	F-Ratio	P Value	Significance
Oestrus, %	75.0 \pm 9.9	77.7 \pm 10.1	82.3 \pm 9.5	87.5 \pm 8.5	0.32	0.812	N.S.
Interval to onset of oestrus*, h	37.2 \pm 1.7 ^a	49.7 \pm 1.3 ^b	50.5 \pm 1.3 ^b	51.0 \pm 0.8 ^b	23.77	0.000	P<0.001
Duration of oestrus*, h	27.2 \pm 0.8 ^b	27.0 \pm 0.8 ^b	24.0 \pm 1.2 ^a	27.8 \pm 0.8 ^b	3.30	0.027	P<0.05
Pregnancy rate*, %	93.3 \pm 6.6 ^b	85.7 \pm 9.7 ^b	42.8 \pm 13.7 ^a	57.1 \pm 13.7 ^a	4.51	0.007	P<0.01
Lambing rate*, %	93.3 \pm 6.6 ^b	85.7 \pm 9.7 ^b	35.7 \pm 13.3 ^a	50.0 \pm 13.9 ^a	6.24	0.001	P \leq 0.001
Twinning rate*, %	33.3 \pm 12.6	35.7 \pm 13.3	7.1 \pm 7.1	21.4 \pm 11.4	1.31	0.281	N.S.

* **Group 1:** "Long-term FGA-PMSG"; **Group 2:** "Short-term FGA-PMSG"; **Group 3:** "Ovsynch"; **Group 4:** "Short-term FGA + Ovsynch"; * The number of treated ewes and those in oestrus were 57 only; a total of 14 ewes (of 5, 4, 3 and 2 ewes from Group I-V, respectively) with no signs of oestrus were excluded from the analyses; ^{a,b} Means (\pm SEM) having different superscripts within the same row are significantly different from each other ($P<0.05$); **F-ratio:** Frequency ratio; **P value:** Probability value; **Significance:** Statistical significance; **N.S.:** not significant ($P>0.05$)

(33%), as compared to that in our study (80%). The low rates of oestrus reported might be due mainly to the physiological status and breed type of ewes used therein.

Interval to onset of oestrus upon the sponge withdrawal was markedly shorter in the long-term treatment as compared to others. Herein, the oestrus interval with the long-term FGA-PMSG was 37.2 h, similar to 38.1–44.7 h in Awassi and Dorper ewes [27,28]. Small differences between the durations might be related to a smaller number of animals, shorter duration of sponge administration and a higher dose of PMSG used in the latter [29]. Herein, the interval was 49.7 h after the short-term FGA, similar to 46.2 h in a previous study [30]. However, higher times of 69–70 h [24,25] were also reported elsewhere. Martemucci and D'Alessandro [26], using the Ovsynch reported that the interval was 59 h, while it was 50 h herein. Small differences of oestrus intervals might be due to the individual ewes in different breeds and to different management conditions used.

The duration of oestrus was shorter (24 h) in ewes treated with the Ovsynch, as compared to the other protocols. The durations of oestrus with both the FGA protocols were similar to those in previous studies [27,29]. However, the durations of oestrus observed (24–27.9 h) were shorter than the 34.9 h in Awassi ewes [24], but longer than the 18.7 h in Dorper ewes [28]. Differences might be related mainly to the season as well as to other co-factors (e.g. protocol, feeding, etc.).

The present results of long- or short-term FGA were not markedly different based on the pregnancy, lambing and twinning rates. Likewise, Ustuner et al. [24] reported that both the long- and short-term resulted in similar fertility (22% vs. 20%, resp.) in Awassi ewes during the season. By contrast, the short-term (6 d) treatment resulted in a higher pregnancy rate due likely to the ovulation of newly recruited growing follicles [10]. Therein, however, the rates of pregnancy and lambing with both durations did not differ widely. Meanwhile, pregnancy and lambing rates of Ovsynch, either alone or its combination with short-term FGA, were markedly lower than the FGA-sponge administrations, regardless of its duration. In this respect, we simply consider that, the conventional Ovsynch protocol originally developed for cattle species [15] may be inappropriate for sheep. This could simply be attributable to abrupt species differences in the seasonality and durations of oestrus cycles (21 d vs. 16–17 d) for cattle and sheep, respectively [2]. Apparently, the standard Ovsynch protocol should be modified for the ewe, as having different follicular and luteal dynamics. In a broader sense, the variations might also be due to different synchronisation protocols, the type of hormones and breed used [3,8]. In this respect, we considered that progesterone-primed treatment would mimic the luteal phase likely to be followed by the propagation of follicular development upon the sponge withdrawal. Indeed, the FGA-PMSG presumably ensure both an increase in the

number of females in oestrus [2,8] and the reduction of short cycles especially in late breeders, as leading them to enter into the breeding season more efficiently. In another study, however, similar lambing rates (47–50%) were reported with either the Ovsynch alone or its combination with the short-term FGA [16,31]. This was also the case herein.

The higher pregnancy/lambing rates observed in the FGA-PMSG based groups may be due to the PMSG action. In fact, the Ovsynch protocol mainly acts as the source of exogenous doubling GnRH (mainly provoking the endogenous LH release), while the PMSG acts mainly as the source of exogenous FSH that seems to be more advantageous over the LH *per se*. It was thought that the standard Ovsynch (injections on day zero–7–9) used in ewes might be inappropriate, as reported elsewhere [16]. Hence, it would be logical that both different days of the injections and the administrative sequence of GnRH (before or after the FGA) should be studied further [16]. However, progesterone plus PMSG might be well enough for stimulating the ovarian cycle, eventually leading to a successful lambing at early, mid- or late in the breeding season [3].

Overall, the findings showed that different synchronisation protocols could markedly alter the onset/interval (shortest with the FGA-PMSG) and duration of oestrus as well as the rates of pregnancy and lambing (lowest with the Ovsynch) in fat-tailed ewes during the breeding season. In that, however, there was only a numerical (but not significant) improvement in reproductive parameters (e.g. lambing) as compared to the standard Ovsynch protocol studied herein. Hence, not only the protocol itself but also the follicular/luteal dynamics should be studied further for a better understanding of its *in vivo* regulatory mechanisms of exogenous hormones in sheep. Nevertheless, our study would provide some clues for the future synchronisation trials in sheep. In this respect, we might recommend some modifications (such as studying the durations between GnRH and PGF_{2α} administrations) of present synchronisation protocols to achieve further improvements in the ultimate fertility. Additionally, future investigations of synchronisation with a higher number of animals from other breeds should also comprise an early determination of the plasma progesterone levels in- and outside the breeding seasons.

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REFERENCES

1. Martin GB, Oldman CM, Cognie Y, Pearce DT: The physiological responses of anovulatory ewes to the introduction of rams - A review. *Livestock Prod Sci*, 15, 219–247, 1986.

2. **Ptaszynska M:** Ovine reproduction. In, *Compendium of Animal Reproduction*. 6th Revised ed., Intervet Int. bv (The Netherlands). pp.125-147, 2011.
3. **Yıldız S, Uzun M, Kaya M, Uçar Ö, Çenesiz M:** Effects of rams and luteal or follicular phase ewes on preovulatory LH surge characteristics in ewes. *Turk J Vet Anim Sci*, 28, 669-673, 2004.
4. **Robinson TJ, Moore NW, Holst PJ, Smith JF:** The evaluation of several progestogens administered in intravaginal sponges for the synchronization of estrus in the entire cyclic Merino ewe. In, Robinson TJ (Ed): *Control of the Ovarian Cycle in the Sheep*. White and Bull PTY Ltd. United States. pp.76-91, 1967.
5. **Gordon IR:** Controlled reproduction in sheep and goats. In, *Artificial Control of Estrus and Ovulation*. CAB International, New York, pp.87-89, 1997.
6. **Fukui Y, Ishikawa D, Ishida N, Okada M, Itagaki R, Ogiso T:** Comparison of fertility of estrous synchronized ewes with four different intravaginal devices during the breeding season. *J Reprod Dev*, 45, 337-343, 1999.
7. **Gomez JD, Balasch S, Gomez LD, Martino A, Fernandez N:** A comparison between intravaginal progestagen and melatonin implant treatments on the reproductive efficiency of ewes. *Small Rumin Res*, 66, 156-163, 2006.
8. **Ucar O, Kaya M, Yildiz S, Onder F, Cenesiz M, Uzun M:** Effect of progestagen/PMSG treatment for oestrus synchronisation of Tuj ewes to be bred after the natural breeding season. *Acta Vet Brno*, 74, 385-393, 2005.
9. **Viñoles C, Forsberg M, Banchemo G, Rubianes E:** Effect of long term and short term progestagen treatment on follicular development and pregnancy rate in cyclic ewes. *Theriogenology*, 55, 993-1004, 2001.
10. **Viñoles C, Meikle A, Forsberg M, Rubianes E:** The effect of sublethal levels of exogenous progesterone on follicular dynamics and endocrine patterns during the luteal phase of the ewe. *Theriogenology*, 51, 1351-1361, 1999.
11. **Martin GB, Milton J, Davidson R, Banchemo-Hunzicker G, Lindsay D, Blache D:** Natural methods for increasing reproductive efficiency in small ruminants. *Anim Reprod Sci*, 82-83, 231-245, 2004.
12. **Hawk HW, Cooper BS:** Sperm transport into the cervix of the ewes after regulation of estrus with prostaglandin or progestogen. *J Anim Sci*, 44, 638-643, 1977.
13. **Ataman MB, Aköz M:** GnRH-PGF_{2α} and PGF_{2α}-PGF_{2α} synchronization in Akkaraman cross-breed sheep in the breeding season. *Bull Vet Inst Pulawy*, 50, 101-104, 2006.
14. **Beck NFG, Jones M, Davies B, Peters AR, Williams SP:** Oestrus synchronization in ewes: The effect of combining a prostaglandin analogue with a GnRH agonist (buserelin). *Anim Sci*, 62, 85-87, 1996.
15. **Pursley JR, Mee MO, Wiltbank MC:** Synchronization of ovulation in dairy cows using PGF_{2α} and GnRH. *Theriogenology*, 44, 915-923, 1995.
16. **Deligiannis C, Valasi I, Rekkas CA, Goulas P, Theodosiou E, Lainas T, Amiridis GS:** Synchronization of ovulation and fixed time intrauterine insemination in ewes. *Reprod Domest Anim*, 40, 6-10, 2005.
17. **Holtz W, Sohnrey B, Gerland M, Driancourt MA:** Ovsynch synchronisation and fixed-time insemination in goats. *Theriogenology*, 69, 785-792, 2008.
18. **Ucar O, Cenesiz M, Kaya M, Yildiz S:** Effects of body condition score upon reproduction in sheep (in Turkish, review). *Bultendif Veteriner Bulten*. 29, 2-5, 2008.
19. **Mutiga ER, Mukasa-Mugerwa E:** Effect of the method of estrus synchronization and PMSG dosage on estrus and twinning in Ethiopian Menze sheep. *Theriogenology*, 38, 727-734, 1992.
20. **Ataman MB, Aköz M, Fındık M, Saban E:** Geçiş dönemi başındaki Akkaraman melezi koyunlarda farklı dozda flourogestene acetate, norgestomet ve PGF_{2α} ile senkronize östrüslerin uyarılması. *Kafkas Univ Vet Fak Derg*, 15 (5): 801-805, 2009.
21. **Rekik M, Lassoued N, Yacoubi C:** Reproductive performances in ewe lambs of the Queue Fine de l' Quest breed and their D'Man crosses following synchronisation. *Small Rumin Res*, 45, 75-78, 2002.
22. **Rajamahendran R, Raniowski J, Ravindran V:** Effect of PMSG and ram contact on the reproductive performance of progestagen-treated ewes during breeding and anestrus seasons. *Small Rumin Res*, 10, 341-347, 1993.
23. **Hashemi M, Safdarian M, Kafi M:** Estrous response to synchronisation of estrus using different progesterone treatments outside the natural breeding season in ewes. *Small Rumin Res*, 65, 279-283, 2006.
24. **Ustuner B, Gunay U, Nur Z, Ustuner H:** Effects of long and short-term progestagen treatments combined with PMSG on oestrus synchronisation and fertility in Awassi ewes during the breeding season. *Acta Vet Brno*, 76, 391-397, 2007.
25. **Ali A:** Effect of time of eCG administration on follicular response and reproductive performance of FGA-treated Ossimi ewes. *Small Rumin Res*, 72, 33-37, 2007.
26. **Martemucci G, D'Alessandro AG:** Estrous and fertility responses of dairy ewes synchronized with combined short term GnRH, PGF_{2α} and estradiol benzoate treatments. *Small Rumin Res*, 93, 41-47, 2010.
27. **Özyurtlu N, Küçükaslan I, Çetin Y:** Characterization of oestrous induction response, oestrous duration, fecundity and fertility in Awassi ewes during the non-breeding season utilizing both CIDR and intravaginal sponge treatments. *Reprod Domest Anim*, 45, 464-467, 2010.
28. **Zelege M, Greyling JPC, Schwalbach LMJ, Muller T, Erasmus JA:** Effect of progestagen and PMSG on oestrus synchronisation and fertility in Dorper ewes during the transition period. *Small Rumin Res*, 56, 47-53, 2005.
29. **Turk G, Gur S, Sonmez M, Bozkurt T, Aksu EH, Aksoy H:** Effect of exogenous GnRH at the time of artificial insemination on reproductive performance of Awassi ewes synchronized with progestagen-PMSG-PGF_{2α} combination. *Reprod Domest Anim*, 43, 308-313, 2008.
30. **Ungerfeld R, Rubianes E:** Short term primings with different progestogen intravaginal devices (MAP, FGA and CIDR) for ewes eCG-estrous induction in anestrus ewes. *Small Rumin Res*, 46, 63-66, 2002.
31. **Titi HH, Kridli RT, Alnimer MA:** Estrus synchronization in sheep and goats using combinations of GnRH, progestagen and PGF_{2α}. *Reprod Domest Anim*, 45, 594-599, 2010.

The Axon Number of the Rat Sciatic Nerve: A Stereological Study

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Summary

This study was designed to determine with stereological methods the number of axons in the sciatic nerve as a result of peripheral nerve blockage following injection of lidocaine and prilocaine into hind limb muscle. Nine adult female Sprague-Dawley rats weighted 150-200 g were used in the study. Ketamine was employed intraperitoneal to the rats as 50 mg/kg. The prilocaine and lidocaine were equally (5 mg/ml) injected to the left and right limbs of the 4 rats. As a control group, isotonic sodium chloride (0.9%) was performed as 0.2 ml into the extremities of the remaining 5 rats. When the application completed, surrounding muscles and soft tissues as well as sciatic nerve were dissected as 0.5x1.0 cm length. In order to determine neurotoxic effects of lidocaine and prilocaine on sciatic nerve, the number of axons was computed by unbiased stereological method. Following of the square root transformation, Mann-Whitney U test was performed to compare groups. When the unbiased comparisons were executed to both groups, the effects of lidocaine and prilocaine on sciatic nerve were found statistically non-significant.

Keywords: Axons number, Sterology, Rat, Sciatic nerve

Sıçan Siyatik Sinir Akson Sayısı: Stereolojik Bir Çalışma

Özet

Bu çalışma, Lidokain ve prilokain'in arka ekstremité kası enjeksiyonunu takiben oluşan periferik sinir blokajı sonucunda siyatik sinirde akson sayısını stereolojik metotlarla belirlemek için planlandı. Bu çalışmada 150-200 g ağırlığında dokuz adet Sprague-Dawley tipi erişkin dişi sıçan kullanıldı. Sıçanlara Ketalar 50 mg/kg intraperitoneal uygulandı. Dört adet dişi sıçanın sağ ekstremitelerine eşit miktarda prilokain 5 mg/ml (0.2 ml) ve sol ekstremitelerine lidokain 5 mg/ml (0.2 ml) verildi. Kontrol grubu olarak %0.9 izotonik sodyum klorür kalan 5 adet sıçan ekstremitelerine 0.2 ml uygulandı. Uygulama sona erince çevre kas ve yumuşak dokular siyatik sinir ile beraber 0.5x1.0 cm uzunlukta çıkarıldı. Lidokain ve prilokainin siyatik sinir üzerine nörotoksik etkilerini ortaya koymak için tarafsız stereolojik metod ile akson sayısı hesaplandı. Bu özellik bakımından denek ve kontrol gruplarını karşılaştırmada karekök transformasyonu yapıldıktan sonra Mann-Whitney U testi kullanıldı. Her iki grup arasında yapılan karşılaştırma tarafsız stereolojik hesaplamalarla yapıldığında sıçanlara uygulanan Lidokain ve prilokain akut olarak siyatik sinir akson sayılarına etkileri istatistiksel olarak anlamlı bulunmadı.

Anahtar sözcükler: Akson sayısı, Stereoloji, Rat, Siyatik sinir

INTRODUCTION

Regional anesthesia with a nerve -blocking is a useful technique for day-stay surgery to the hand ^[1]. Regional anesthetic techniques provide efficient postoperative analgesia ^[2,3]. The most often used local anesthetics for regional anesthesia in Europe is prilocaine and lidocaine. They have a relatively short duration of action and are

the least toxic of the amino-amide local anesthetics ^[4,5]. Peripheral nerve injury is a rare but sometimes has the devastating complication of regional anesthesia.

Neurotoxicity of local anesthetics *per se* is not new although it is largely ignored by clinicians ^[6]. Laboratory



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evidence suggests that local anesthetic neurotoxicity is at least partially dose and concentration dependent ^[7]. Studies of local anesthetic-induced nerve injury have eliminated several possible mechanisms of toxicity. For example glucose included in the local anesthetic solution is not neurotoxic which suggests that toxicity results from an action of the local anesthetic itself ^[8]. Therefore we investigated stereological the acute neurotoxic effects of lidocaine and prilocaine on sciatic nerve using injection technique of continuous peripheral nerve blockade.

MATERIAL and METHODS

Animals

This study was approved by the Animal Use Ethics Commission from Yüzüncü Yıl University (2003/03-01). Nine female rats (150-200 g) were kept in a standard plastic cage on sawdust bedding in an air-conditioned room (22°C) under a 12/12 h light/dark cycle and fed *ad-libitum*.

Chemicals

The animals were anesthetized by ketamine hydrochloric (50 mg/kg) intraperitoneally without muscle relaxation and placed in prone position. The rats were separated as a control (n=5) and experiment (n=4) groups. Under the general anesthesia by using peripheric nerve-stimulator (2 Hz and 0.3 ms B/Braun Stimuplex) and a nerve stimulator needle (22 G x 2-0.7 x 50 mm) skin was incised. Proximal hindlimb muscle and sciatic nerve were exposed. The nerve catheters (20 gauges) were inserted into the sciatic nerve sheaths and fixed bilaterally into two extremities along the sciatic nerve trace. After this procedure skin and subcutaneous tissue were closed by single stitches.

Four animals after placing the catheters were administered lidocaine (5 mg/ml) in left side and prilocaine (5 mg/ml) in right side with a rate of 0.2 ml for a total period of 6 h by opening 1 mA. In control group (n=3) the animals were treated with normal saline by same rate and doses on both sides. After six hour from this procedure, the sciatic nerve with 0.5×1.0 cm volume was cut out with surrounding tissues by removing the stitches and withdrawing the catheters. The animals were sacrificed.

Histological Analysis

The entire sciatic nerves were removed bilaterally en bloc from all rats. The nerves were stretched to in situ length by pinning onto a card and then fixed with 2.5% glutaraldehyde in 0.1 M sodium cacodylate (pH 7.4) for 4-6 h in 4°C. After fixation nerve samples about 0.5 cm long that were taken from the nerves were rinsed in 0.1 M sodium cacodylate (pH 7.4) twice. Specimens were post fixed in 1% osmium tetroxide for 1 h dehydrated in an ascending alcohol series and propylene oxide took into for a 50:50 mixture propylene oxide and Epon 812 for 24 h. These

procedures were completed by embedding the tissues 48 h in Epon Embedding Kit (Fluka Chemie GmbH, Switzerland). Semi-thin and ultra-thin sections (of 1 mm and 90 nm thickness, respectively) were cut by an ultramicrotome (Super Nova Reichert-Yung, Austria) and stained with 1% toluidine blue.

Stereology

Stereological analyses of sciatic nerves were done according to principles described previously ^[9]. A manual stereological workstation composed of a digital camera (Nikon COOLPX5400, Tokyo, Japan) a manual dial indicator controlled specimen stage and a light microscope (Nikon Microphot-FX Tokyo, Japan) was used for axon number counting ^[10]. To obtain an estimation of total myelinated axon number in an unbiased manner the axon profiles in the nerve cross-section are sampled with equal probability regardless of shape size orientation and location that means each sampled item was selected with a systematic random manner ^[11]. For this aim we used an area fraction approach in the application area of unbiased counting frame was 900 mm². A counting frame was placed on to a monitor and the sampled area was chosen by a systematic uniform random manner via dial indicator controlled specimen stage. Meander sampling of each sectioned nerve profiles was done in 70 µm-70 µm step size in a systematic-random manner. This ensures that all locations within a nerve cross-section were equally represented and that all axon profiles were sampled with an equal probability regardless of shape size orientation and location ^[12,13].

Descriptive statistics were presented as Mean, Median SD, Minimum and Maximum values for axon count. After square-root transformation Mann-Whitney U test was used to compare treatment and control groups for this trait 5% values was considered to statistical significant; MINITAB statistical pocket programmed was used for all statistical.

RESULTS

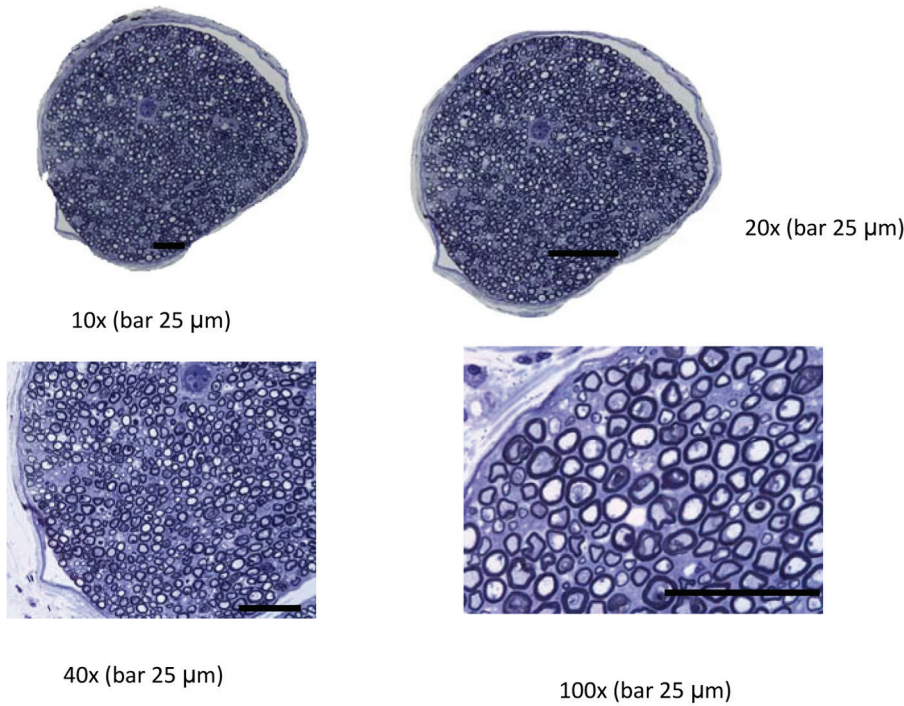
In our study axon number of sciatic nerve was observed on histological sections from the region applied isotonic injection in both treated and control groups. The neurotoxicity was not observed in both lidocaine and prilocaine group statistically (P>0.05) (Table 1, Fig 1).

DISCUSSION

Peripheral nerve blockade with a local anesthetic provides excellent pain relief but its clinical utility for the treatment of acute or postoperative pain is sometimes limited by a short duration of effect ^[14,15]. After placing the nerve catheter it is important to provide fast onset of the block to improve the workflow within the operating theatre. This aim can be reached by using a drug with fast onset ^[16].

Table 1. Descriptive statistics of the axon number of the groups treated and control ($P>0.05$)**Tablo 1.** Denek ve kontrol gruplarının akson sayılarına ait tanımlayıcı istatistik ($P>0.05$)

Groups	N	Mean	SE Mean	StDev	Minimum	Median	Maximum	CV
Lidokain	4	59.74	3.67	7.34	52.70	58.10	70.07	0.1228
Control Left	5	69.53	5.38	12.03	55.46	76.93	80.32	0.1730
Control Right	5	70.09	4.12	9.20	58.75	71.42	83.19	0.1313
Prilokain	4	54.20	3.59	7.17	47.82	52.25	83.19	0.1323

**Fig 1.** Cross-section of sciatic nerve**Şekil 1.** Siyatik sinirin enine kesiti

Local anesthetics are a group of drugs defined by their ability to prevent sodium entry into axons thereby preventing the generation of propagated action potentials in axons. However they have other actions such as prevention of axonal sprouting and effects on G-protein-coupled receptors and on conductance of ions in addition to sodium that might be important in the management of pain [17]. The transport of the amide local anesthetics such as lidocaine and prilocaine is by passive diffusion this transport is dependent on the pH across biological membranes in the case of the amide local anesthetics. Higher concentrations of the anesthetic will be found in the tissue or compartment with lower pH [18].

Prilocaine is medium-long-acting local anesthetics with a fast onset of action [16]. Prilocaine stabilizes the neuronal membrane by inhibiting the ionic fluxes required for the initiation and conduction of impulses thereby effecting local anesthetic action. Local anesthetics bind selectively to the intracellular surface of sodium channels to block influx of sodium into the axon [19].

Studies of clinical adverse effects of intrathecal local anesthetics suggest cauda equina syndrome results from local anesthetic neurotoxicity [20]. Lidocaine toxicity is not

the result of an immediate and irreversible breakdown in the integrity of the plasma membrane as would be reflected by a rapid and permanent loss in membrane potential. The mechanism of the lidocaine-induced depolarization currently is unknown but may reflect the simultaneous blockade of ion channels and pumps responsible for the maintenance of neuronal resting potential [21].

Peripheral nerve injury is occurs as a complication of regional anesthesia. Ischemia is one of the causative mechanisms. This may result from changes in peripheral blood flow caused by the local anesthetic itself and/or a vasopressor adjuvant. Peripheral nerves have a dual blood supply of intrinsic exchange vessels in the endoneurium and an extrinsic plexus of supply vessels in the epineurial space that crosses the perineurium to anastomose with the intrinsic circulation. The extrinsic supply is known to be responsive to adrenergic stimuli [22].

Persistent reductions in peripheral nerve blood flow (NBF) can lead to pathological changes in the structure of nerve fibers and their supporting cells [23].

The intravenous regional anesthesia (IVRA) is a technique suitable for hand and forearm surgery and

safe when performed with care by expert practitioners. The most often used local anaesthetic for IVRA in Europe is prilocaine. It has a relatively short duration of action similar to that of lidocaine ^[4] and is the least toxic of the amino-amide local anesthetics ^[5].

Although the axonal membrane repolarizes after wash of lidocaine an irreversible conduction block was observed in axons after 3 min exposure of the axon to lidocaine at concentrations as low as 40 mM ^[24]. Concentration dependence of lidocaine induced irreversible conduction loss. That we observed recovery of soma action potential after exposure of DRG neurons to even higher concentrations of lidocaine raises the possibility that different mechanisms underlie neuronal death and loss of conduction. Because an increase $[Ca^{++}]$ can disrupt cytoskeleton ^[25,26] and disruption of cytoskeleton can decrease excitability ^[27]. Lidocaine-induced increase in $[Ca^{++}]$ possibly could contribute to both cell death and long-lasting conduction failure. Regional anesthesia-induced nerve injury may in fact require a combined mechanical and chemical insult ^[28].

In rat sciatic nerve relative neural toxicity and relative motor nerve conduction blockade were assessed for two amide-linked local anesthetics (etidocaine and lidocaine) and two ester-linked local anesthetics (chlorprocaine and procaine). The nerve fiber injury and edema were assayed by light microscopic examination of nerve tissue sampled two days after perineural (next to the sciatic nerve) injection of various concentrations of the local anesthetics. Both nerve injury and edema increased with concentration of local anesthetics but injury was frequently present in nerve fascicles with little or no edema. In previous studies the amplitude of the electrical activity elicited from the interosseous muscles of the foot following ipsilateral electrical stimulation at the sciatic notch was monitored for up to are minutes to assess the extent of motor nerve blockade. The resulting low concentration-response curves were analyzed for differences in potency. Both for injury and for conduction block the order of decreasing potency was etidocaine, lidocaine, chlorprocaine and procaine. These results are not consistent with the proposal that ester-linked agents are more likely than other local anesthetic agents to cause nerve injury ^[29]. With specific regard to lidocaine neurotoxicity an irreversible block of impulse conduction was induced in the frog sciatic nerve within a clinical concentration range of 1% to 2% lidocaine ^[24]. In rats increasing doses of intrathecal 5% hyperbaric lidocaine resulted in persistent sacral sensory deficits associated with motor weakness ^[30]. All axons counts were based on the gold standard ^[31] and fractionators technique ^[12,32].

In conclusion we found that the comparison between the infusion of lidocaine and/or prilocaine and the placebo showed no significant difference in the histological axon numbers ($P > 0.05$).

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REFERENCES

- Petronella R, Janzen M, Vipond AJ, Bush DJ, Hopkins PH:** A comparison of 1% prilocaine with 0.5% ropivacaine for outpatient-based surgery under axillary brachial plexus block. *Anesth Analg*, 93, 187-191, 2001.
- Richman JM, Liu SS, Courpas G, Wong R, Rowlingson AJ, McGready J, Cohen SR, Wu CL:** Does continuous peripheral nerve block provide superior pain control to opioids? A meta-analysis. *Anesth Analg*, 102, 248-257, 2006.
- Block BM, Liu SS, Rowlingson AJ, Cowan AR, Cowan JA, Wu CL:** Efficacy of postoperative epidural analgesia: A meta-analysis. *JAMA*, 290, 2455-2463, 2003.
- Valli H, Rosenberg PH, Hekali R:** Comparison of lidocaine and prilocaine for intravenous regional anaesthesia of the whole lower extremity. *Reg Anaesth*, 12, 128-134, 1987.
- Truant AP:** Local anesthetic and toxicologic properties of citanest. *Acta Anaesthesiol Scand*, 16, 19-22, 1965.
- Lambert LA, Lambert DH, Strichartz GR:** Irreversible conduction block in isolated nerve by high concentrations of local anesthetics. *Anesthesiology*, 80, 1082-1093, 1994.
- Ready LB, Plumer MH, Haschke RH:** Neurotoxicity of intrathecal local anesthetics in rabbits. *Anesthesiology*, 63, 364-370, 1985.
- Sakura S, Chan VW, Ciriales R, Drasner K:** The addition of 7.5% glucose does not alter the neurotoxicity of 5% lidocaine administered intrathecally in the rat. *Anesthesiology*, 82 (1): 236-240, 1995.
- Larsen JO:** Stereology of nerve cross-sections. *J Neurosci Methods*, 85, 107-118, 1998.
- Canan S, Aktas A, Ulkay MB, Colakoglu S, Ragbetli MC Ayyildiz M, Geuna S, Kaplan S:** Prenatally expose to a non-steroidal anti-inflammatory drug or saline solution impairs sciatic nerve morphology: A stereological and histological study. *Int J Develop Neurosc*, 26, 733-738, 2008.
- Gundersen HJG, Jensen EB, Kieu K, Nielsen J:** The efficiency of systematic sampling in stereology-reconsidered. *J Microsc*, 193, 199-211, 1999.
- Gundersen HJG:** Stereology of arbitrary particles. A review of unbiased number and size estimators and the presentation of some new ones in memory of William R. Thompson. *J Microsc*, 143, 3-45, 1986.
- Geuna S, Tos P, Battiston B, Guglielmone R:** Verification of the two dimensional disector a method for the unbiased estimation of density and number of myelinated nerve fibers in peripheral nerves. *Ann Anat*, 182, 23-34, 2000.
- Duncan L, Wildsmith JAW:** Liposomal local anaesthetics. *Br J Anaesth*, 75, 260-261, 1995.
- Kuzma PJ, Kline MD, Calkins MD, Staats PS:** Progress in the development of ultra-long-acting local anesthetics. *Reg Anesth*, 22, 543-551, 1997.
- Scott DB, McClure JB, Giasi RM, Seo J, Covino BG:** Effects of concentration of local anesthetic drugs in extradural block. *Br J Anaesth*, 52, 1033-1037, 1980.
- Heavner JE:** Local anesthetics. *Curr Opin Anaesthesiol*, 20 (4): 336-342, 2007.
- Cosovic E, Mornjakovic Z, Susko I, Alicelebic S:** Quantification of some qualitative changes of the rabbit sciatic nerve after the application of lidocaine into the mesoneurium. *Med Arh*, 61 (4): 207-211, 2007.
- Sadove MS, Jobgen SE, Heller NF, Rosenberg R:** Methemoglobinemia-An effect of a new local anesthetic L-67 (Prilocaine). *Acta Anesth Scand*, 9 (16): 175-182, 2008.
- Auroy Y, Narchi P, Messiah A, Litt L, Rouvier B, Samii K:** Serious

complications related to regional anesthesia: Results of a prospective survey in France. *Anesthesiology*, 87, 479-486, 1997.

21. Lambert DH, Strichartz GR: Every problem is an opportunity or one person's poison is another person's remedy. *Reg Anesth Pain Med*, 23, 3-6, 1988.

22. Myers RR, Heckman HM: Effects of local anesthesia on nerve blood flow: Studies using lidocaine with and without epinephrine. *Anesthesiology*, 71, 757-762, 1989.

23. Myers RR, Kalichman MW, Reisner LS, Powell HC: Neurotoxicity of local anesthetics: Altered perineural permeability edema and nerve fiber injury. *Anesthesiology*, 64, 29-35, 1986.

24. Bainton CR, Strichartz GR: Concentration dependence of lidocaine-induced irreversible conduction loss in frog nerve. *Anesthesiology*, 81, 657-667, 1994.

25. Neely MD, Gesemann M: Disruption of microfilaments in growth cones following depolarization and calcium influx. *J Neurosci*, 14, 7511-7520, 1994.

26. Waxman SG, Black JA, Ransom BR, Stys PK: Protection of the axonal cytoskeleton in anoxic optic nerve by decreased extracellular calcium. *Brain Res*, 614, 137-145, 1993.

27. Sakai H, Matsumoto G, Murofushi H: Role of microtubules and axonin in membrane excitation of the squid giant axon. *Adv Biophys*, 19, 43-89, 1985.

28. Gentili F, Hudson AR, Hunter D, Kline DG: Nerve injection injury with local anesthetic agents: A light and electron microscopic fluorescent microscopic and horseradish peroxidase study. *Neurosurg*, 6, 263-272, 1980.

29. Kalichman MW, Moorhouse DF, Powell HC, Myers RR: Relative Neural toxicity of local anesthetics. *J Neuropathol Exp Neurol*, 52, 3, 1993.

30. Drasner K, Sakura S, Chan VW, Bollen AW, Ciriales R: Persistent sacral sensory deficit induced by intrathecal local anesthetic infusion in the rat. *Anesthesiology*, 80, 847-852, 1994.

31. Kaplan S, Geuna S, Ronchi G, Ulkay MB, von Barthold CS: Calibration of the stereological estimation of the number of myelinated axons in the rat sciatic nerve: A multicenter study. *J Neurosci Methods*, 187 (1): 90-99, 2010.

32. Altunkaynak ME, Altunkaynak BZ, Unal S, Vuraler O, Unal B: Morphological alterations of sciatic nerve during development from rat fetuses to adults: A histological, stereological and embryological study. *Kafkas Univ Vet Fak Derg*, 15 (5): 687-697, 2009.

The Impacts of Prebiotic Supplementation on Humoral and Cell-Mediated Immune Responses of Broilers under Combined Stresses Caused by Feed Restriction and *Salmonella enteritidis* Challenge

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Summary

This experiment was conducted to investigate the impacts of prebiotic on humoral and cell-mediated immune responses of broilers under combined stresses caused by feed restriction and *Salmonella enteritidis* challenge. 320 male broilers (Ross 308) at a 23 factorial arrangement in randomized complete block design were divided into 8 treatment groups with 4 replicates per each. The Impact of these factors and their interaction on cell-mediated immune response to cutaneous basophil hypersensitivity (CBH) test, antibody titer response against Newcastle disease virus (NDV) and sheep red blood cell (SRBC), and the relative weight of lymphoid organs were measured. Prebiotic caused an increase and feed restriction caused a decrease in cell-mediated immune responses on days 15 and 30 respectively, and interaction among the factors was differ ($P<0.05$). Also, factors' interaction had a significant effect on antibody titer response against NDV ($P<0.05$) at day 27. *Salmonella* and feed restriction significantly reduced the relative weight of thymus at day 21 and bursa of Fabricius at day 42, respectively ($p < 0.05$). Differences among other measured parameters were not significant ($P>0.05$).

Keywords: Prebiotic, Feed restriction, *Salmonella*, Humoral immune response, Cell-mediated immune response

Prebiyotik İlavesinin Yem Kısıtlaması ve *Salmonella enteritidis* İnokulasyonu İle Kombine Stres Altındaki Broylerlerde Sıvısal ve Hücreyel İmmün Yanıt Üzerine Etkileri

Özet

Bu çalışma, prebiyotik ilavesinin yem kısıtlaması *Salmonella enteritidis* inokulasyonu ile kombine stres altındaki broylerlerde sıvısal ve hücreyel immün yanıt üzerine etkileri araştırmak üzere gerçekleştirildi. Rastgele tam blok tasarım içinde 23 faktoriyel düzenlemede 320 erkek boyler (Ross 308) 4 tekrar olmak üzere 8 tedavi grubuna bölündü. Bu faktörlerin ve onların etkileşiminin hücreyel immün cevap üzerin etkilerini belirlemek üzere kutanöz bazofil aşırı duyarlılık testi (CBH), Newcastle hastalığı (NDV) virüsü ve koyun eritrositlerine (SRBC) karşı gelişen antikor titreleri ve lenfoid organların göreceli ağırlıkları belirlendi. Hücreyel immün yanıtı prebiyotik 15. günde arttırırken yem kısıtlamasını 30.günde azalttığı ve faktörler arası etkileşimin farklı ($P<0.05$) olduğu gözlemlendi. Ayrıca, faktörlerin etkileşiminin 27. gündeki NDV'ye karşı gelişen antikor titresinin gelişimi üzerine önemli etkisinin ($P<0.05$) olduğu belirlendi. *Salmonella* ve yem kısıtlamasının, timus ve bursa fabricius'un göreceli ağırlığını, sırasıyla 42. ve 21. günde azalttığı belirlendi ($P<0.05$). Diğer ölçülen parametreler arasında farklılıklar anlamlı bulunmadı ($P>0.05$).

Anahtar sözcükler: Prebiyotik, Yem kısıtlaması, *Salmonella*, Sıvısal bağışıklık cevabı, Hücreyel bağışıklık cevabı

INTRODUCTION

In recent years, feed restriction has been used for improving broilers' performance and their carcass quality. One of the common methods is restriction in the starter

feeding period on the base of compensatory growth. It has been reported that decrease in broilers' primary growth reduces the rate of metabolic reactions; therefore there



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is less need to oxygen, and the metabolic disorders such as ascites and SDS (sudden death syndrome) resulted from fast growth are prevented ^[1]. Beside the advantages, feed restriction has some disadvantages too. Limitations in access to food make stress for the chickens, and this stress changes the microbial population of their gut. It has been shown that hunger in birds reduces the amount of *Lactobacilli* spp. in the crop ^[2]. Also there are reports on the suppressing effects of feed restriction on some immune system parameters in chickens ^[3,4].

Furthermore, broilers commonly meet several internal and external stressors like stocking density, temperature, transportation, feed contamination and microbial infections ^[5]. Among the microbial infections, *Salmonella* spp. is one of the main, and it is the leading cause of human food-borne infections associated with consumption of poultry products in the world ^[6].

The reduced immunocompetence caused by feed restriction beside the effects of bacterial enteric pathogens like *Salmonella* spp. make severe stresses in broilers. In such a condition, using additives such as prebiotics for improving the immune function and gut microflora is helpful. Prebiotics beneficially affect the host by making changes in the population of gut microflora in favor of useful bacteria ^[7] as well as increasing its humoral and cell-mediated immune responses ^[8]. In addition, prebiotics help the host resist against the bacterial infections ^[9].

Several studies have been done on the useful effects of prebiotics on infections ^[10,11] and the negative effects of feed restriction on immune system ^[3,4]. Gursoy et al. ^[12] reported that exposure to combined stresses may have synergistic, additive, or adaptive effects on the immune system parameters. In the literature, there was no report concerning the effect of dietary prebiotic on immune responses of broilers under combined stresses caused by infection and feed restriction. Therefore, our purpose in this study was to investigate the effect of prebiotic supplementation on immune system parameters of broiler chickens under feed restriction and *Salmonella* challenge.

MATERIAL and METHODS

Birds, Diets, and Management

Three hundred twenty 1-d-old male broilers (Ross 308) were obtained from a commercial hatchery. At a 2³ factorial arrangement in randomized complete block design, they were divided into 8 treatment groups with 4 replicates and 10 broilers in each replicate, and were then put in special cages. Environmental temperature in the first week of life was 32°C and decreased to 20°C until the end of the experiment. During the first week, 22 h of light was provided with a reduction to 20 h afterward. Chickens had access to *ad libitum* water and a diet based on corn

and soybean meal which provided 3.000 kcal/kg ME and 21.7% CP up to day 21, and 3075 kcal/kg ME and 19.25% CP from day 21 to day 42. This basal diet was unmedicated.

Prebiotic Supplementation

The prebiotic was gained from TechnoMOS (Biochem, Lohne, Germany). It was used at the amount of 0.1% of the diet. This product consists of mannanoligosaccharide (MOS) and β -1,3-Glucan and has been obtained from *Saccharomyces cerevisiae*.

Feed Restriction Procedure

From day 7 to day 14, the daily feed intake of the groups getting *ad libitum* consumption was measured, and 80% of this feed was given to feed restricted treatment groups the next day.

Salmonella Culturing, Counting and Challenge

Salmonella enteritidis (PTCC 1709) was provided freeze-dried from the Persian Type Culture Collection (IROST, Tehran, Iran) isolated from the liver of chickens. Freeze-dried inoculum was grown in nutrient broth media at 37°C for 8 h and passed to fresh nutrient broth for 3 incubation periods. Counting colony-forming units (cfu) was done with Neobar lam at day 10. After counting, chickens in the challenged groups received 2.6×10^5 cfu/chick of passaged medium through oral gavage with utilizing micropipette on the same day.

Treatment Groups

The treatment groups included 1. a control group with no supplementation and challenge, 2. a group under feed restriction (Control + R), 3. a *Salmonella* challenged group (Control + S), 4. a group under feed restriction and *Salmonella* challenge (Control + R + S), 5. a prebiotic treated group, 6. a prebiotic treated and feed restricted group (Prebiotic + R), 7. a prebiotic treated under *Salmonella* challenge (Prebiotic + S), 8. a prebiotic treated under feed restriction and *Salmonella* challenge (Prebiotic + R + S).

Immunological Tests

Cell-mediated immune response was assessed according to the method of Corrier and DeLoach ^[13] by a cutaneous basophil hypersensitivity (CBH) test using phytohemagglutinin P (PHA-P) produced by Gibco company (Gibco, Invitrogen Corporation, Scotland, UK). At days 15 and 30, the toe web of the right foot (2 chickens per pen) was measured with a Caliper with an accuracy of 0.01 mm. Immediately after measurement, 100 μ g of PHA-P (suspended in 0.1 mL of PBS) was injected into the toe web. The toe web swelling was measured 24 h after injection. The response was measured by subtracting the skin thickness of the first measurement from the skin thickness 24 h after dermal injection. At day 7, vaccination against Newcastle disease virus (NDV) was done by instilling

B1 strain of NDV (Intervet International BV, Boxmeer, Netherlands) in the chickens' eyes, and 2 birds per each pen were injected in breast muscle by 0.1 ml of 5% sheep red blood cell (SRBC) diluted in PBS. The chickens were bled from the wing vein at days 17 and 27, and sera were collected individually in separate sterile vials. The hemagglutination inhibition and hemagglutination tests were done to determine the antibody titer response against NDV^[14] and SRBC^[15], respectively.

At days 21 and 42, a chicken from each replicate pen was weighed and then killed by cervical cutting. Carcasses were dissected immediately after euthanasia. Thymus, bursa of Fabricius, and spleen were then precisely removed and weighed separately on a sensitive digital scale.

Statistical Analysis

All data were subjected to GLM procedures of SPSS version 18 software as a 2³ factorial arrangement of treatments in randomized complete block design that included prebiotic supplementation, feed restriction and *Salmonella* challenge as the main factors and their respective interactions. The treatment means were compared by Duncan's multiple range tests. Probability values of less than 0.05 ($P < 0.05$) were considered significant.

RESULTS

As it is shown in *Table 1*, there were significant differences among treatments regarding the cell-mediated immune

response to CBH test ($P < 0.05$). Prebiotic caused an increase at day 15, and feed restriction caused a decrease at day 30 in toe web thickness, and interaction among the factors was significant ($P < 0.05$). There were no significant differences among treatments and factors' interaction regarding antibody titer responses against SRBC at days 17 and 27, and to NDV at day 17 ($P > 0.05$); but there were significant differences among factors' interaction regarding antibody titer response against NDV at day 27 ($P < 0.05$).

Table 2 shows the relative weight of lymphoid organs at days 21 and 42. *Salmonella* challenge had a significant effect on thymus relative weight at day 21 ($P < 0.05$). Feed restriction significantly reduced the bursa of Fabricius at day 42 ($P < 0.05$). There were no significant differences among treatments and factors' interaction regarding relative weight of spleen at days 21 and 42 ($P > 0.05$).

DISCUSSION

In this study, prebiotic had advantageous and feed restriction had disadvantageous effects on cell-mediated immune response to PHA-P injection. Savino^[3] expressed that severe feed restriction can cause significant decrease of mitogenic lymphocyte responses in vitro. Also in the study done by Hangalapura et al.^[4], feed restriction reduced cell-mediated immune response which is likely because the cellular components of immune system are energy demanding. Savino and Dardenne^[16] reported that infections as well as malnutrition can cause thymus atrophy leading

Table 1. Cell-mediated immune response to PHA-P, and antibody titer response against NDV and SRBC

Tablo 1. PHA-ye karşı hücresel immun yanıt ve NDV ve SRBC'ye karşı antikor yanıtı

Treatment	CBH ¹ Test (mm)		Antibody Titer (log ₂) Against NDV ²		Antibody Titer (log ₂) Against SRBC ³	
	Day 15	Day 30	Day 17	Day 27	Day 17	Day 27
Control	0.643 ^{bc}	0.580 ^{cd}	3.25	4.625 ^{ab}	2	2.5
Control + R ⁴	0.695 ^{abc}	0.528 ^d	2.5	5.250 ^a	2	3
Control + S ⁵	0.783 ^{ab}	0.888 ^a	3.75	5.625 ^a	1.25	3.5
Control + R + S	0.480 ^c	0.503 ^d	3.25	3.250 ^b	2.25	3.375
Prebiotic	0.905 ^a	0.843 ^{ab}	3.25	5.000 ^a	2	2.625
Prebiotic + R	0.693 ^{abc}	0.530 ^d	3.25	4.500 ^{ab}	2.5	3.5
Prebiotic + S	0.840 ^{ab}	0.728 ^{abc}	3	4.750 ^a	2	3.5
Prebiotic + R + S	0.890 ^a	0.663 ^{bcd}	2.75	4.875 ^a	2.5	3.625
SEM	0.052	0.043	0.412	0.458	0.330	0.592
P-value						
Prebiotic	0.002	0.135	0.672	0.775	0.122	0.607
Restriction	0.061	0.001	0.212	0.116	0.043	0.421
<i>Salmonella</i>	0.786	0.093	0.672	0.506	0.597	0.171
P×R×S ⁶	0.007	0.003	0.672	0.011	0.296	0.941

1. Cutaneous basophil hypersensitivity reaction to phytohemagglutinin P injection into the toe web skin, and measured as skin swelling before and after injection, 2. Antibody response against Newcastle disease virüs, 3. Antibody response against Sheep Red Blood Cell, 4. Restriction in feed, 5. *Salmonella* challenge, 6. Interaction among prebiotic, feed restriction and *Salmonella* challenge, ^{a,b,c,d} Within the same column, means with different superscripts are significantly differ ($P < 0.05$)

Table 2. The relative weights (% of live BW) of lymphoid organs at days 21 and 42
Tablo 2. 21. ve 42. günlerde lenfoid organların göreceli (% canlı ağırlığa göre) ağırlığı

Treatment	Day 21			Day 42		
	Bursa (%)	Thymus (%)	Spleen (%)	Bursa (%)	Thymus (%)	Spleen (%)
Control	0.213	0.373 ^a	0.078	0.220 ^a	0.218	0.125
Control + R ¹	0.250	0.263 ^{ab}	0.075	0.188 ^{ab}	0.193	0.110
Control + S ²	0.268	0.288 ^{ab}	0.075	0.228 ^a	0.275	0.120
Control + R + S	0.208	0.258 ^{ab}	0.085	0.140 ^{bc}	0.140	0.133
Prebiotic	0.243	0.363 ^{ab}	0.080	0.223 ^a	0.165	0.118
Prebiotic + R	0.218	0.323 ^{ab}	0.075	0.180 ^{abc}	0.188	0.110
Prebiotic + S	0.260	0.228 ^b	0.083	0.218 ^a	0.210	0.105
Prebiotic + R + S	0.253	0.310 ^{ab}	0.083	0.125 ^c	0.200	0.105
SEM	0.023	0.042	0.011	0.018	0.043	0.013
P-value						
Prebiotic	0.596	0.726	0.081	0.566	0.610	0.198
Restriction	0.407	0.424	0.935	0.001	0.235	0.793
<i>Salmonella</i>	0.329	0.04	0.570	0.065	0.610	1.00
P×R×S ³	0.91	0.726	0.807	0.923	0.527	0.601

1. Restriction in feed, 2. *Salmonella* challenge, 3. Interaction among prebiotic, feed restriction and *Salmonella* challenge, ^{a,b,c} Within the same column, means with different superscripts are significantly differ (P<0.05)

to decrease in cellular immune response mediated by T-cells. As Hooge ^[17] expressed, MOS advantages are more noticeable in stressor conditions which is in agreement with our findings showing that MOS increased cell-mediated immune response against the stresses caused by feed restriction and *Salmonella* challenge. The mechanism of prebiotic's function in immunity regulation is not yet exactly known. But some studies show that *Salmonella* infections stimulate inflammatory cytokine, interleukin-1 production ^[10,11]. Furthermore, MOS can competitively adsorbs pathogens from mannose and prevent their attack to gut epithelium ^[18]; this can cause increase in energy intake which is helpful in cell-mediated immunity.

Antibody titer responses against NDV had significant differences among treatment groups at day 27. The group under feed restriction and *Salmonella* challenge had the least antibody titer response against NDV and its difference with the group which was supplemented with prebiotic under the same condition, was significant. This indicates the advantageous effect of prebiotic on immunity in agreement with Alavi et al. ^[19], but in contrast with Silva et al. ^[20] findings. Also, Fanoosi and Torki ^[21] did not observe any decrease in antibody titer response against NDV under 10% feed restriction, but Jahanpour et al. ^[22] reported that feed restriction as amount as 75% standard catalogue during days 8 to 14 decreased antibody titer response against NDV. There was no significant difference among treatments regarding antibody titer response against SRBC. Khalaji et al. ^[23] and Hangalapura et al. ^[4] found similar results using prebiotic in broilers and feed restriction in line chickens respectively; but Khajavi et al. ^[24] reported

that antibody titer response increased against SRBC under moderate feed restriction in broilers which is in contrast with Jahanpour et al. ^[22] who reported that feed restriction as amount as 75% standard catalogue during days 8 to 21 decreased antibody titer response against SRBC. Riberio et al. ^[25] expressed that prebiotic effect on IgG against *Salmonella enteritidis* is not significant in broilers; whereas Cetin et al. ^[26] and Woo et al. ^[27] indicated that IgG level increases by MOS supplementation in turkeys and layers respectively. These contradictions might be because of differences in strain and age of the birds, the prebiotic type, feed restriction severity and intensity of exposure to *Salmonella*. However it seems that stresses and nutrient intake decrease have less suppressive effect on humoral immunity than on cell-mediated immunity.

There was a significant difference regarding the relative weight of thymus at day 21 among treatments. The treatment group with prebiotic under *Salmonella* challenge had the least relative weight of thymus. The significant difference between this treatment and the control treatment indicates that despite using prebiotic, *Salmonella* caused a decrease in the weight of thymus. Savino and Dardenne ^[16] expressed that infections cause thymus gland Atrophy. At day 42, the relative weight of bursa of Fabricius had a significant difference among treatments. The treatment group under prebiotic supplementation, restriction and *Salmonella* challenge had the least relative weight of bursa of Fabricius. Feed restriction and *Salmonella* infection stresses might be the main reason of decrease in the relative weight of this lymphoid organ. The plasma corticosterone level can increase because of feed

restriction^[4] and microbial challenge^[28] stresses. The high level of plasma corticosterone can cause a decrease in the immunocompetence and the weight of lymphoid organs.

The results of this study show that using prebiotics during combined stresses can have some advantages in broilers. Although feed restriction has beneficial effects on broilers, its suppressive impact on immunity system especially when used severely must be considered. These suppressive impacts are more noticeable in cell-mediated immunity than humoral immunity, because the cellular immunity components are more sensitive to energy demand. Therefore, prebiotics are helpful in such a condition as well as in *Salmonella* infections.

REFERENCES

- Acar N, Sizemore FG, Leach GR, Wideman RF, Owen JRL, Barbat GF:** Growth of broiler chickens in response to feed restriction regimens to reduce ascites. *Poult Sci*, 74 (5): 833-843, 1995.
- Hinton A, Buhr RJ, Ingram KD:** Physical, chemical and microbiological changes in the crop of broiler chickens subjected to incremental feed withdrawal. *Poult Sci*, 79 (2): 212-218, 2000.
- Savino W:** The thymus gland is a target in malnutrition. *Eur J Clin Nutr*, 56 (3): S46-S49, 2002.
- Hangalapura BN, Nieuwland MGB, Reilingh GDV, Buyse J, Brand HVD, Kemp B, Parmentier HK:** Severe feed restriction enhances innate immunity but suppresses cellular immunity in chicken lines divergently selected for antibody responses. *Poult Sci*, 84 (10): 1520-1529, 2005.
- Kang SY, Ko YH, Moon YS, Sohn SH, Jang IS:** Effects of the combined stress induced by stocking density and feed restriction on hematological and cytokine parameters as stress indicators in laying hens. *Asian-Aust J Anim Sci*, 24 (3): 414-420, 2011.
- van Immerseel F, Cauwerts K, Devriese LA, Haesebrouck F, Ducatelle R:** Feed additives to control *Salmonella* in poultry. *World Poult Sci J*, 58 (4): 501-513, 2002.
- Gibson GR, Roberfroid MB:** Dietary modulation of the human colonic microbiota: Introducing the concept of prebiotics. *J Nutr*, 125 (6): 1401-1412, 1995.
- Kim GB, Seo YM, Kim CH, Paik IK:** Effect of dietary prebiotic supplementation on the performance, intestinal microflora, and immune response of broilers. *Poult Sci*, 90 (1): 75-82, 2011.
- Patterson JA, Burkholder KM:** Application of prebiotics and probiotics in poultry production. *Poult Sci*, 82 (4): 627-631, 2003.
- Maassen CB, van Holten-Neelen C, Balk F, den Bak-Glashouwer MJ, Leer RJ, Laman JD, Boersma WJ, Claassen E:** Strain-dependent induction of cytokine profiles in the gut by orally administered *Lactobacillus* strains. *Vaccine*, 18 (23): 2613-2623, 2000.
- Swaggerty CL, Kaiser P, Rothwell L, Pevzner IY, Kogut MH:** Heterophil cytokine mRNA profiles from genetically distinct lines of chickens with differential heterophil-mediated innate immune responses. *Avian Pathol*, 35 (2): 102-108, 2006.
- Gursoy E, Cardounel A, Hu Y, Kalimi M:** Biological effects of long-term caloric restriction: Adaptation with simultaneous administration of caloric stress plus repeated immobilization stress in rats. *Exp Biol Med*, 226 (2): 97-102, 2001.
- Corrier DE, DeLoach JR:** Evaluation of cell-mediated, cutaneous basophil hypersensitivity in young chickens by an interdigital skin test. *Poult Sci*, 69 (3): 403-408, 1990.
- Marquardt WW, Synder DB, Savage PK, Kadavil SK, Yancey FS:** Antibody response to Newcastle disease virus given by two different routes as measured by ELISA and hemagglutination-inhibition test and associated tracheal immunity. *Avian Dis*, 29 (1): 71-79, 1985.
- Wegmann T, Smithies O:** A simple hemagglutination system requiring small amounts of red cells and antibodies. *Transfusion*, 6 (1): 67-75, 1966.
- Savino W, Dardenne M:** Nutritional imbalances and infections affect the thymus: consequences on T-cell-mediated immune responses. *Proc Nutr Society*, 69 (4): 636-643, 2010.
- Hooze DM:** Meta-analysis of broiler chicken pen trials evaluating dietary mannanoligosaccharide, 1993-2003. *Int J Poult Sci*, 3 (3): 163-174, 2004.
- Fernandez F, Hinton M, van Gils B:** Dietary mannan-oligosaccharides and their effect on chicken caecal microflora in relation to *Salmonella enteritidis* colonization. *Avian Pathol*, 31 (1): 49-58, 2002.
- Alavi SAN, Zakeri A, Kamrani B, Pourakbari Y:** Effect of prebiotics, probiotics, acidifier, growth promoter antibiotics and synbiotic on humoral immunity of broiler chickens. *Global Veterinaria*, 8 (6): 612-617, 2012.
- Silva VK, da Silva JDT, Torres KAA, de Faria Filho DE, Hada FH, de Moraes VMB:** Humoral immune response of broilers fed diets containing yeast extract and prebiotics in the prestarter phase and raised at different temperatures. *J Appl Poult Res*, 18 (3): 530-540, 2009.
- Fanooci M, Torki M:** Effects of qualitative dietary restriction on performance, carcass characteristics, white blood cell count and humoral immune response of broiler chicks. *Global Veterinaria*, 4 (3): 277-282, 2010.
- Jahanpour H, Seidavi A, Qotbi AAA:** Effects of intensity and duration of feed quantitative restriction on broiler immune system. *Annals Bio Res*, 3 (6): 2656-2661, 2012.
- Khalaji S, Zaghari M, Nezafati S:** The effects of mannan-oligosaccharides on cecal microbial populations, blood parameters, immune response and performance of broiler chicks under controlled condition. *Afr J Biochem Res*, 5 (5): 160-164, 2011.
- Khajavi MS, Rahimi S, Hassan ZM, Kamali MA, Mousavi T:** Effect of feed restriction early in life on humoral and cellular immunity of two commercial broiler strains under heat stress conditions. *Br Poult Sci*, 44 (3): 490-497, 2003.
- Ribeiro AML, Vogt LK, Canal CW, Cardoso MR, Labres RV, Streck AF, Bessa MC:** Effects of prebiotics and probiotics on the colonization and immune response of broiler chickens challenged with *Salmonella enteritidis*. *Brazil J Poult Sci*, 9 (3): 193-200, 2007.
- Cetin N, Guclu BK, Cetin E:** The effect of prebiotics and mannan-oligosaccharide on some haematological and immunological parameters in turkey. *J Vet Med A*, 52 (3): 263-267, 2005.
- Woo KC, Kim CH, Paik IK:** Effects of supplementary immune modulators (MOS, Lectin) and organic acid mixture (organic acid F, organic acid G) on the performance, profile of leukocytes and erythrocytes, small intestinal microflora and immune response in laying hens. *Kor J Anim Sci Technol*, 49 (4): 481-490, 2007.
- Khansari DN, Murgu AJ, Faith RE:** Effects of stress on the immune system. *Immunol Today*, 11 (5): 170-175, 1990.

Effects of Low Doses of Bisphenol A on Primordial Germ Cells in Zebrafish (*Danio rerio*) Embryos and Larvae ^{[1][2]}

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Summary

Primordial germ cells are the precursors of gametes in sexually reproducing organisms. The migration and homing process of primordial germ cells are prone to environmental effects and the endocrine system hormones. One of the environmental pollutants which humans are exposed to is bisphenol A (BPA). BPA which is an endocrine disrupter generally used for making plastics harder. In this study we investigated the effects of low doses of BPA (4 mg/L and 8 mg/L) on primordial germ cells at the zebrafish embryos and larvae. Whole mount in situ hybridization for germ cell marker gene vasa showed that upon exposure to BPA, the primordial germ cell number increased and these cells ectopically localized outside the normal germ cell niche in zebrafish. Migration of primordial germ cells into ectopic locations and morphological changes in these cells were also proved by histological studies. Additionally, results of acridine orange staining to detect apoptotic cells showed that low doses of BPA did not increase apoptosis. Results of the present study showed that low doses of BPA exposure increased the number of primordial germ cells and induced ectopic primordial germ cell localization in the zebrafish embryos.

Keywords: Zebrafish, Primordial germ cell, Bisphenol A

Düşük Dozlardaki Bisfenol A'nın Zebra Balığı (*Danio rerio*) Embriyo ve Larvalarında Primordiyal Germ Hücreleri Üzerine Etkisi

Özet

Primordiyal germ hücreleri, eşeyli üreyen canlılarda gametlerin öncül hücreleridir. Endokrin sistem hormonları ve çevresel etkenler, primordiyal germ hücrelerinin göçünü etkilemektedir. İnsanların maruz kaldığı çevresel kirleticilerden biri de bisfenol A (BPA)'dır. Endokrin sistemde hasar oluşturan biri olan BPA, genellikle plastik sertleştirme işleminde kullanılmaktadır. Bu çalışmada zebra balığı embriyo ve larvalarında düşük dozlardaki (4 mg/L and 8 mg/L) BPA'nın etkileri incelenmiştir. Bir germ hücresi belirteci gen olan vasa ile yapılan whole mount in situ hibridizasyon deneyleri sonucunda BPA'nın primordiyal germ hücre sayısında artışa sebep olduğu ve bu hücrelerin göç yolu dışındaki (ektopik) bölgelerde bulunduğu tespit edilmiştir. Primordiyal germ hücrelerinin ektopik bölgelere göçü ve bu hücrelerdeki morfolojik değişiklikler histolojik çalışmalarla da doğrulanmıştır. Buna ek olarak, düşük dozlarıyla yapılan boyamalarda apoptozisin tespitinde kullanılan akrinin turuncusu boyaması sonucunda BPA'nın apoptozisi artırmadığı saptanmıştır. Bu çalışma sonuçları, düşük dozlardaki BPA'nın primordiyal germ hücre sayısını artırdığını ve bu hücrelerin ektopik bölgelere göç etmesine yol açtığını ortaya koymaktadır.


Anahtar sözcükler: Zebra Balığı, Primordiyal germ hücresi, Bisfenol A

INTRODUCTION

Primordial germ cells (PGCs) constitute an embryonic cell type that migrate to gonadal precursors and form the gametes. Thus PGCs are the cells that provide the

permanence of the species and are crucial for the development of a new organism in the next generation ^[1]. In many organisms PGCs are specified during early

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embryonic development and they contain germ granules [2-7].

PGC migration must be finely regulated as it follows a complex pathway through various developing tissues [8]. The ultimate location of PGCs in zebrafish is between the endodermal cells the yolk sac which are close to allantois, at early stages of the embryonic development [9]. They migrate into dorsal mesentery and reaches in the gonad primordia with amoeboid movements. The pathway which PGCs follows defined as "gonadal pathway" [10-13]. PGC migration in zebrafish takes place during the first 24 h of the embryonic development [14-16].

In zebrafish, germ line lineage can be traced throughout embryogenesis by means of transcripts of *vasa* [17] which is a gene that is first identified in *Drosophila melanogaster* as a maternal determinant of germ cell formation [18]. In zebrafish, *vasa* is a molecular marker for PGC [19,20]. *Vasa* protein can be detected in the germline cells in early embryos, because that it specifically located into polar granules, which are located where the germ cells are specified. PGCs are specified at four positions along the margin of the blastodisc and start migrating dorsally during gastrulation [16]. PGCs move towards the intermediate targets around somites 1-3 at 10.5 h post fertilization (hpf), and subsequently to the final target region at the level of somites 8-10 at 13 hpf. At 24 hpf, PGCs localize to the junction between the yolk ball and yolk extension in the gonadal region, forming compact clusters [16].

Several chemicals or environmental pollutants can cause cellular damage in PGCs which has ramifications in different diseases/disorders in organisms [21,22]. Therefore, investigating the migratory behaviour and regulations of PGCs is an intriguing biological question. Additionally, human are exposed to many of the environmental toxicants everyday. Information obtained from experiments on the reproductive systems of the experimental animals can provide important findings about the impact of humans.

Endocrine disrupting chemicals (EDCs) are compounds in the external environment that modulate the physiology of the endocrine system and often cause health disorders [23]. Bisphenol A (2 bis(4-hydroxyphenyl) propane, BPA) is one of the most potent endocrine disrupting chemicals which mimics estrogen *in vivo* and *in vitro* [24] and therefore is classified as xenoestrogen. BPA is widely used in industry for making plastics harder. It's found in polycarbonate plastics (baby feeding bottles, carboys) [25] and inside of epoxy coated cans [26]. With the increasing usage of these materials in daily life, humans are exposed to BPA in an escalating manner, which could cause endocrine disorders. High concentrations of estrogens stimulate PGC growth *in vitro* through the somatic cells of the gonadal ridges [27]. BPA acts by several different mechanisms to perturb the early stages of oogenesis and also affects primordial germ cells, by influencing mitotic proliferation and the timing of meiotic entry [28].

Duan and Zhu [29] investigated the toxicity of BPA on the growth of zebrafish embryos and found the median embryo lethal concentration (LC50) for 24 h as 16.36 ± 0.40 mg/L. They also mentioned that all embryos died at 25 mg/L dose at 24 hpf. So in the present study, doses were chosen according to LC50 value of bisphenol A on zebrafish embryos. We carried out zebrafish embryos 4 mg/L and 8 mg/L BPA and investigated the effects on primordial germ cells of the zebrafish embryos. In 24hpf zebrafish embryos we determined the location of PGCs with whole mount *in situ* experiments. With histological analysis we observed the migration pathway of PGCs for two weeks.

MATERIAL and METHODS

This study was approved by Kocaeli University Animal Researches Ethics Committee (HADYEK- 5/3 - 2011).

Material

All animal experiments and procedures were carried out in accordance with the recommendations and official guidelines for animal handling and research. The temperature and humidity were kept at 28.5°C and 61%, respectively. Sexually matured zebrafish were fed daily with *Artemia* sp. and Tetra- Min® Hauptfutter (Tetra Werke, Germany) under standardized conditions (20-L glass aquaria, $28 \pm 1^\circ\text{C}$, light/dark cycle = 14 h/10 h) and after egg laying embryos were collected immediately. Under a stereo microscope, fertilized and unfertilized eggs were separated. Fertilized embryos were transferred into special small embryo aquariums.

Methods

BPA Exposure

BPA, 2 bis(4-hydroxyphenyl), (Sigma Aldrich, CAS No: 50-05-7) was dissolved in 1% dimethylsulfoxide (DMSO) to generate desired concentrations for treatment of the embryos. Two control and 2 experimental groups were used as: control (untreated), solvent control group (1% DMSO), 4 mg/L BPA and 8 mg/L BPA. DMSO and BPA were applied from 1 h post fertilization until sacrificing the embryos.

Whole Mount *In Situ* Hybridization

24-hpf-old embryos from control and BPA treatment groups were collected and fixed in 4% paraformaldehyde (PFA, Sigma) overnight at 4°C. After removing chorion, the embryos were washed in methanol series and stored in 100% MeOH at -20°C . Digoxigenin-labelled antisense RNA probes were generated for *vasa* gene as described Miyake et al. [30]. Whole mount *in situ* hybridizations with chromogenic substrates Nitro Blue Tetrazolium (NBT)/5-Bromo-4-Chloro-3-Indoyl phosphate (BCIP) were on the 24-hpf-embryos performed as previously described [31,32].

Acridine Orange Staining

After removing the chorion of the zebrafish embryos 5 µg/L acridine orange (Sigma) was added into the embryo rearing medium. Embryos were incubated in acridine orange solution for 30 min. They were anaesthetised with 0.04% of 3-aminobenzoic acid methyl ester (MESAB) and imaged under dissecting fluorescent microscope.

Histology

Embryos and larvae (0-2 weeks old) were fixed in Bouin's fixative for 8 h. After fixation, tissue was dehydrated in ascending concentrations of ethanol, equilibrated in xylene. The tissues were then embedded in paraffin wax and cut into 5-7 µm sections on a microtome. The sections were mounted on glass slides and stained with haematoxylin and eosin, toluidine blue and Best's carmine before examination under a light microscope.

Statistical Analysis

Statistical analysis were performed by using IBM SPSS Statistics 5.0 programme. Quantitative data was given as mean ± SD. T test were used for analyses and $P < 0.05$ was considered as statistical significance value. For statistical analysis, 7 samples were used in each group ($n=7$).

RESULTS

Results of Expression of vasa Gene in Primordial Germ Cells

Vasa gene is a germ cell marker [19,20]. At embryonic stages of development, vasa transcripts are concentrated in an electron-dense structure, the putative zebrafish germ plasm [20,33].

The location and number of PGCs were detected at 24 h post fertilization (Fig. 1 and Fig. 2). In each group, 7 embryos were tested. PGCs that migrate the correct and ectopic locations were counted. In control (untreated) and DMSO groups PGCs migrate to the target region correctly (Fig. 1-a and 1-b). When compared the 4 mg/L BPA and 8 mg/L BPA groups with control ones, there was an increase in the number of PGCs (Fig. 2) and a significant portion of germ cells were at ectopic locations (Fig. 1-c and 1-d). It was observed that 1% DMSO and BPA treatment increased the PGCs numbers in comparison to the untreated group. t test results showed that DMSO and BPA treatment increased the total PGC number (Table 1). The number of PGCs increased in BPA treated group (Table 2). BPA treatment also increased ectopic PGCs (Table 3). Our data suggest that BPA caused an increase in the number

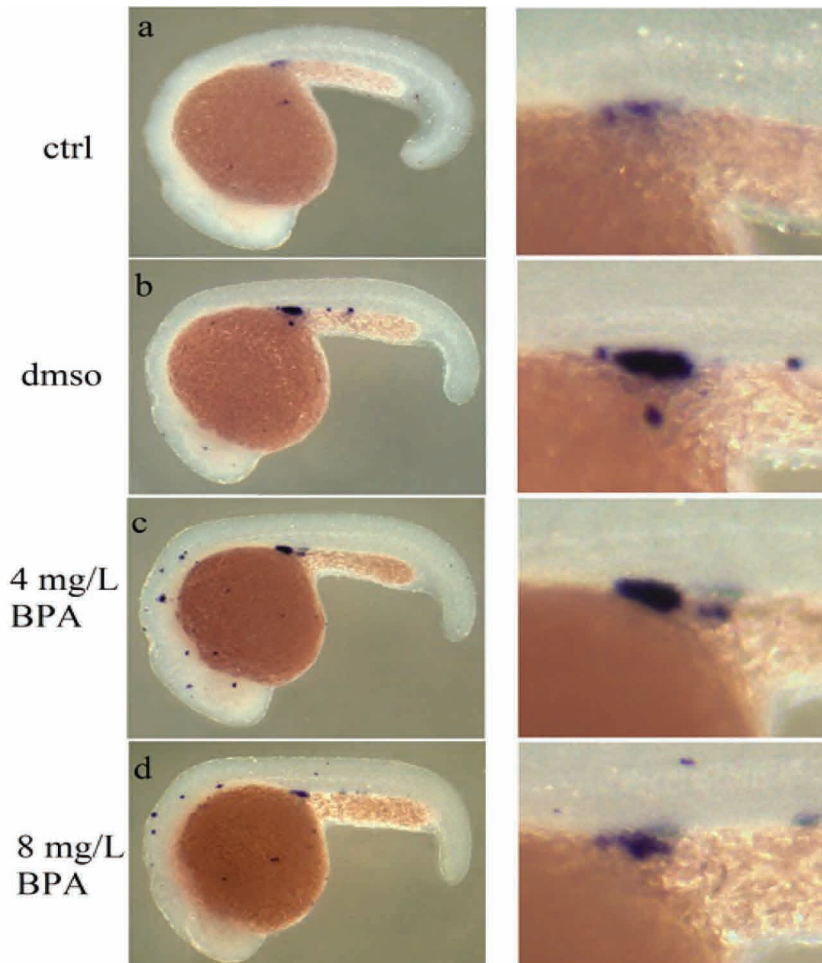


Fig 1. Whole mount in situ hybridization results of 24 hpf old zebrafish embryos. Magnifications at the right side show PGCs which locate the right direction. **a)** Untreated (ctrl) group, PGCs located to the right direction, **b)** 1% DMSO (solvent control) group, PGCs located to the right direction but there are few ectopic cells, **c)** 4 mg/L BPA treated group, most of PGCs are ectopic, **d)** 8 mg/L BPA treated group, most of PGCs are ectopic

Şekil 1. 24 saatlik zebra balığı embriyolarında whole mount in situ hibridizasyon sonuçları. Sağ taraftaki büyütülmüş kısımlar doğru bölgede bulunan primordiyal germ hücrelerini (PGH) göstermektedir. **a)** Kontrol grubu, PGH doğru bölgede bulunmaktadır, **b)** Çözücü kontrol (%1 DMSO) grubu, PGH doğru bölgede bulunmaktadır fakat birkaç hücre ectopik bölgede bulunmaktadır, **c)** 4 mg/L BPA uygulanmış grup, pek çok PGH ectopik bölgelerdedir, **d)** 8 mg/L BPA uygulaması yapılmış grup, pek çok PGH ectopik bölgededir

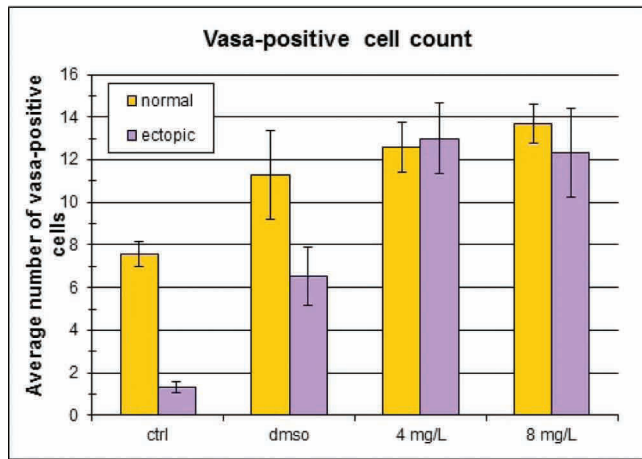


Fig 2. General comparement of control and BPA treatment groups. When compare with control group, as a result of BPA treatment more vasa-positive cells (PGCs) were detected at ectopic and right locations (n=7)

Şekil 2. Kontrol ve BPA gruplarının genel karşılaştırması. Kontrol grubu ile karşılaştırıldığında BPA uygulaması sonucunda vasa pozitif hücreler (PGH) doğru ve ektopik bölgelerde tespit edilmişlerdir (n=7)

Table 1. Total (ectopic + gonadal region) PGC frequency at 24 hpf zebrafish embryos

Tablo 1. 24 saatlik zebra balığı embriyolarındaki toplam (ektopik + gonadal bölgedeki) PGH sıklığı

Group	Total PGCs Frequency			*
	Average	Standart Deviation		
Control (untreated) (n=7)	4.43	± 0.94		a
1% DMSO (n=7)	9.00	± 1.45		b
4 mg/L BPA (n=7)	13.50	± 0.81		c
8 mg/L BPA (n=7)	13.71	± 0.98		c

* There are significant difference between samples that are shown with different letters (P<0.05)

Table 2. PGC frequency at gonadal region at 24 hpf zebrafish embryos

Tablo 2. 24 saatlik zebra balığı embriyolarındaki gonadal bölgedeki PGH sıklığı

Group	PGCs Frequency at Gonadal Location			*
	Average	Standart Deviation		
Control (untreated) (n=7)	7.57	± 0.65		a
1% DMSO (n=7)	11.29	± 2.22		a,b
4 mg/L BPA (n=7)	12.57	± 1.25		b
8 mg/L BPA (n=7)	13.71	± 0.99		b

* There are significant difference between samples that are shown with different letters (P<0.05)

of PGCs, although all of these cells cannot migrate into the prospective primordia and remained at ectopic locations.

Results of Acridine Orange Staining

In the second part of the study we investigated the effects of BPA on the apoptosis frequency in zebrafish

Table 3. PGC frequency at ectopic locations at 24 hpf zebrafish embryos

Tablo 3. 24 saatlik zebra balığı embriyolarındaki ektopik bölgelerdeki PGH sıklığı

Group	PGCs Frequency at Ectopic Locations			*
	Average	Standart Deviation		
Control (untreated) (n=7)	1.29	± 0.29		a
1% DMSO (n=7)	6.71	± 1.57		b
4 mg/L BPA (n=7)	14.43	± 1.00		c
8 mg/L BPA (n=7)	13.71	± 1.77		c

* There are significant difference between samples that are shown with different letters (P<0.05)

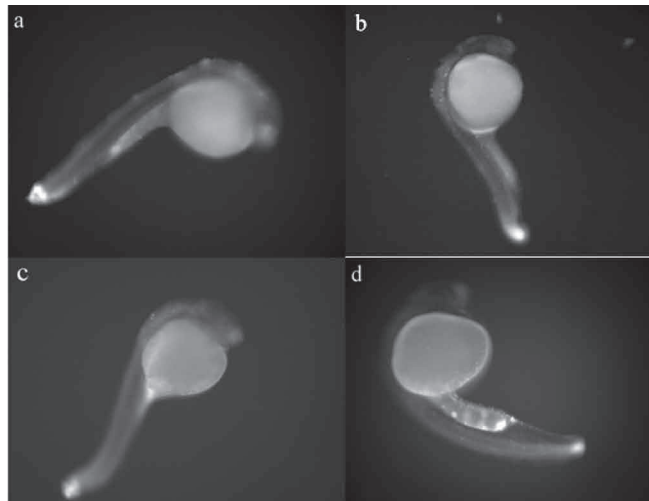


Fig 3. Acridine orange staining results a) Control group b) Solvent control group (1% DMSO) c) 4 mg/L BPA treated group d) 8 mg/L BPA treated group. When compare BPA treatment groups with control ones, it was not seen any differences between the groups

Şekil 3. Akridin turuncusu boyama sonuçları a) Kontrol grubu b) Çözücü kontrol grubu (%1 DMSO) c) 4 mg/L BPA uygulaması yapılmış grup d) 8 mg/L BPA uygulaması yapılmış grup. Kontrol grupları BPA uygulaması yapılmış gruplarla karşılaştırıldığında gruplar arasında bir fark görülmemiştir

embryos. Acridine orange is a nucleic acid fluorochrome which emits fluorescence when it intercalates into the DNA [34]. BPA treatment did not change apoptosis frequency and the frequency was in normal range that normally prevails during the embryonic development (Fig. 3) of zebrafish embryos.

Histological Studies

Morphology of primordial germ cells were detected with histological methods during 2 weeks. The embryos of the 8 mg/L BPA group could survived 7 days maximally. PGCs which are bigger than somatic cells and have large nucleus, nuage material and glycogen granules were detected by H&E, toluidine blue and Best's carmine stains. It was found that results of histological studies were supported by whole mount *in situ* results. In DMSO and BPA treated groups, increase in the number of PGCs which located at ectopic regions were detected.

At 3 day post fertilization (dpf) embryos, PGCs were observed at the dorsal part of yolk sac in untreated group. PGC structure was normal (Fig. 4-a). Location of PGCs were shown with arrows (Fig. 4-b,c,d). In BPA treated groups, decrease at glycogen granules in PGCs were observed (Fig. 4-c,d).

At 7 dpf prelarvae, PGCs were observed near muscle tissue (Fig. 5-a,b,c,d). In untreated and DMSO group, the shapes of migrating PGCs were ameboid (shown with star) (Fig. 5-a,b). In BPA treated groups, most of PGCs did not have ameboid shape (Fig. 5-c,d).

At 14 dpf larvae, in untreated and DMSO groups, PGCs were observed at gonadal region (Fig. 6-a,b). PGCs could not migrate gonadal region at 4 mg/L BPA treated group.

In this group, PGCs were observed near somites (Fig. 6-c). Decrease at nuage material of PGCs were also detected in this group (Fig. 6-c,d).

DISCUSSION

We conducted this study to evaluate the effects of BPA on primordial germ cells of zebrafish (*Danio rerio*) embryos because very little is known about the effects of BPA on primordial germ cells in vertebrates. There are not any study the effects of BPA on primordial germ cells. so, our study is pioneer about this subject.

Avcı et al.^[35] determined the effects of nonylphenol on growth parameters and antioxidant defense system

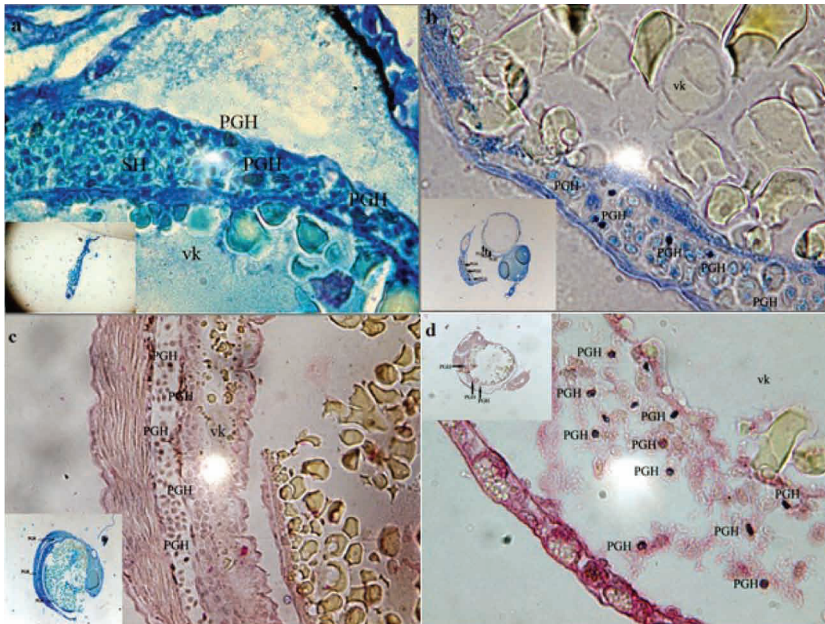
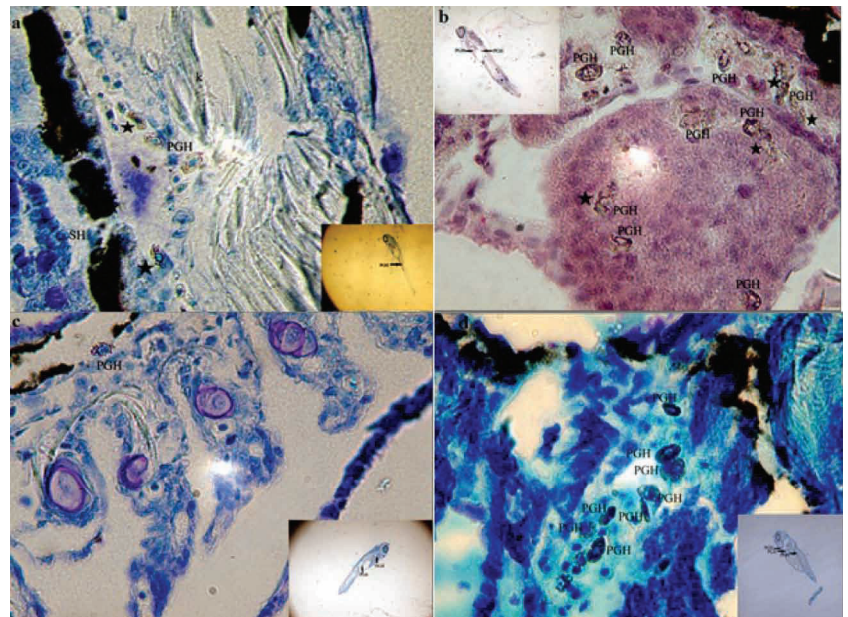


Fig 4. Histologic sections of 3 dpf embryos. Location of PGCs were shown with arrows. PGH: primordial germ cell, SH: somatic cell, vk: yolk sac, a) Untreated group, Toluidine blue staining (x4), (x100), b) Solvent control group (1% DMSO), Toluidine blue staining (x10), (x100), c) 4 mg/L BPA treated group, Toluidine blue and Best carmine staining (x10), (x100), d) 8 mg/L BPA treated group, Best carmine staining (x10), (x100)

Şekil 4. 3 günlük embriyoların histolojik kesiti. PGH bulunduğu yerler oklar ile gösterilmiştir. PGH: Primordiyal germ hücresi, SH: somatik hücre, vk: vitellüs kesesi, a) Kontrol grubu, Toluidine mavisi boyaması (x4), (x100), b) Çözücü kontrol grubu (%1 DMSO), Toluidine mavisi boyaması (x10), (x100), c) 4 mg/L BPA uygulaması yapılmış grup, Toluidine mavisi ve Best Carmine boyaması (x10), (x100), d) 8 mg/L BPA uygulaması yapılmış grup, Best carmine boyaması (x10), (x100)

Fig 5. Histologic sections of 7 dpf prelarvae. Location of PGCs were shown with arrows. Migrating PGCs were mentioned with star. PGH: primordial germ cell, SH: somatic cell, k: muscle tissue, a) Untreated group, Toluidine blue staining (x10), (x100), b) Solvent control group (1% DMSO), Best carmine staining (x10), (x100), c) 4 mg/L BPA treated group, PGC at ectopic region, Toluidine blue staining (x10), (x100), d) 8 mg/L BPA treated group, Toluidine blue staining (x10), (x100)

Şekil 5. 7 günlük prelarvaların histolojik kesiti. PGH bulunduğu yerler oklar ile gösterilmiştir. Göç eden PGH yıldız ile gösterilmiştir. PGH: Primordiyal germ hücresi, SH: somatik hücre, k: kas dokusu. a) Kontrol grubu, Toluidine mavisi boyaması (x10), (x100), b) Çözücü kontrol grubu (%1 DMSO), Best carmine boyaması (x10), (x100), c) 4 mg/L BPA uygulaması yapılmış grup, ektopik bölgedeki PGH, Toluidine mavisi boyaması (x10), (x100) d) 8 mg/L BPA uygulaması yapılmış grup, Toluidine mavisi boyaması (x10), (x100)



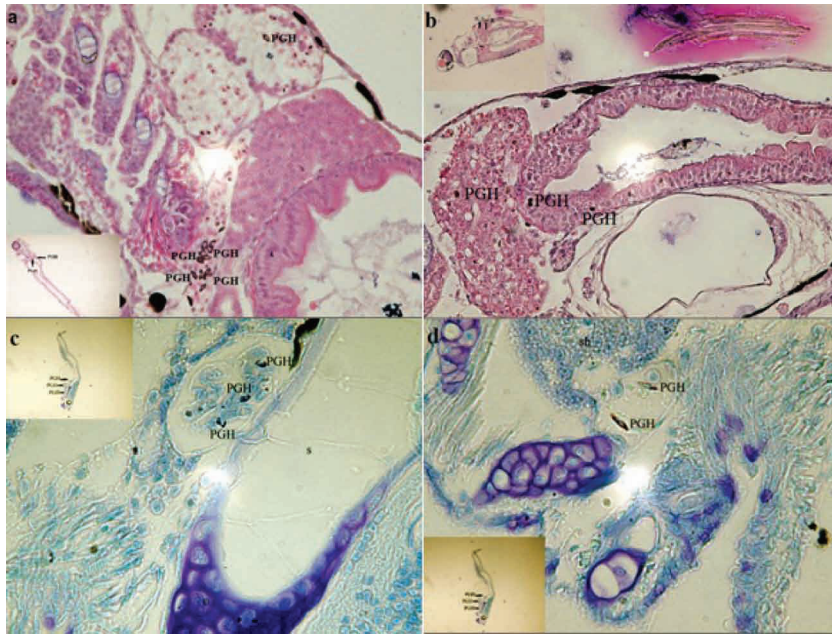


Fig 6. Histologic sections of 14 dpf larvae. Location of PGCs were shown with arrows. PGH: primordial germ cell, sh: somatic cell, s: somites, a) Untreated group H&E staining (x10), (x40), b) Solvent control group (1% DMSO), H&E staining (x20), (x40), c) 4 mg/L BPA treated group, Toluidine blue staining (x10), (x100), d) 4 mg/L BPA treated group, PGC at ectopic region, Toluidine blue staining (x10), (x100)

Şekil 6. 14 günlük larvaların histolojik kesiti. PGH bulunduğu yerler oklar ile gösterilmiştir. PGH: Primordiyal germ hücresi, sh: somatik hücre, s: somitler, a) Kontrol grubu, H&E boyaması (x10), (x100), b) Çözücü kontrol grubu (%1 DMSO), H&E boyaması (x20), (x40), c) 4 mg/L BPA uygulaması yapılmış grup, Toluidine mavisi boyaması (x10), (x100), d) 4 mg/L BPA uygulaması yapılmış grup, ektopik bölgedeki PGH, Toluidine mavisi boyaması (x10), (x100)

Japanese quails (*Coturnix japonica*) and they found that nonylphenol has dramatic effects on egg production rate and anti-oxidant system in Japanese quails (*Coturnix japonica*). In addition, Willey and Krone^[21] investigated effects of endosulfan and nonylphenol on the primordial germ cell population in pre-larval zebrafish embryos during the first day of development. In this study, vasa antisense RNA probe was used for whole mount in situ hybridization. As a result of the study, they proved that endosulfan treatment cause decrease the number of PGCs in 5th and 6th somites, increase the number of PGCS in 7th and 8th somites. Between 9th and 13th somites any change in the number of PGCs didn't observed. In nonylphenol treated group they found decrease in the number of PGCs between 6th and 7th somites. As a result, these chemicals cause alternations in the distrubition of PGCs along the posterior and anterior axis. In our study, we proved that BPA treatment cause ectopic localization of PGCs. Besides, it cause increase in the number of total PGCs.

Lawson et al.^[28] studied the effects of low doses of BPA on gene expression in the fetal mouse ovary and they suggested that BPA acts to down-regulate mitotic cell-cycle genes, raising the possibility that fetal BPA exposure may act to limit expansion of the primordial germ cell population. On contrary, we found an increase in the number of PGC population after BPA exposure in zebrafish embryos.

Doitsidou et al.^[36] found that SDF-1a chemokine is the guidance for primordial germ cell attraction. In our study, it's obvious that BPA treatment cause an increment at the number of ectopic primordial germ cells. This result can be interpreted with 2 ways. First, BPA can inhibit SDF-1a chemokine signal mechanism. Second, BPA treatment increses primordial germ cell production and the ectopic

cells that can be seen from figures are migrating cells to the targets.

In some references it's mentioned that BPA induces apoptosis at rodents. Li et al.^[6] gave high doses of BPA to mice and they used TUNNEL assay, immunohistochemistry and western blotting techniques. As a result they found that high doses of BPA exposure induces apoptosis and upregulation of fas/fasI and caspase-3 expression in testes of mice. Similarly, Wang et al.^[37] found that BPA induced germ cell apoptosis in testes. These studies were done at tissues of adult individuals. On contrary in our study we studied with embryos and we could not see any apoptotic effects of low doses of BPA on zebrafish embryos. Similarly to our results, Oka et al.^[38] studied the effects of BPA on apoptosis in central neural cells during early development of *Xenopus* and they also couldn't observe apoptotic effects in early stages.

REFERENCES

1. Raz, E: Primordial Germ Cell Development in Zebrafish. *Cell Dev Biol*, 13, 489-495, 2002.
2. Eddy EM: Germ plasm and the differentiation of germ cell line. *Int Rev Cytol*, 43, 229-280, 1975.
3. Saffman E, Lasko P: Germline Development in vertebrates and invertebrates. *Cell Mol Life Sci*, 55, 1141-1163, 1999.
4. Schisa JA, Pitt JN, Priess JR: Analysis of RNA associated with P granules in germ cells of *C. elegans* adults. *Dev*, 128, 1287-1298, 2001.
5. Seydoux G, Strome S: Launcing the germ line in *Caenorhabditis elegans*: Regulation of gene expression in early germ cells. *Dev*, 128, 3275-3283, 1999.
6. Li W, Deng F, Wang H, Zhen Y, Xiang F, Sui Y, Li J: Germ cell-less expression in zebrafish embryos. *Dev Growth Differ*, 48 (5): 333-338, 2006.
7. Ozhan-Kizil G, Havemann J, Gerberding M: Germ cells in the crustacean *Parhyale hawaiiensis* depend on vasa protein for their maintenance but not for their formation. *Dev Biol*, 327 (1): 230-239, 2009.

8. Richardson BE, Lehmann R: Mechanisms guiding primordial germ cell migration: strategies from different organisms. *Nat Rev Mol Cell Biol*, 11, 37-49, 2010.
9. Felici DM: Twenty years of research on primordial germ cells. *Int J Dev Biol*, 45, 519-522, 2001.
10. Coucouvanis EC: Molecular and Cellular Aspects of Prenatal Gametogenesis in the Mouse. *MSc Thesis*. Stanford University. 1993.
11. Carlson BM: Patten's Foundations of Embryology. Department of Anatomy and Cell Biology. University of Michigan. USA. 1996.
12. Baser H, Eisenbeiss S, Neumann M, Reichmann FM, Thiss B, Thiss C, Raz E: Transition from non-motile behaviour to directed migration during early PGC development in zebrafish. *J Cell Sci*, 118, 4027-4038, 2005.
13. Koc ND: The effects of deltamethrin manipulation on during PGCs' migration pathway of zebrafish (*Branchydanio rerio*). *PhD Thesis*, Marmara University, TURKEY, 2008.
14. Weidinger G, Wolke U, Kopranner M, Klinger M, Raz E: Identification of tissues and patterning events required for distinct steps in early migration of zebrafish primordial germ cells. *Dev*, 126, 5295-5307, 1999.
15. Weidinger G, Wolke U, Kopranner M, Thisse C, Thisse B, Raz E: Regulation of zebrafish primordial germ cell migration by attraction towards an intermediate target. *Dev*, 129, 25-36, 2002.
16. Saito T, Kazeto RG, Kawakami Y, Nomura K, Tanaka H, Adachi S, Arai K, Yamaha E: The mechanism for primordial germ-cell migration is conserved between Japanese eel and zebrafish. *PLoS One*, 6 (9): 1-8, 2011.
17. Nagai T, Yamaha E, Arai K: Histological differentiation of primordial germ cells in zebrafish. *Zool Sci*, 18, 215-223, 2001.
18. Raz, E: The function and regulation of vasa-like genes in germ-cell development. *Gen Biol*, 1 (3): 1017.1-1017.6, 2000.
19. Olsen LC, Aasland R, Fjose A: A vasa-like gene in zebrafish identifies putative primordial germ cells. *Mech Dev*, 66, 95-105, 1997.
20. Yoon C, Kawakami K, Hopkins N: Zebrafish vasa homologue RNA is localized to the cleavage planes of 2-and 4-cell-stage embryos and is expressed in the primordial germ cells. *Dev*, 124, 3157-3165, 1997.
21. Willey JB, Krone PH: Effects of endosulfan and nonylphenol on the primordial germ cell population in pre-larval zebrafish embryos. *Aquat Toxicol*, 54 (1-2): 113-123, 2001.
22. Katayama K, Ueno M, Yamauchi H, Nakayama H, Doi K: Erthylnitrosourea induced apoptosis in primordial germ cells of the rat fetus. *Exp and Toxicol Pathol*, 54 (3): 193-196, 2002.
23. Crews D, McLachlan JA: Epigenetics, evolution, endocrine disruption, health, and disease. *Endocrinol*, 147 (6): 4-10, 2006.
24. Kim YJ, Yun HJ, Ryu JC: Expression profiling of estrogen responsive genes on bisphenol A, 4-nonylphenol and 17 β -estradiol treatment using in house cDNA microarray. *Bio Chip J*, 5 (1): 86-94, 2011.
25. Cao XL, Corriveau J: Migration of bisphenol a from polycarbonate baby and water bottles into water under severe conditions. *J Agric Food Chem*, 56, 6378-6381, 2008.
26. Kang HJ, Kondo F, Katayama Y: Human exposure to bisphenol A. *Toxicol*, 226 (2-3): 79-89, 2006.
27. Behrens GHGM, Klinger FG, Eskild W, Grotmol T, Haugen TB, Felici M: Akt/PTEN signaling mediates estrogen-dependent proliferation of primordial germ cells *in vitro*. *Mol Endocrinol*, 17 (12): 2630-2638, 2003.
28. Lawson C, Gieske M, Murdoch B, Ye P, Li Y, Hassold T, Hunt PA: Gene expression in the fetal mouse ovary is altered by exposure to low doses of bisphenol A. *Biol Reprod*, 84, 79-86, 2011.
29. Duan ZH, Zhu L: Toxicity of bisphenol a on the growth of zebrafish embryos. *Acta Hydrobiol Sin*, 30 (6): 638-642, 2006.
30. Miyake A, Saito T, Kashiwagi T, Ando D, Yamamoto A, Suzuki T, Nakatsuji N, Nakatsuji T: Cloning and pattern of expression of the shirou-uo vasa gene during embryogenesis and its roles in PGC development. *Int J Dev Biol*, 50, 619-625, 2006.
31. Reifers F, Adams J, Mason IJ, Schulte-Merker S, Brand, M: Overlapping and distinct functions provided by fgf17, a new zebrafish member of the Fgf8/17/18 subgroup of Fgfs. *Mech Dev*, 99 (1-2): 39-49, 2000.
32. Kizil C, Otto GW, Geisler R, Nusslein-Volhard C, Antos CL: Simplet controls cell proliferation and gene transcription during zebrafish caudal fin regeneration. *Dev Biol*, 325 (2): 329-340, 2009.
33. Knaut H, Pelegri F, Bohmann K, Schwarz H, Nusslein-Volhard C: Zebrafish vasa RNA but not its protein is a component of the germ plasm and segregates asymmetrically before germline specification. *J Cell Biol*, 149, 875-888, 2000.
34. Nafisi S, Saboury AA, Keramat N, Neault JF, Tajmir-Riahi HA: Stability and structural features of DNA intercalation with ethidium bromide, acridine orange and methylene blue. *J Mol Struct*, 827 (1-3): 35-43, 2007.
35. Avci G, Uguz C, Bayram I, Erdogan M, Kucukkurt I, Dosay Akbulut M, Özdemir M, Lenger OF, Iscan M, Togan I: Effects of nonylphenol on growth parameters and antioxidant defense system in Japanese quails (*Coturnix japonica*). *Kafkas Univ Vet Fak Derg*, 16 (4): 537-546, 2010.
36. Doitsidou M, Reichman-Freid M, Stebler J, Kopranner M, Dorries J, Meyer D, Esguerra CV, Leung T, Raz E: Guidance of primordial germ cell migration by the chemokine SDF-1. *Cell*, 111, 647-659, 2002.
37. Wang Q, Zhao XF, Ji YL, Wang H, Liu P, Zhang C, Zhang Y, Xu DX: Mitochondrial signaling pathway is also involved in bisphenol A induced germ cell apoptosis in testes. *Toxicol Lett*, 199 (2): 129-135, 2010.
38. Oka T, Adati N, Shinkai T, Sakuma K, Nishimura T, Kurose K: Bisphenol A induces apoptosis in central neural cells during early development of *Xenopus laevis*. *Biochem Biophys Res Commun*, 312 (4): 877-882, 2003.

Importance of Vaccination Against Atrophic Rhinitis in Pigs on Average Daily Gain and Mortality Rate

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Summary

The aim of this study was to investigate the effect of pigs, sows and piglets vaccination against atrophic rhinitis (AR) on the production results, the average daily gain (ADG) and mortality rate. Unvaccinated piglets, originating from vaccinated sows had a slightly higher ADG and only slightly less mortality than the control piglets (unvaccinated piglets and their mothers). Vaccinated piglets had a significantly higher ($P<0.01$) ADG and almost twice lower mortality rate than piglets unvaccinated, whether originating from vaccinated or unvaccinated mothers. Based on the observed results it is concluded that vaccination of pigs against AR significantly increases ADG and reduces mortality, and vaccination of sows against AR only slightly improves production performance and it is not economically justified for the studied farm.

Keywords: Atrophic rhinitis, Vaccination, Average daily gain, Mortality rate, Pig

Domuzlarda Atropik Rinite Karşı Aşılamanın Ortalama Günlük Kazanç ve Mortalite Oranı Üzerindeki Önemi

Özet

Bu araştırmanın amacı, domuzların, dişi domuzların ve domuz yavrularının atropik rinite (AR) karşı aşılama çalışmalarının üretim sonuçları, günlük kazanç oranı (ADG) ve mortalite oranı üzerine etkisini araştırmaktır. Aşılanmış domuz, yavruları aşılanmış ya da aşılanmamış annelerden olup olmadığına bakılmaksızın aşılanmamış yavrulara göre yüksek ADG ($P<0.01$) ve neredyse iki kat düşük mortalite oranı gösterdiler. Bu sonuçlara dayanarak, domuzlarda AR'e karşı aşılamanın önemli derecede ADG'ni arttırdığı ve mortaliteyi düşürdüğü, ve dişi domuzların AR'e karşı aşılanmasının sadece az miktarda üretim performansını arttırdığı ve bunun çalışılan çiftliklerde ekonomik olarak anlamlı olmadığı sonucuna varılmıştır.

Anahtar sözcükler: Atropik rinit, Aşılama, Ortalama Günlük Kazanç, Mortalite Oranı, Domuz

INTRODUCTION

Gram-negative bacteria *Pasteurella multocida* and *Bordetella bronchiseptica* are the primary infectious etiologic factors of atrophic rhinitis (AR). It is characterized by the formation of thick dry crusts in a roomy nasal cavity, which are result of progressive wasting away or decrease in size (atrophy) of the mucous nasal lining (mucosa) and underlying bone ^[1]. The various symptoms include foetor (strong offensive smell), crusting/nasal obstruction, nosebleeds, anosmia (loss of smell) or cacosmia (hallucination of disagreeable odour), secondary

infection, maggot infestation, nasal deformity, pharyngitis, otitis media and even, rarely, extension into the brain and its membranes ^[2]. Atrophic rhinitis can be classed as primary or, where it is a consequence of another condition or event, secondary. Infections of pigs with *P. multocida* and *B. bronchiseptica* can also result in bronchopneumonia ^[3,4]. Moreover, AR causes significant global economic loss in swine production due to growth retardation ^[5].

Vaccines to control AR have exploited knowledge of



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the pathogenesis of AR. Vaccines have been assessed for efficacy in reducing clinical signs and lesions of AR following both natural challenge in field trials and artificial challenge in controlled experiments [5]. Researchers have also evaluated the efficacy of AR vaccines in improving the growth rate of pigs, eliminating infection by toxigenic *P. multocida* and *B. bronchiseptica* from a farm, and inducing immune responses in sows and piglets to these bacteria [6,7].

On the basis of this, the objective of this investigation was to evaluate the effect of commercial vaccines against AR on the average daily weight gain and mortality rate.

MATERIAL and METHODS

Experimental Animals

The experiment was performed on a pig farm, with a capacity of 2500 sows, with an intensive way of keeping the pigs infected with AR enclosed. Infection status was assessed by serologic testing and detection of *Pasteurellae* and *Bordetellae* genom in serum. The experiment was performed on 4 groups of sows and their piglets (A, B, C and D) of the breeds Landras x Large Yorkshire hybrid (F₁), Large Yorkshire, and Landrace. Group A piglets originated from group A sows, group B piglets originated from group B sows, group C piglets originated from group C sows, and group D piglets originated from group D sows. All the animals had tags on their ears and a tattooed number.

Vaccinations of Animals

Animals were vaccinated with commercially vaccine against AR to the manufacturer's instructions (IDT Biologika GmbH). The vaccine had inactivated bacteria *Bordetella bronchiseptica* and inactivated bacteria *Pasteurella multocida* (A type and D type) and toxoid (dermonecrotxin).

Vaccination of Sows

A total of 103 pregnant sows were used in this study. The health of the animals before vaccination was clinically normal. Two programs were performed: in groups A and B 50 sows were vaccinated twice at 5 and 2 weeks before parturition, and in groups C and D, 53 sows were unvaccinated. The vaccine was applied in 4 ml dosages, subcutaneous, behind the ear.

Vaccinations of Piglets

A total of 627 piglets were used in this study. Two programs were performed: in groups A and C 304 piglets were vaccinated twice at 35 and 56 days of age (the vaccine was applied in 1 ml dosages, subcutaneous, behind the ear), and in groups B and D 323 piglets were unvaccinated.

Average Daily Gain and Mortality

Each of the groups A and B contained 25 sows and their 152 piglets, group C contained 26 sows and their 152 piglets and group D (control group) contained 27 sows and their 198 piglets. The piglets were selected by random sampling method. After weaning, all piglets were housed in one building and were divided by 10 piglets per box. Average daily gain and mortality were monitored from 28th to 175th day of life.

Statistical Analyses

The data were processed by ANOVA and Post Hoc Test was used for comparison of the means of treatments. Statistical significance of differences between means was determined at the level of $P < 0.01$.

RESULTS

Influence of Vaccination on Average Daily Gain of Pigs (ADG)

Table 1 presents the results of the ADG in tested piglets. In group A piglets (sows and piglets vaccinated) ADG values were significantly higher ($P < 0.01$) than in group B piglets (sows vaccinated only). Between group A and group C piglets (vaccinated pigs only) there was no significant difference ($P > 0.01$) in ADG. Group A piglets had significantly higher ($P < 0.01$) ADG than group D piglets (control group). Group B piglets had significantly lower ($P < 0.01$) ADG than group C piglets. At the same time, group B piglets had higher ($P < 0.01$) ADG compared to control piglets (group D). Vaccinated piglets (group C) had significantly higher ($P < 0.01$) ADG compared to the control group.

Influence of Vaccination on Mortality of Piglets

To determine the effect of vaccination against AR on mortality of piglets, mortality was calculated using the

Table 1. Average daily gain

Tablo 1. Ortalama günlük kazanç

Group	Frequency	Percent	Mean	SD	MIN	MAX
GROUP A	152	23.2	682	33.36	620	740
GROUP B	152	23.2	632	45.10	570	730
GROUP C	152	23.2	678	49.90	570	765
GROUP D	198	30,3	615	64.64	480	740
Total	654	100.0				

initial number of the piglets in each observed group on the 28th day of life. Group C piglets had the lowest number of deaths (5.3%), which is almost twice lower value compared to the control piglets (group D). Unvaccinated piglets whose mothers were vaccinated (group B) had a very high mortality (9.2%), almost as in control group piglets. Vaccinated piglets, whose mothers were also vaccinated, had a mortality of 6.6% (Table 2).

Table 2. Mortality rate (%)

Tablo 2. Mortalite oranı (%)

Group			
A	B	C	D
6.6	9.2	5.3	10.1

DISCUSSION

The results of this study clearly indicate that vaccination of pigs against AR on studied farms significantly influenced the increase of the ADG and reduced mortality of piglets. Vaccinated piglets, originating from vaccinated and non-vaccinated sows had about 10% higher daily gain, reducing the feeding duration for 10 days as determined in their studies by Liao et al.^[5]. In vaccinated piglets there were almost half the deaths than in non-vaccinated pigs^[8]. Unvaccinated piglets, originating from vaccinated sows had a slightly higher ADG and only slightly less mortality than the control piglets. This is a consequence of the termination of colostral immunity and the occurrence of infection in the same piglets^[9,10]. Evaluation of the safety and efficacy vaccines in this study suggest that they have potential in reducing the cost of AR in commercial farms. However, based on the experience of others^[8,11], these vaccines are unlikely to eliminate the causative agents of AR, *P. multocida* and *B. bronchiseptica* from a farm. By vaccinating the piglets better production results were achieved. This justifies the vaccination economically and at the same time on the studied farm there is no need for sows vaccination. The vaccinated piglets originating from both vaccinated and unvaccinated sows show no significant difference in ADG and mortality rate is almost identical. Nielsen^[12] found that there is no difference in ADG between piglets originating from vaccinated and unvaccinated mothers during lactation period. Considered that, as well as the results obtained in this study, where there is almost no significant difference in production results between unvaccinated piglets originating from vaccinated and unvaccinated sows, economic justification was not confirmed vaccination of sows on farm examined. In the previous researches performed by other authors^[13,14] it has been established that vaccination of older pigs, has little benefit against clinical AR and is generally less effective than sow vaccination. However recent results in researches show that there is no difference between ADG and mortality rate with pigs derived from vaccinated and

non-vaccinated sows. Distinction in ADG and mortality rate shall be shown in at a later stage of life, aged 90 days and over, so pigs vaccination has been recommended in herds in which AR causes have been established^[15].

Vaccination of piglets has unquestionable beneficial effect but current vaccines are not able to eliminate the bacteria. Optimal results can only be obtained when vaccination is combined with husbandry and management measures such as all-in/all-out production, correct vaccination, hygiene, reduction of stocking density and avoiding of stress^[16].

REFERENCES

1. Lee JM, Kang SY, Park SI, Woo HJ and Kwon MS: Molecular cloning and characterization of the gene for outer membrane protein H in a *Pasteurella multocida* (D:4) isolate from pigs with atrophic rhinitis symptoms in Korea. *J Microbiol Biotechnol*, 14, 1343-1349, 2004.
2. Sakano T, Okada M, Taneda A, Mukai T, Sato S: Effect of *Bordetella bronchiseptica* and serotype D *Pasteurella multocida* bacterin-toxoid on the occurrence of atrophic rhinitis after experimental infection with *B. bronchiseptica* and toxigenic type A *P. multocida*. *J Vet Med*, 59 (1): 55-57, 1997.
3. Brockmeier SL, Register KB, Magyar T, Lax AJ, Pullinger GD, Kunkle RA: Role of the dermonecrotic toxin of *Bordetella bronchiseptica* in pathogenesis of respiratory disease in swine. *Infect Immun*, 70, 481-490, 2002.
4. Davies RL, MacCorquodale R, Baillie S, Caffrey B: Characterization and comparison of *Pasteurella multocida* strains associated with porcine pneumonia and atrophic rhinitis. *J Med Microbiol*, 52, 59-67, 2003.
5. Liao CM, Huang C, Hsuan SL, Chen ZW, Lee WC, Liu CI, Winton JR, Chien MS: Immunogenicity and efficacy of three recombinant subunit *Pasteurella multocida* toxin vaccines against progressive atrophic rhinitis in pigs. *Vaccine*, 24, 27-35, 2006.
6. To H, Someno S, Nagai S: Development of a genetically modified nontoxic *Pasteurella multocida* toxin as a candidate for use in vaccines against progressive atrophic rhinitis in pigs. *Am J Vet Res*, 66 (1): 113-118, 2005.
7. Hsuan SL, Liao CM, Huang C, Winton JR, Chen ZW, Lee WC, Liao JW, Chen TH, Chiou CJ, Yeh KS, Chien MS: Efficacy of a novel *Pasteurella multocida* vaccine against progressive atrophic rhinitis of swine. *Vaccine*, 27, 2923-2929, 2009.
8. Kabay MJ, Mercy AR, Lloyd JM and Robertson GM: Vaccine efficacy for reducing turbinate atrophy and improving growth rate in piggeries with endemic atrophic rhinitis. *Australian Vet J*, 69 (5): 101-103, 1992.
9. Tizard IR: Veterinary Immunology. 6th ed., London, WB Saunders, 89: p. 223, 2000.
10. Rooke AJ, Carranca C, Bland MI, Sinclair GA, Ewen M, Bland CVI, Edwards AS: Relationship between passive absorption of immunoglobulin G by the piglet and plasma concentrations of immunoglobulin G at weaning. *Livestock Prod Sci*, 81, 223-234, 2003.
11. Foged NT, Nielsen JP, Jorsal SE: Protection against progressive atrophic rhinitis by vaccination with *Pasteurella multocida* toxin purified by monoclonal antibodies. *Vet Rec*, 125, 7-11, 1989.
12. Nielsen JP, Foged NT, Sorensen V, Barfod K, Bording A and Petersen SK: Vaccination against progressive atrophic rhinitis with a recombinant *Pasteurella multocida* toxin derivative. *Can J Vet Res*, 55, 128-138, 1991.
13. Giles CJ, Smith IM: The value of vaccinating pigs with *Bordetella bronchiseptica*. *Pig Vet Soc Proc*, 9, 61, 1982.
14. Giles CJ, Smith IM: Vaccination of pigs with *Bordetella bronchiseptica*. *Vet Bull*, 53, 327, 1983.
15. Kang ML, Shin SW, Rayamahji N, Seo YS, Lee SI, Lee WH, Yoo HS:

Field efficacy of a combined vaccine supplemented with recombinant *Pasteurella multocida* toxin subunits against atrophic rhinitis. *Korean J Vet Res*, 48 (1): 53-60, 2008.

16. Haesebrouck F, Pasmans F, Chiers K, Maes D, Ducatelle R, Decostere A: Efficacy of vaccines against bacterial diseases in swine: what can we expect? *Vet Microbiol*, 100, 255-268, 2004.

Kontrol Günü Süt Verimlerinin Zaman Serisi Yöntemi ile Modellenmesi ^{[1][2]}

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Özet

Bu çalışmanın amacı kontrol günü süt verimlerinin zaman serisi yöntemi ile modellenmesi ve en isabetli öngörülerini sağlayan kontrol günü sayısının belirlenmesidir. Bu amaçla 1070 süt siğirına ait 10700 kontrol günü kaydı kullanılmıştır. Veri seti her birinde 5350 kayıt bulunan iki guruba ayrılmıştır. Bu gruplardan biri model parametrelerini tahmin amacı ile, diğeri ise modelin öngörü başarısını değerlendirmek ve en isabetli öngörülerini sağlayan kontrol günü sayısını belirlemek amacı ile kullanılmıştır. Çalışma sonucunda ARIMA(2,0,0)(1,1,1)₁₀ modeli uygun model olarak belirlenmiş ve öngörü değerleri bu model kullanılarak elde edilmiştir. Söz konusu modelin tahmin edilen parametre değerleri ile gösterimi $(1-B^{10})^1y_t = [(1-0.99129B^{10})/(1-0.36889B-0.06934B^2)(1-0.08352B^{10})]a_t$ şeklindedir. Gerçek değerler ile öngörü değerleri arasında yüksek ve istatistiksel olarak anlamlı korelasyonlar saptanmıştır. Sonuçlar zaman serisi yaklaşımının süt veriminin öngörüsünde kullanışlı olabileceğini ortaya koymaktadır.

Anahtar sözcükler: Kontrol günü, Süt verimi, Zaman serisi, ARIMA, Öngörü

Modeling the Test Day Milk Yields via Time Series Method

Summary

The aim of this study is to model the test day milk yields via time series methodology and to determine the number of test days which provide the most accurate forecasts. For this purpose, 10700 test day records belonging to 1070 dairy cattle were used. Data were divided into two groups of 5350 records in each. One set of observations was used to model parameters, while the remaining was used for evaluating the forecast power of the model and for determining the number of test day records which provide the most accurate forecasts. ARIMA(2,0,0)(1,1,1)₁₀ model was determined to be suitable and it was used to obtain the forecast values. The expression of the model using estimated parameter values is $(1-B^{10})^1y_t = [(1-0.99129B^{10})/(1-0.36889B-0.06934B^2)(1-0.08352B^{10})]a_t$. Statistically significant and high correlations were determined between the actual and forecast values. The results indicated that the time series approach can be useful for prediction of milk yields.

Keywords: Test day, Milk yield, Time series, ARIMA, Forecast

GİRİŞ

Süt verimini modellemeye yönelik çalışmaların 1900'li yılların başlarından itibaren yürütüldüğü bilinmektedir ^[1]. Siğiraların süt veriminin modellenmesi için yapılan çalışmalar kısmi ya da tamamlanmış laktasyon kayıtlarından elde edilen günlük, haftalık ya da aylık süt miktarları üzerinden doğrusal ya da doğrusal olmayan deterministik modeller tahminlemek üzerine kurulmuştur ^[2]. t zamanındaki süt verimi y_t ; $y_t = f(t) + \varepsilon_t$ veya $y_t = f(t) \times \varepsilon_t$ gibi iki mümkün şekilde modellenebilir ^[3]. Burada, f(t) sürekli ve laktasyonun uzun-

luğuna bağlı olarak tüm zaman aralıklarında tanımlı bir fonksiyon iken ε_t , şansa bağlı hata terimlerini simgelemektedir ^[4]. Wood, Gamma, Ali-Schaeffer, Glasbey, Logaritmik, Wilmlink, Goodall modelleri laktasyon eğrisinin modellenmesinde yaygın olarak kullanılmaktadır ^[5-8].

Farklı sağım günlerinde ölçülen kontrol günü süt verimlerinin genetik olarak farklı değişkenler olarak adlandırılması ve çok değişkenli yöntemlerle analiz edilmesi



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mümkündür ^[9]. Bununla birlikte, yaygın olan tek değişkenli yaklaşıma göre her bir laktasyonda farklı zamanlarda alınan süt verimleri aynı deneme ünitesindeki tekrarlanan ölçümler olarak adlandırılabilir ^[10]. Laktasyon dönemi süresince belirli zaman aralıklarıyla yapılan ölçümler zamana bağlı olduğu için kurulan regresyon modellerindeki artıklar arasında bir ilişkinin olabileceği düşünülebilir. Aynı şekilde her kontrolde elde edilen verimin bir önceki verimlerle ilişkisi de söz konusu olmaktadır ^[3].

Süt veriminin modellenmesinde kullanılan farklı bir yaklaşım ise Deluyker ve ark.^[2], tarafından yapılan çalışmada ortaya konmuştur. İlgili çalışmada Holstein ırkı ineklere ait süt verim kayıtları zaman serisi yöntemi ile modellenmiştir. Macciotta ve ark.^[9,11], Sarda koyunu ve Simmental ırkı ineklere ait kontrol günü süt verimlerini zaman serisi yöntemi ile modelledikleri çalışmalarını yayınlamışlardır. Söz konusu çalışmalar, zaman serisi yönteminin, laktasyon eğrisinin şeklini tanımlamada ve belirli sayıda kontrol günü kaydı mevcut olduğunda hayvanların devam eden laktasyondaki süt verimlerinin öngörüsünde kullanışlı olduğunu ortaya koymuştur.

Bu çalışmanın amacı daha çok ekonomi alanında kullanılan ve bir seriye ait değerlerin kendi geçmiş değerleri, güncel ve geçmiş dönem rassal artıklarının ağırlıklı toplamı yardımıyla modellenmesine olanak sağlayan tek değişkenli zaman serisi analizi ile süt sığırlarının kontrol günü süt verimlerinin modellenmesidir. Ayrıca, zaman serisi modellerinin öngörü başarısından yararlanarak en isabetli öngörülerini sağlayan kontrol günü sayısının belirlenmesi de amaçlanmıştır.

MATERYAL VE METOT

Hayvan Materyali

Bu çalışmanın hayvan materyalini İngiltere Ulusal Süt Kayıtları Birliği'nde kaydı tutulan ve ilk kontrolleri Kasım 1988 ile Ekim 1989 tarihleri arasında yapılmış olup 10 adet verim kaydı bulunan 1.070 İngiliz Siyah Alaca düvesi oluşturmaktadır. Verim kayıtları yaklaşık olarak birer aylık aralıklarla alınmıştır. Her bir kontrol günü süt verimi 24 saatlik süre içerisinde alınan bireysel günlük süt verimlerinin toplamıdır ve kg cinsinden kayıt edilmiştir.

Zaman Serisi Analizi

Bir zaman serisi, zaman içerisinde sıralı olarak toplanan gözlem değerlerinin dizisi olarak tanımlanabilir ^[12]. Zaman vasfının gün, hafta, ay, üç ay, yıl vb. periyotlarını simgeleyen şıkları $t=1,2,...,T$ olarak ifade edilirse, Y değişkenine ait bu dönemlerde elde edilen gözlem değerleri de $y_1, y_2, ..., y_T$ şeklinde gösterilirler ^[13].

Zaman serisi ile iyi uyum gösteren bir modelin belirlenme aşamasında ilk olarak serinin durağan olması veya olmaması, mevsimsel özelliklere sahip olup olmaması gibi

özellikler ön plana çıkmaktadır ^[14]. Durağan olmayan Y serisinin ortalamada durağan hale getirilmesi için d -kez farkının alınması gerekiyorsa, durağan w_t serisi B olarak ifade edilen geri kaydırma işlemcisi yardımıyla $w_t = (1-B)^d y_t$ şeklinde gösterilmektedir ^[12]. Mevsimsel seyri olan serinin mevsimselliğini ortadan kaldırmak amacı ile D -kez fark alma işlemi uygulandığında durağan w_t serisi ise $w_t = (1-B^s)^D y_t$ şeklinde gösterilmektedir ^[14].

Zaman serisinin belirli bir noktada aldığı değer, y_t , serinin p adet geçmiş değeri ve bir şok ile tanımlanan bir fonksiyona sahip ise, serinin altında yatan veri yaratma süreci p 'inci mertebeden otoregresif süreç, $AR(p)$, olarak tanımlanmaktadır ^[15]. Bir zaman serisinin bugünkü değeri, mevcut ve q adet geçmiş dönemdeki rassal şokun ağırlıklı toplamı ise süreç q mertebesinde hareketli ortalama, $MA(q)$, sürecidir ^[4]. Gerçek hayatta karşılaşılan pek çok durumda ise her iki süreci de içeren karma otoregresif hareketli ortalama modeli, $ARMA(p,q)$, ile çalışmak söz konusu olmaktadır.

Zaman serilerinde gözlenen periyodik ya da mevsimsel hareketler, birbirini izleyen ölçüm dönemlerinde aynı ölçüm zamanları arasında gözlenen benzer yükselme ya da düşme hareketleri olarak tanımlanmaktadır. Bu tür serilerde mevsimsel ilişkinin yanında ardışık gözlemler arasında da ilişki ortaya çıkabilmektedir. $ARIMA(p,d,q)(P,D,Q)_s$ olarak gösterilen çarpımsal mevsimsel $ARIMA$ modelleri ile mevsim içerisindeki ve mevsimler arasındaki bağımlılık yapısı birlikte modellenmektedir ^[4].

$$(1-B)^d (1-B^s)^D y_t = \frac{(1 + \theta_1 B + \dots + \theta_q B^q)(1 + \Theta_1 B^s + \dots + \Theta_Q B^{Qs})}{(1 - \phi_1 B - \dots - \phi_p B^p)(1 - \Phi_1 B^s - \dots - \Phi_P B^{Ps})} a_t$$

Burada, θ mevsimsel olmayan MA parametrelerini, q mevsimsel olmayan MA parametre sayısını, ϕ mevsimsel olmayan AR parametrelerini, p mevsimsel olmayan AR parametre sayısını, Φ mevsimsel AR parametrelerini, P mevsimsel AR parametre sayısını, Θ mevsimsel MA parametrelerini ve Q mevsimsel MA parametre sayısını simgelemektedir.

Zaman serisini oluşturan sürecin olasılık dağılımını belirlemek ve kesin bir tanımlamasını yapmak için otokorelasyon fonksiyonu (OKF) ve kısmi otokorelasyon fonksiyonu (KOKF) kullanılmaktadır ^[14].

Veri Setinin Hazırlanması ve İstatistiksel Analizler

Klasik zaman serisi tertibinden farklı olarak bu çalışmada kontrol günü süt verimlerini içeren veri seti Macciotta ve ark.^[9,11] tarafından uygulanan bir yaklaşımla hazırlanmıştır (Tablo 1). Macciotta ve ark.^[9], indeks değişkeninin, içerdiği anlam ne olursa olsun çıktığı, yani Y değerlerini, sıralamada kullanılan bir araç olduğunu ve bu nedenle herhangi bir zaman kavramına bağlı olmak zorunda olmadığını belirtmişlerdir. Veri setinin izlenecek yönetime uygun olarak düzenlenmesi amacıyla öncelikle

hayvanlar rastgele sıralanmış, ardından laktasyon kayıtları her bir hayvan içerisinde sıraya konmuştur. Toplamda 10.700 laktasyon kaydına karşılık gelen gözlemlere 1'den 10.700'e kadar bir indeks değeri atanmıştır. Örneğin 26 numaralı indeks değeri 3. sıradaki hayvanın 6. laktasyon kaydını belirtmektedir.

Veri seti 535 hayvanın 5350 kontrol günü süt verimi kaydını içeren iki ayrı kısma ayrılmıştır. Model parametreleri SAS programının ARIMA prosedüründe ilk 535 hayvana ait kayıtlar için en çok olabilirlik yöntemi ile tahmin edilmiştir^[16]. Modelin öngörü başarısı model tahmin basamağında elde edilen parametre değerleri ve ikinci gruptaki 535 hayvanın kayıtları kullanılarak değerlendirilmiştir. En isabetli öngörülerini sağlayan minimum kayıt sayısını belirlemek amacı ile, her bir hayvanın bilindiği varsayılan 2, 3, 4, 5, 6, 7 ve 8 kaydı kullanılarak geriye kalan, sırasıyla 8, 7, 6, 5, 4, 3 ve 2 kaydını öngörmek şeklinde bir algoritma işletilmiştir. Gerçek değerler ile öngörü değerleri arasındaki korelasyonlar SAS programının CORR prosedürü kullanılarak elde edilmiştir.

Zaman serisi analizi daha çok ekonomi alanında uygulama olanağı bulduğundan kullanılan terminoloji de

bu alana özgüdür. Çalışmada, ele alınan veri seti dikkate alındığında; zaman-indeks, yıl-her bir hayvana ait laktasyon dönemi, ay-kontrol günü ve mevsim-laktasyonun pik verim dönemi ya da artış veya azalış seyri izlediği dönemler vb. kavramlarının karşılığı olarak yorumlanmalıdır.

BULGULAR

*Tablo 2'*de model kurma basamağında kullanılan veri setine ilişkin bazı tanımlayıcı istatistikler sunulmuştur. Beklendiği gibi en yüksek ortalama süt verimi 2. kontrol gününde saptanmış olup ortalamalar tipik laktasyon eğrisine benzer bir seyir izlemektedir. *Şekil 1* kontrol günü süt verimlerinin zaman serisi grafiğini göstermektedir. *Şekilden* de görüleceği gibi süt verimi belirli bir bandın dışına çıkmamakta ve alçalma ya da yükselme yönünde herhangi bir eğilim göstermemektedir.

*Şekil 2'*de sunulan otokorelasyon katsayılarının 0,10, 20,..., gecikmelerinde zirve yapması periyodik bileşenin varlığına işaret etmektedir. OKF(1), OKF(10) ve OKF(20) değerleri yüksek derecede bir ilişkinin varlığını ortaya koymaktadır. Benzer şekilde yüksek korelasyonlar OKF(5), OKF(15) ve OKF(25)'te de saptanmıştır. *Şekil 2'*den korelasyonların güven sınırları dışına çıktığı ve bu nedenle de

Tablo 1. Çalışmada kullanılan veri setinin yapısı

Table 1. Structure of the data used in the study

Sıgır	Kontrol Günü	Verim	İndeks
1	1	*	1
1	2	*	2
1	3	*	3
1	4	*	4
1	5	*	5
.			.
.			.
1070	7	*	10697
1070	8	*	10698
1070	9	*	10699
1070	10	*	10700

* hayvanın ilgili kontrol günündeki toplam süt verimi (kg)

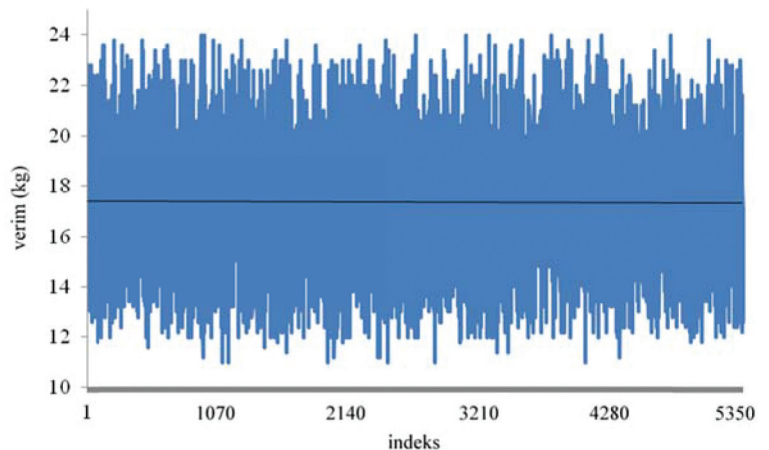
Tablo 2. Veri setine ilişkin bazı tanımlayıcı istatistikler

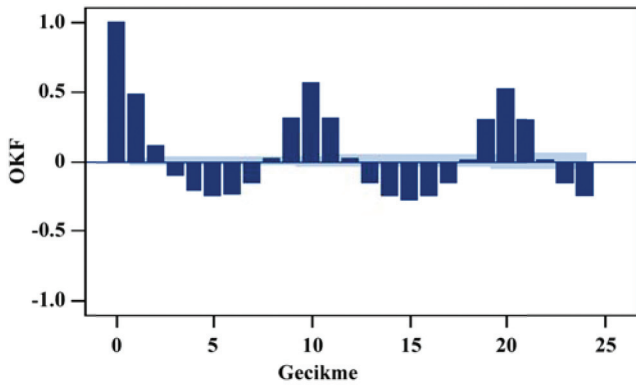
Table 2. Some descriptive statistics of the data

Kontrol Günü	N	Ortalama	Std. Sapma
1	535	19.18	2.118
2	535	20.30	1.878
3	535	19.56	1.840
4	535	18.52	1.731
5	535	17.63	1.787
6	535	16.89	1.790
7	535	16.51	1.902
8	535	15.94	1.916
9	535	15.15	1.809
10	535	14.06	1.384

Şekil 1. Süt verimlerinin zaman serisi grafiği

Fig 1. Time sequence plot of milk yield data





Şekil 2. Orjinal verilere ait OKF grafiği

Fig 2. ACF plot of original data

Tablo 3. Parametre tahminleri

Table 3. Parameter estimates

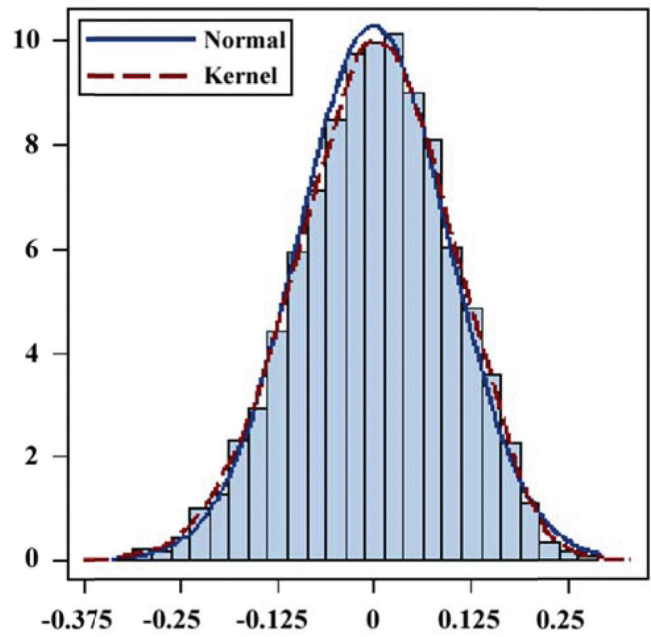
Parametre	Tahmin	Standart Hata	t-Değeri	P	Gecikme
SMA	0.99129	0.0022861	433.61	<.0001	10
AR1	0.36889	0.01362	27.08	<.0001	1
AR2	0.06934	0.01361	5.09	<.0001	2
SAR	0.08352	0.01377	6.07	<.0001	10

SMA (Seasonal MA): Mevsimsel MA, AR1-2 (Autoregressive): Otoregresif parametreler, SAR (Seasonal AR): Mevsimsel AR parametresi

istatistiksel olarak önemli oldukları görülmektedir. Bu durum veri setinin oluşturulma şekli düşünüldüğünde beklenen bir sonuçtur. Her ne kadar çalışmanın materyalini oluşturan kontrol günü süt verim kayıtları rastgele sıralanmış olsa da teorik olarak her bir laktasyonda aynı dönemde pik verimine ulaşıldığı bilinmektedir. Periyodik gecikmelerde ortaya çıkan bu yüksek otokorelasyonlar ve bu otokorelasyonların aynı seviyede varlığını devam ettirmesi, güçlü bir mevsimsellik göstergesi olup veri setine mevsimsel fark işlemi uygulanması gerektiğini göstermektedir.

Mevsimsel farkı alınmış seriye ait OKF incelenmiş ve gecikme uzunluğu arttıkça OKF'nun hızla azalıp sıfırı kestiği, bununla birlikte orjinal seriye ilişkin OKF'nda 10 ve 10'un katlarında gözlenen periyodik davranışın giderildiği belirlenmiştir. Buna karşın 1. ve 10. gecikmelerdeki otokorelasyonların model oluşturulurken dikkate alınması gerektiği saptanmıştır. Bu nedenle MA sürecinin mevsimsel olmayan ve mevsimsel olan kısımlarını simgeleyen, q ve Q mertebeleri, 1 olarak belirlenmiştir. Mevsimsel farkı alınmış seriye ait KOKF'nun sıfırı ikinci gecikmede kestiği görüldüğünden mevsimsel olmayan AR mertebesinin 2 olduğu düşünülmüştür. Ayrıca ilk iki mevsimsel gecikmede önemli olan kısmi otokorelasyonların da dikkate alınması gerektiği saptanmıştır.

Analizin sonraki aşamasında, aday model olarak belirlenen çarpımsal-mevsimsel ARIMA modeline ait parametreler tahmin edilmiş ve mevsimsel AR mertebesinin 2 olması durumunda ilgili parametrenin anlamsız olduğu ($P>0.05$) görüldüğünden analiz dışında tutulmasına karar



Şekil 3. Artıkların histogramı ile normal ve kernel yoğunlukları

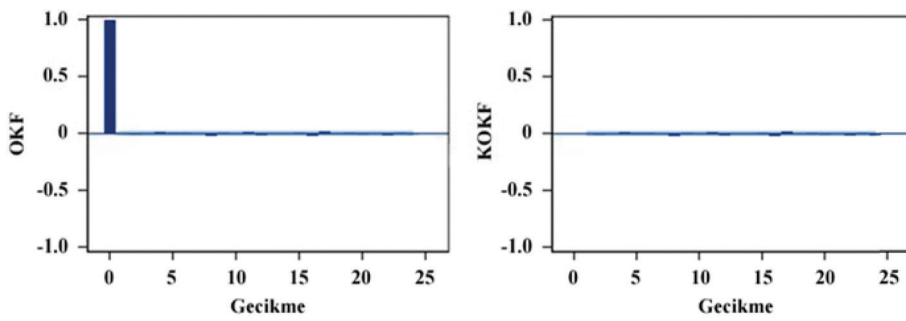
Fig 3. Histogram with normal and kernel density plots of the residuals

verilmiştir. Serinin D=1 olmak üzere 1 kez mevsimsel farkı alındığından nihai model ARIMA(2,0,0)(1,1,1)₁₀ modeli olarak belirlenmiş ve bu modelin en çok benzerlik yöntemi ile tahmin edilen parametre değerleri Tablo 3'te sunulmuştur. Söz konusu model parametrelerinin tümünün istatistiksel olarak önemli olduğu belirlenmiştir ($P<0.05$). Bununla birlikte, model artıklarına ilişkin ilk 48 gecikme için hesaplanmış olan ki-kare değerleri için $P>0.05$ olduğu saptanmış ve modele herhangi bir AR ya da MA terimi eklenmesine gerek olmadığı sonucuna ulaşılmıştır.

Model artıklarının Şekil 3'te yer alan dağılımı incelendiğinde artıkların yaklaşık sıfır ortalama etrafında normal dağılışa sahip olduğu görülmektedir. Artıklara ilişkin OKF ve KOKF'karı da modelin veri setini tanımlamada uygun olduğunu göstermektedir (Şekil 4). Model artıklarına ilişkin otokorelasyon ve kısmi otokorelasyonların tümü güven sınırları içerisinde kaldığından, bir başka ifade ile model artıkları arasında istatistiksel açıdan önemli korelasyonlar olmadığından modelin doğru bir şekilde tanımlandığı sonucuna ulaşılmıştır. Model, tahmin edilen parametre değerleri ile aşağıdaki gibi gösterilmektedir:

$$(1 - B^{10})^1 y_t = \frac{(1 - 0,99129B^{10})}{(1 - 0,36889B - 0,06934B^2)(1 - 0,08352B^{10})} a_t$$

Çalışmada gerçek değerler ile modelden tahmin edilen değerler arasındaki korelasyon 0.90 olarak bulunmuştur. Mevcut gözlem sayısı arttıkça model yardımı ile öngörüsü yapılan değerler ile serinin gerçek değerleri arasındaki korelasyonların artma eğiliminde olduğu görülmektedir (Tablo 4). Bununla birlikte, öngörüsü yapılacak kontrol günü (ay) son kontrol kaydının alındığı tarihten uzaklaştıkça sözü



Şekil 4. Artıklara ait OKF ve KOKF grafikleri

Fig 4. ACF and PACF plots of the residuals

Tablo 4. Mevcut gözlem değerleri ile öngörü değerleri arasındaki korelasyonlar
Table 4. Correlations among the actual and forecasted values

Öngörüsü Yapılan Kontrol Günü (ay)	Mevcut Gözlem Sayısı						
	2	3	4	5	6	7	8
3	0.74						
4	0.74	0.82					
5	0.74	0.79	0.81				
6	0.75	0.78	0.80	0.85			
7	0.76	0.78	0.80	0.85	0.87		
8	0.76	0.78	0.79	0.83	0.87	0.87	
9	0.75	0.77	0.78	0.81	0.83	0.84	0.87
10	0.74	0.75	0.76	0.79	0.80	0.80	0.80

Tüm korelasyonlar için $P < 0.001$ dir

edilen değerler arasındaki korelasyonlar azalmaktadır. Çalışmanın bulguları 6 kaydın mevcut olduğu durumun serinin gelecekteki değerlerinin öngörüsü için yeterli seviyede bilgi içerdiğini göstermektedir.

TARTIŞMA ve SONUÇ

Macciotta ve ark.^[9], Sarda koyunlarına ait mevsimsellikten arındırılmış kontrol günü süt verimlerini modelledikleri çalışmalarında çarpımsal mevsimsel ARIMA(2,0,0)(1,0,1)₇ modelini uygun model olarak belirlemişlerdir. Bu model çalışmamızda belirlenen model ile örtüşmektedir. Bunun yanında Macciotta ve ark.^[11], Simmental ırkı inekler üzerinde yürüttükleri çalışmalarında çarpımsal mevsimsel ARIMA(1,0,1)(1,0,1)₈ modelini, Deluyker ve ark.^[2] ise Holstein ırkı ineklere ait kontrol günü süt verim kayıtlarını modelledikleri çalışmalarında ARIMA(0,1,1) modelini uygun model olarak belirlemişlerdir.

Macciotta ve ark.^[11], çalışmalarında süt verimi için 2 adet mevcut kontrol günü kaydını 3. kayıt gününe ait verimleri öngörmeye kullanmışlar ve gerçek değerler ile öngörü değerleri arasındaki korelasyonları 0.85 olarak saptamışlardır. Aynı şekilde 3, 4, 5 ve 6 kaydın olduğu durumlar için de bir sonraki kayda ilişkin öngörüler ile gerçek değerler arasında tüm durumlar için 0.85'lik korelasyonlar saptamışlardır. Buna karşın çalışmamızda 6 gözlemin

mevcut olduğu durumda 7 ve 8. gözlem değerlerine ilişkin gerçek değerler ve öngörü değerleri arasında her iki kontrol günü için 0.87'lik korelasyonlar saptanmıştır. Sözü edilen çalışmada korelasyonlar 0.85-0.44 arasında değişim göstermekte iken çalışmamızda tüm durumlar için korelasyonlar 0.87-0.74 aralığındadır.

Serilerin değerleri arasında az da olsa gözlenen farklılıkların daha homojen hayvan grupları ile çalışılması durumunda daha da azalması beklenmektedir. Ayrıca model tahmini yapılan hayvan grubu ile modelin öngörü başarısının değerlendirildiği hayvan grubunun da benzer koşullarda yetiştirilen ve genetik potansiyel olarak da birbirinin benzeri hayvanlar olmaları daha isabetli öngörüler yapılmasına olanak sağlayacaktır. Bu açıdan bakıldığında çalışmada ele alınan yöntemin sürü ya da işletme bazında kullanılması uygun olacaktır. Özellikle otomatik kontrol sistemlerinin kullanıldığı işletmelerde bireyler için tahmin edilen değerlerden büyük sapmalar görülmesi durumunda bu hataların değerlendirilmesi gerekmektedir.

KAYNAKLAR

1. Grossman M, Kuck AL, Norton HW: Lactation curves of purebred and crossbred dairy cattle. *J Dairy Sci*, 69 (1): 195-203, 1986.
2. Deluyker HA, Shumway RH, Wecker WE, Azari AS, Weaver LD: Modeling daily milk yield in Holstein cows using time series analysis. *J Dairy Sci*, 73, 539-548, 1990.
3. Goodall EA, Sprevak D: A Note on stochastic model to describe the milk yield of a dairy cow. *Anim Prod*, 38, 133-136, 1984.
4. Chatfield C: Time Series Forecasting, CRC Press LLC, NW, 2000.
5. Kellogg DW, Urquhart NS, Ortega AJ: Estimating Holstein lactation curves with a gamma curve. *J Dairy Sci*, 60, 1308-1315, 1977.
6. Keskin İ, Çilek S, İlhan F: Polatlı Tarım İşletmesi'nde yetiştirilen Siyah Alaca sığırların laktasyon eğrisi özellikleri. *Kafkas Univ Vet Fak Derg*, 15 (3): 437-442, 2009.
7. Druet T, Jaffrezic F, Boichard D, Ducrocq V: Modeling lactation curves and estimation of genetic parameters for first lactation test-day records of French Holstein cows. *J Dairy Sci*, 86, 2480-2490, 2003.
8. Sherchand L, McNew RW, Kellogg DW, Johnson ZB: Selection of a mathematical model to generate lactation curves using daily milk yields of Holstein cows. *J Dairy Sci*, 78, 2507-2513, 1995.
9. Macciotta NPP, Cappio-Borlino A, Pulina G: Time series autoregressive integrated moving average modeling of test-day milk yields of dairy ewes. *J Dairy Sci*, 83, 1094-1103, 2000.
10. Van der Werf J: Random Regression in Animal Breeding (Course Notes). Jaboticabal, SP Brazil, November, 2001.

11. Macciotta NPP, Vicario D, Pulina G, Cappio-Borlino A: Test day and lactation yield predictions in Italian Simmental cows by ARMA methods. *J Dairy Sci*, 85, 3107-3114, 2002.

12. Box GEP, Jenkins GM: Time Series Analysis: Forecasting and Control (Revised Edition), Holden-Day, USA, 1976.

13. Akgül I: Geleneksel Zaman Serisi Yöntemleri, 1. Baskı, Der Yayınları,

İstanbul, 2003.

14. Akgül I: Zaman Serilerinin Analizi ve ARIMA Modelleri, 1. Baskı, Der Yayınları, İstanbul, 2003.

15. Yaffee R, McGee M: Introduction to Time Series Analysis and Forecasting with Applications of SAS and SPSS. Academic Press, NY, 2000.

16. SAS Institute Inc.: SAS/STAT User's Guide, Version 9.2, 2009.

Molecular Characteristics of *Staphylococcus aureus* Isolates from Buffaloes Milk^[1]

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Summary

The aim of the present study was to analyse the *aroA* and X region of *spa* genes on *Staphylococcus aureus* isolates collected from milks of native buffaloes in north-west of Iran. For this purpose, seventy five *S. aureus* isolates were examined by PCR-RFLP. Amplification of *AroA* gene revealed a single amplicon with a size of approximately 1,153 bp, but restriction profile analysis of this amplicon yielded three different genotypes including genotype A (9.3 %), B (88%) and N (2.7%). Amplification of X region of the *spa* gene showed an amplicon of 1110; 1130; 1160; 1190 bp in 19, 15, 12 and 29 isolates, respectively. *HhaI* digestion of this amplicons demonstrated that the isolates having the same PCR band size were in the same PCR-RFLP patterns and there was not variation into isolates belonged to the same PCR amplicon.

Keywords: *Staphylococcus aureus*, *aroA* gene, *spa* gene, PCR-RFLP

Manda Sütünden İzole Edilen *Staphylococcus aureus* Türlerinin Moleküler Karakterizasyonu

Özet

Bu çalışmanın amacı, İran'ın Kuzey Batısı'nda yerli mandaların sütlerinden izole edilen *Staphylococcus aureus* izolatlarında *spa* genlerinin *aroA* ve X bölgelerini analiz etmektir. Bu amaçla, yetmiş beş *S. aureus* izolatı PCR-RFLP yöntemi ile incelendi. *AroA* gen amplifikasyonu, yaklaşık olarak 1,153 bp büyüklüğünde olan tek bir amplicon oluşturmaya rağmen bu amplicon restriksiyon profil analizi ile genotip A (9.3%), B (88%) ve N (2.7%) olmak üzere üç farklı genotipi oluşturduğu belirlendi. *spa* geninin X bölgesinin amplifikasyonu 19, 15, 12 ve 29 izolatla sırasıyla 1110, 1130, 1160 ve 1190 bp büyüklüğünde fragman ürettiği gözlemlendi. Bu ampliconun *HhaI* enzimi ile kesimi, aynı PCR bant büyüklüğüne sahip izolatların aynı PCR-RFLP profillerinde olduğunu ve aynı PCR ampliconuna ait izolatlar içinde varyasyon olmadığını gösterdi.

Anahtar sözcükler: *Staphylococcus aureus*, *aroA* gen, *spa* geni, PCR-RFLP

INTRODUCTION

Buffaloes account for about 5 percent of the milk production in the world but this rate is greater in the Near East and Asia. Buffalo milk is also considered as an important milk source in Iran^[1]. Clinical and subclinical mastitis are associated in decrease of milk production^[2].

Among pathogenic bacteria which have been isolated as etiological agents of mastitis in ruminants, *Staphylococcus aureus* has been mentioned as the main one. The prevalence of subclinical and clinical bovine, caprine and

ovine mastitis caused by *S. aureus* ranges from 5 to 50%, 5.6 to 17%, 0.22 to 11% in different countries, respectively^[3]. However, only a few studies on staphylococcal mastitis concerning buffalo infections have been published. Staphylococcal mastitis is important in ruminants of Iran and mastitis caused by this organism is a major cause of economical losses in the Iranian dairy industry^[4]. *S. aureus* is also one of the main pathogens isolated from buffaloes in this country^[5]. Some studies confirm this subject in other countries in Asia^[6,7].



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Different molecular methods such as PCR - restriction fragment length polymorphism (PCR-RFLP) analysis of the virulence genes [8-10], multilocus sequence typing [11], pulsed-field gel electrophoresis [12-14], multilocus enzyme electrophoresis [15,16] have been used molecular characterization of the *S. aureus* isolates. Molecular characterization of *S. aureus* isolated from bovine milk samples was studied by many researchers [17]. However, a few papers studied molecular profile of *S. aureus* isolated from buffaloes in worldwide [3,6].

In contrast of some molecular studies on bovine milk isolates of *S. aureus* in Iran [4,18], there are not similar studies in buffalo isolates. The aim of the present study was to characterize analysis of the *aroA* and *spa* gene of *S. aureus* isolates collected from native buffaloes in villages around of Tabriz city, Capital of East Azerbaijan province in Iran.

MATERIAL and METHODS

Samples

Seventy five *S. aureus* isolates were isolated from milk of buffaloes in Tabriz, Northwest of Iran from the microbiology laboratory of Islamic Azad University, Tabriz branch. All isolates had been confirmed by standard biochemical tests [4] and stored in brain heart infusion (BHI) with 30% glycerol at -70°C, previously.

DNA Extraction

A colony of Brain heart infusion (BHI) agar was transferred to a clean microtube and it was added 500 µl lysis buffer (pH 8) including (containing 5 mol NaCl, 1 mol Tris-base, 0.5 mol EDTA and CTAB 2%). Then incubated in 60-65°C for 2 h and centrifuged at 12.000 x g for 5 min. The pellet was resuspended in CHCl₃-isoamyl alcohol (24:1), and centrifuged for five min at 12.000 x g. Pellet was incubate with RNAase in 37°C for 30 min, then was resuspended in cold isopropanol and transfer to -20°C for 15 min. In the next step, ethanol 70% was added to

supernatant and centrifuged for one min at 12.000 x g. Finally, 50 µl TE-buffer was add to pellet and stored as DNA template.

Confirmation Isolates by PCR

All isolates were confirmed as *S. aureus* by the detection of *nuc* gene by PCR described by Brakstad et al. [19].

Molecular Typing by *aroA* Gene Restriction Profile Analysis

The *aroA* gene was amplified by PCR using the special primers [8]. PCR products were digested with 50 U *TaqI* (Fermentas) at 65°C for 3 h. All digested fragments were assessed by electrophoretic separation in 2% (w/v) agarose gels.

Molecular Typing by X Region of the *spa* Gene Restriction Profile Analysis

Amplification of the repeat region of the *S. aureus* protein A gene (*spa*) was performed by using a specific primer set as described previously [18,19]. PCR products were digested with 50 U *HhaI* (Fermentas) at 37°C for 3 h. All digested fragments were assessed by electrophoretic separation in 2% (w/v) agarose gels.

RESULTS

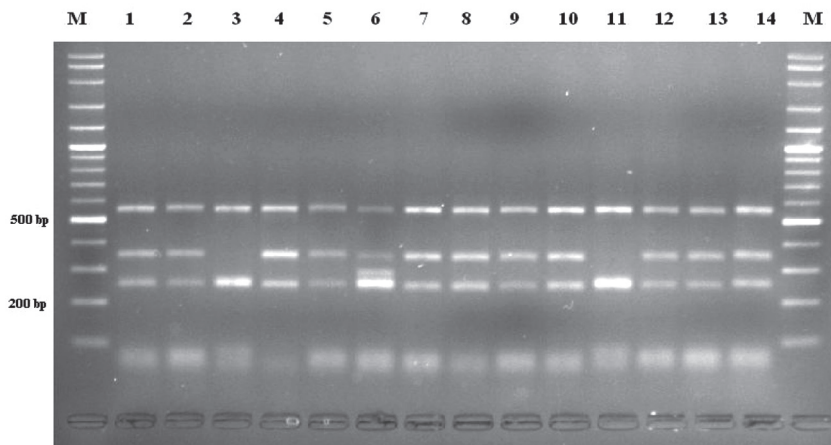
According to amplification of the *nuc* gene, all 75 isolates confirmed as *S. aureus*.

AroA Gene Restriction Profile Analysis

AroA gene amplification revealed a single amplicon with an expected size of approximately 1.153 bp, but restriction profile analysis of mentioned gene by U *TaqI* enzyme indicated three differentiated profile. Sixty-six of the 75 isolates (88%) shows a profile known as genotype B and 7 isolates (9.3%) as genotype A. Two isolates (2.7%) have 4 detectable bands and were considered a different genotype showed in this study (Fig. 1).

Fig 1. PCR-amplified *aroA* gene digested with the *TaqI*. Lane M: DNA marker, Lanes 1,2,4,5,7 to 10,12 to 14 Genotype B, Lanes 3,11 Genotype A, Lane 6 new genotype in this study (N)

Şekil 1. PCR ile amplifiye edilen *aroA* gen amplikonunun *TaqI* ile kesimi. Sıra M: DNA marker, Sıralar 1,2,4,5,7 'den 10,12'den 14'e Genotip B, Sıralar 3,11 Genotip A, Sıra 6 bu çalışmadaki yeni genotip (N)



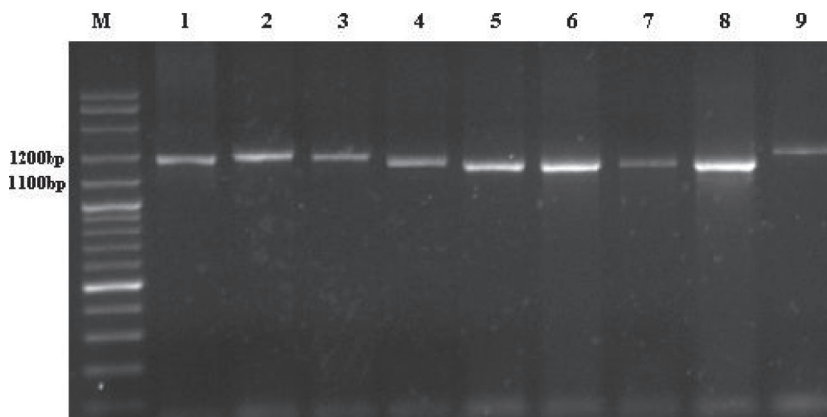
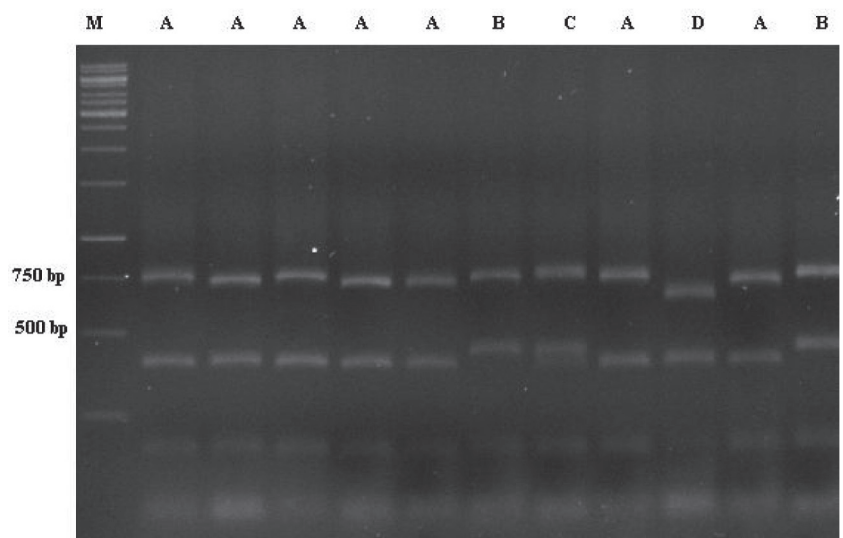


Fig 2. Agarose gel electrophoresis of PCR products obtained from amplification of the *spa* gene. **Lane M:** DNA marker, **Lanes 1,2,3:** 1160, **Lanes 4,7:** 1130, **Lanes 5,6,8:** 1110 and **Lane 9:** 1190 bp

Şekil 2. *spa* geninin amplifikasyonu sonucu elde edilen PCR ürünlerinin agaroz jel elektroforezi. **Sıra M:** DNA marker, **Sıralar 1,2,3:** 1160, **Sıralar 4,7:** 1130, **Sıralar 5,6,8:** 1110 ve **Sıra 9:** 1190 bp

Fig 3. PCR-amplified *spa* gene digested with the *HhaI*. **Lane M:** DNA marker, **Lane A:** pattern produces by 1190 bp band size, **Lane B:** pattern produces by 1130 bp band size, **Lane C:** pattern produces by 1160 bp band size and **Lane D:** pattern produces by 1110 bp band size

Şekil 3. PCR ile amplifiye edilen *spa* gen ampiklonunun *HhaI* ile kesimi. **Sıra M:** DNA marker, **Sıra A:** 1190 bp bant büyüklüğü üreten profil, **Sıra B:** 1130 bp bant büyüklüğü üreten profil, **Sıra C:** 1160 bp bant büyüklüğü üreten profil ve **Sıra D:** 1110 bp bant büyüklüğü üreten profil



Analysis of x Region of the *spa* Gene

All of the 75 *S. aureus* isolates were PCR positive for X region of *spa* gene. Amplification of the *spa* gene showed an amplicon of 1110; 1130; 1160; 1190 bp in 19, 15, 12 and 29 isolates, respectively (Fig. 2).

PCR-RFLP of X region of the *spa* gene demonstrated 4 profiles. Isolates with same PCR band size were same into the PCR-RFLP patterns and there was not variation into isolates belonged to same PCR amplicon (Fig. 3).

DISCUSSION

This paper is the first report describing molecular characteristic of *S. aureus* isolated from buffalo's milk. Studies of the molecular epidemiology of *S. aureus* collected from cows have been already published in the literature worldwide [17] and in Iran [4,18], but there is a few published studies of *S. aureus* isolated from buffalo [3,22].

Similar to *S. aureus* isolated from other source amplicon size of *aroA* gene is a 1,153-bp fragment and this confirmed *aroA* gene is specific for *S. aureus*, hence it could be used

as a powerful tool for the identification of *S. aureus* isolates in buffaloes, too.

AroA gene restriction profile studies showed none of *S. aureus* isolated from cattle were genotype C or D in previous [4,8,23]. Our study confirms this again in buffaloes once. It may suggest that genotype C or D have link to isolates from human origins. Marcos et al. [8] reported these two genotype especially D (44%) from human isolates. In animal, instead of genotype C or D, there are different RFLP patterns with various bands named N reported up to 50% in *S. aureus* with bovine origin [4,23]. In this study, we reported 2.7% these ones in buffalo's isolates. High prevalence of genotype B in animal origin isolates in the some studies, included present study, showed this genotype may has been link to animal source [4,8]. In contrast, Marcos et al. [8] reported low prevalence of genotype B in human isolates.

Koreen, et al. [24] suggested a new typing method for *S. aureus* based on sequencing of X region of staphylococcal protein A (*spa*) gene instead of expensive, time-consuming methods such as pulsed-field gel electrophoresis (PFGE), multilocus enzyme electrophoresis (MLEE), multilocus sequence typing (MLST). The X region of *spa* gene is very highly polymorphism that reflex epidemiological variation

among different *S. aureus* isolates. As a substitution for Koreen, et al.^[24] method, Wichelhaus et al.^[20] designed X region of *spa* gene based on PCR-RFLP, described previously. We used the same method in this study.

The amplicon size of X region of *spa* gene in different *S. aureus* has varieties because of polymorphism of this region of *spa* gene. The amplicon size of X region of *spa* gene in this study has not report in previous papers and it may be belonged to new *spa* type. However, the PCR-RFLP of the mentioned region of each different amplicon size have not produced variation in internal of each groups and this suggest sequence based *spa* typing for determination of new *spa* type of *S. aureus* from buffaloes origin. On the other hand, *spa* gene variation may be related with the geographical and host factors similar to that of *aroA* gene.

REFERENCE

1. Bromandi Jazi M: Buffaloes culture. Applied -scientific High educational institute, Tehran, 2006.
2. Seegers H, Fourichon C, Beaudeau F: Production effects related to mastitis and mastitis economics in dairy cattle herds. *Vet Res*, 34, 475-491, 2003.
3. Aires-de-Sousa M, Parente CESR, Vieira-da-Motta O, Bonna ICF, Silva DA, Lencastre H: Characterization of *Staphylococcus aureus* isolates from buffalo, bovine, ovine, and caprine milk samples collected in Rio de Janeiro State, Brazil. *Appl Environ Microbiol*, 73 (12): 3845-3849, 2007.
4. Dastmalchi Saei H, Ahmadi M: Genotyping of *Staphylococcus aureus* isolated from bovine mastitis based on PCR-RFLP analysis of the *aroA* gene. *Comp Clin Pathol*, 19, 163-168, 2010.
5. Beheshti R, Eshratkhan B, Shayegh, J, Giasi Ghalehkandi J, Dianat V, Valiei K: Prevalence and etiology of subclinical mastitis in Buffalo of the Tabriz region, Iran. *J Am Sci*, 7 (5): 642-645, 2011.
6. Sharma A, Sindhu N: Occurrence of clinical and subclinical mastitis in buffaloes in state of Haryana (India). *Ital J Anim Sci*, 6 (Suppl. 2): 965-967, 2007.
7. Sindhu N, Sharma A, Jain VK: Phenotypic and genotypic characterization of *Staphylococcus aureus* isolated from mastitis cases of cows and buffaloes. *J Immunol Immunopathol*, 10 (2): 119-123, 2008.
8. Marcos JY, Soriano AC, Salazar MS, Moral CH, Ramos SS, Smeltzer MS, Carrasco GN: Rapid identification and typing of *Staphylococcus aureus* by PCR-restriction fragment length polymorphism analysis of the *aroA* gene. *J Clin Microbiol*, 37, 570-574, 1999.
9. Ishino K, Tsuchizaki N, Ishikawa J, Hotta K: Usefulness of PCR-restriction fragment length polymorphism typing of the coagulase gene to discriminate arbekacin-resistant methicillin resistant *Staphylococcus aureus* strains. *J Clin Microbiol*, 45, 607-609, 2007.
10. Strommenger B, Bräulke C, Heuck D, Schmidt C, Pasemann B, Nubel U, Witte W: *spa* Typing of *Staphylococcus aureus* as a frontline tool in epidemiological typing. *J Clin Microbiol*, 46, 574-581, 2008.
11. Enright MC, Day NP, Davies CE, Peacock SJ, Spratt BG: Multilocus sequence typing for characterization of methicillin-resistant and methicillin-susceptible clones of *Staphylococcus aureus*. *J Clin Microbiol*, 38, 1008-1015, 2000.
12. Melles DC, van Leeuwen WB, Snijders SV, Horst-Kreft D, Peeters JK, Verbrugh HA, van Belkum A: Comparison of multilocus sequence typing (MLST), pulsed-field gel electrophoresis (PFGE), and amplified fragment length polymorphism (AFLP) for genetic typing of *Staphylococcus aureus*. *J Microbiol Methods*, 69, 371-375, 2007.
13. Zadoks R, van Leeuwen W, Barkema H, Sampimon O, Verbrugh H, Schukken YH, van Belkum A: Application of pulsed-field gel electrophoresis and binary typing as tools in veterinary clinical microbiology and molecular epidemiologic analysis of bovine and human *Staphylococcus aureus* isolates. *J Clin Microbiol*, 38, 1931-1939, 2000.
14. Zadoks RN, van Leeuwen WB, Kreft D, Fox LK, Barkema HW, Schukken YH, van Belkum A: Comparison of *Staphylococcus aureus* isolates from bovine and human skin, milking equipment, and bovine milk by phage typing, pulsed-field gel electrophoresis, and binary typing. *J Clin Microbiol*, 40, 3894-3902, 2002.
15. Enright MC, Day NP, Davies CE, Peacock SJ, Spratt BG: Multilocus sequence typing for characterization of methicillin-resistant and methicillin-susceptible clones of *Staphylococcus aureus*. *J Clin Microbiol*, 38, 1008-1015, 2000.
16. Kapur V, Sischo WM, Greer RS, Whittam TS, Musser JM: Molecular population genetic analysis of *Staphylococcus aureus* recovered from cows. *J Clin Microbiol*, 33, 376-380, 1995.
17. Ikawaty R, Brouwer EC, Jansen MD, van Duiker E, Mevius D, Verhoef J, Fluit AC: Characterization of Dutch *Staphylococcus aureus* from bovine mastitis using a multiple locus variable number tandem repeat analysis. *Vet Microbiol*, 136 (3-4): 277-284, 2006.
18. Dastmalchi Saei H, Ahmadi M, Mardani K, Batavani RA: Molecular typing of *Staphylococcus aureus* isolated from bovine mastitis based on polymorphism of the coagulase gene in the north west of Iran. *Vet Microbiol*, 137, 202-206, 2009.
19. Brakstad OG, Aasbakk K, Maeland J: A detection of *Staphylococcus aureus* by polymerase chain reaction amplification of the *nuc* gene. *J Clin Microbiol*, 30, 1654-1660, 1992.
20. Wichelhaus TA, Hunfeld KP, Boddingtonhaus B, Kraiczky P, Schafer V, Brade V: Rapid molecular typing of methicillin-resistant *Staphylococcus aureus* by PCR-RFLP. *Infect Control Hosp Epidemiol*, 22, 294-298, 2001.
21. Dastmalchi Saei H, Ahmadi M: Discrimination of *Staphylococcus aureus* isolates on the basis of gene coding protein A using PCR-restriction enzyme analysis, 21 (5): 645-652, 2010.
22. Sindhu N, Sharma A, Jain VK: Coagulase gene based molecular detection of *staphylococcus aureus* directly from mastitic milk samples of murrah buffalo. *Buffalo Bulletin*, 29 (1): 52-59, 2010.
23. El-Huneidi W, Bdour S, Mahasneh: A Detection of enterotoxin genes *seg*, *seh*, *sei*, and *sej* and of a novel *aroA* genotype in Jordanian clinical isolates of *Staphylococcus aureus*. *Diagn Microbiol Infect Dis*, 56, 127-132, 2006.
24. Koreen L, Ramaswamy SV, Graviss EA, Naidich S, Musser JM, Kreiswirth BN: *spa* typing method for discriminating among *Staphylococcus aureus* isolates: Implications for use of a single marker to detect genetic micro- and macrovariation. *J Clin Microbiol*, 42, 792-799, 2004.

Comparison of Growth Curves of Broiler under Different Stocking Densities by Gompertz Model

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Summary

The knowledge of the growth curve in poultry science is very useful for describing growth kinetics and setting commercial management procedures. The objective of this research was to fit the Gompertz growth curve from hatching weights to 42 d-old weights of broilers in 3 stocking density groups. A total of 284 Ross 308 broilers randomly divided into 3 stocking density groups (9, 13 and 17 birds/m²) in this experiment. All birds were weighted weekly. The asymptotic weight (A) of broiler chickens for 9, 13 and 17 bird/m² stocking density groups were 4198.46, 3807.45 and 3999.92 g, respectively (P<0.05). The growth rates (K) of broiler chickens for 9, 13 and 17 bird/m² stocking density groups were 0.055, 0.058 and 0.052, respectively (P<0.01). The coefficient of determination for all stocking density groups were 0.998, 0.997 and 0.996, respectively (P<0.05). Moreover, the mean square error (MSE) value was lowest for 9 bird/m² group (P<0.05). The current study suggested that stocking density of 9 bird/m² was better for the broiler growth of the different stocking densities on the base of mature live weight (A), coefficient of determination (R²) and mean square error (MSE).

Keywords: Growth Curves, Gompertz model, Stocking Densities, Broiler

Farklı Yerleşim Sıklıklarındaki Etlik Piliçlerin Büyüme Eğrilerinin Gompertz Modeli İle Karşılaştırılması

Özet

Kanatlı biliminde büyüme eğrisinin bilinmesi büyümenin tarifinde ve yetiştirme tekniklerinin uygulanmasında çok önemlidir. Bu araştırmanın amacı kuluçkadan 42. gün ağırlığına kadar üç yerleşim sıklığında yetiştirilen etlik piliçlerden elde edilen canlı ağırlıkları, Gompertz büyüme eğrisi modeli ile tahmin etmektir. Denemede 3 yerleşim sıklığında (9, 13 ve 17 etlik piliç/m²) tesadüfen dağıtılan toplam 284 adet Ross 308 etlik piliç kullanılmıştır. Bütün hayvanlar haftalık olarak tartılmıştır. 9, 13 ve 17 etlik piliç/m² yerleşim sıklığında etlik piliçlerin, asimptotik ağırlıkları (A) sırasıyla; 4198.46, 3807.45 ve 3999.92 g olarak (P<0.05); büyüme oranı (K) sırasıyla 0.055, 0.058 ve 0.052 olarak (P<0.01) tahmin edilmiştir. Tüm yerleşim sıklıklarında belirtme katsayısı değeri (R²) sırasıyla 0.998, 997 ve 0.996 olarak bulunmuştur (P<0.05). Ek olarak hata kareler ortalaması (MSE) yerleşim sıklığı 9/m² olan grupta en düşüktür (P<0.05). Bu çalışmanın sonucunda tahmini ergin canlı ağırlığı (A), belirtme katsayısı değeri (R²) ve hata kareler ortalamasına (MSE) göre 9 etlik piliç/m² olan yerleşim sıklığında etlik piliçlerin büyümelerinin daha iyi olduğu söylenebilir.

Anahtar sözcükler: Büyüme eğrisi, Gompertz model, Yerleşim sıklığı, Etlik piliç

INTRODUCTION

Poultry industries face various decisions in the production cycle that include nutrient and mineral supply to birds, cost and type of feed, status of bird health, welfare and environmental issues that affect the profitability of

operation^[1]. The stocking densities in broilers vary widely by countries, husbandry systems and final body weights^[2]. Although the use of high stocking densities can diminish individual growth^[3-6], increase in total production meat



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per unit of floor surface, which results in higher profit. Thus, because of the economic benefits, producers have reluctant to decrease stocking densities^[7,8].

Although there have been various definitions of growth given by different biologists, for the purposes of quantitative analysis, growth was defined as the process of an animal gaining weight with time until it reaches maturity^[9]. Growth functions have been shown to be valuable tools for analyzing growth responses to genetic selection, environmental change^[10] and estimation of daily nutrient requirements for growth^[11]. An appropriate growth functions provide a good way of summarizing the information contained in such data into a few parameters that can be interpreted biologically and physically^[11,12]. Thus, the aim of the present study was to investigate the effect of stocking density on the growth curve of broiler chickens according to Gompertz growth model.

MATERIAL and METHODS

A total of 284 chicks (Ross 308) were selected from a commercial flock. Chickens were wing-tagged at 1 d of age and individual body weights were recorded weekly at the end of each one-week period. The chicks were raised in deep litter system and were subjected to the same management, hygienic and climatic conditions. Chickens were randomly placed into the floor systems in 3 stocking density groups with 3 repetitions for each group. The three stocking densities were 9 (density 1, 66 birds), 13 (density 2, 97 birds), and 17 (density 3, 121 birds), birds per m² and were raised to 42 d of age. The space for density 1, density 2 and density 3 were 2.77 m²; 2.76 m² and 2.82 m² respectively for per replicate.

Chickens were fed a 4-phase feeding program with starter fed to first 11 d (23% CP and 3.050 kcal/kg of ME), grower fed from 12 to 21 d (22% CP and 3.100 kcal/kg of ME), finisher fed from 22 to 35 d (20% CP and 3.200 kcal/kg of ME) and a withdrawal feed from 36 to 42 d (18% CP and 3.200 kcal/kg of ME). Diets were provided *ad libitum* and the birds had free access to water. Birds in all 3 groups were allocated with equal space of feeders and drinkers.

The birds were raised according to a typical commercial management program. The photoperiod was 24 h/day.

Temperature started at 32.0°C and was gradually reduced 1°C degrees every day until 22.0°C was attained, after which temperature remained constant.

The widely used nonlinear growth model, Gompertz function was applied to estimate the mean age-live weight relationship^[13]. The mathematical relation of this model was as follows:

$$W = A \cdot \exp(-B \cdot \exp(-Kt))$$

Where;

(W) is the body weight (g) at age (t); (A) is the asymptotic weight or maximum growth response (g); (B) is the initial weight; (K) is the growth rate; (t) is the age in days.

Statistical Analysis

The Gompertz nonlinear regression model^[14] was employed using the SAS statistical package program^[15]. The results obtained with Gompertz models was evaluated in an Excel spreadsheet. Kolmogorov-Smirnov test was performed to check the assumption of normality for Gompertz models coefficients. However, the estimated coefficients were non homogeneously distributed (P<0.05). Therefore, the coefficient differences were determined by the Kruskal Wallis H test. Dunn's multiple comparison tests was also used to compare groups.

RESULTS

The estimated parameter values by using the Gompertz growth function for live weight of broiler chickens for different stocking densities are given in *Table 1* and growth curves in *Fig 1*.

Stocking density significantly affected the asymptotic weight (P<0.05), initial weight (P<0.01), growth rate (P<0.01), determination of coefficients (P<0.05) and the mean square error (P<0.01).

DISCUSSION

In the current study, predicted asymptotic weights (A) were 4198.455, 3807.447 and 3999.922 g for stocking densities of 9 bird/m², 13 bird/m² and 17 bird/m²,

Table 1. Effects of stocking density on growth parameters estimated by Gompertz model

Tablo 1. Gompertz modelle tahmin edilen büyüme parametreleri üzerine yerleşim sıklığının etkisi

Stocking Density (Bird/m ²)	A	B	K	R ²	MSE
9	4198.46±442.73 ^a	4.83±0.342 ^a	0.055±0.006 ^b	0.998 ^a	2110 ^a
13	3807.45±436.49 ^b	4.78±0.387 ^a	0.058±0.007 ^a	0.997 ^{ab}	2846 ^b
17	3999.92±585.65 ^{ab}	4.56±0.387 ^b	0.052±0.008 ^c	0.996 ^b	2950 ^b
P	<0.019	<0.001	<0.001	<0.011	<0.012

(A) The asymptotic or mature weight (g); (B) the initial weight; (K) the growth rate; (R²) determination of coefficients; (MSE) the mean square error

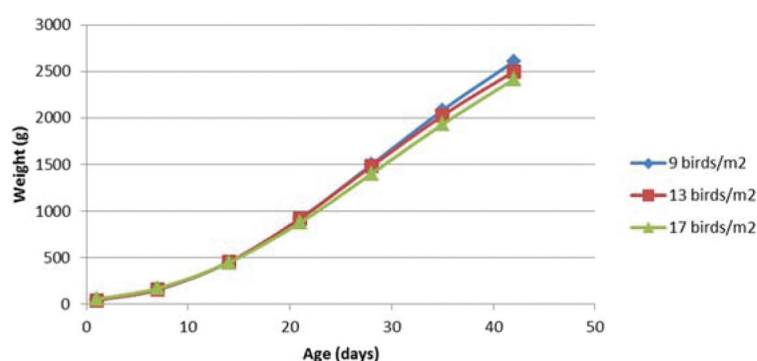


Fig 1. Estimated growth curves of Ross 308 chickens in different stocking density groups (1 to 42 d of age)

Şekil 1. Farklı yerleşim sıklığında 1-42. gün yaşa kadar Ross 308 etlik piliçlerin tahmin edilen büyüme eğrisi

respectively. Differences between stocking densities were observed in the Gompertz function parameters ($P < 0.05$). Based on calculated A value, asymptotic weight for stocking densities is arranged in descending order as 9 bird/m², 17 bird/m² and 13 bird/m². Although Santos et al.^[16] found similar asymptotic weight in their study (4136-4320 g). However, other studies usually reported lower asymptotic weight (Roush et al.^[17], 2936 g; Mignon-Grasteau et al.^[18], 3472 g, and Norris et al.^[19]; 2691-2819 g). Overall 9 bird/m² group had higher estimates for asymptotic weight. Lower stocking density may contribute to animal welfare, reduce stress and consequently resulted in better performance.

The estimation of the initial weight (B) was lower in the stocking density of 17 bird/m², while stocking density of 9 bird/m² showed higher values. The growth rates (K) were ranged from 0.052 to 0.058, showing the early growth rates for chicks in stocking density of 13 bird/m² group compared with others. Different ranges for (K) values had been previously reported. Lower values were reported by Yakupoglu and Atil^[20], Mignon-Grasteau et al.^[18], and Norris et al.^[19]. Similar values for the growth rate K were estimated by Marcato et al.^[21]. However, Santos et al.^[16] and Golomytis et al.^[22] reported higher (K) values when compared with the present study. The higher estimated growth rate values (K) suggested that broiler chickens in stocking density of 13 bird/m² group mature earlier than other chickens. It can be expected that individuals with lower K values would reach to the asymptotic weight (A) later than individuals with higher K values^[23,24].

The determination of coefficients (R^2) of stocking density groups were quite high indicating excellent fit of the data. Differences were observed for R^2 values among the various density groups. All densities have considerably high R^2 values. R^2 values were highest for birds in 9 bird/m² stocking density group ($R^2=0.998$), intermediate for birds in 13 bird/m² stocking density group ($R^2=0.997$) and lowest for birds in 17 bird/m² stocking density group ($R^2=0.996$) ($P < 0.05$). These findings are in agreement with the previous reports^[11,16,17]. Grasteau et al.^[18] have calculated the R^2 values of 0.980 through Gompertz models for broilers. In the current study, the mean square error (MSE) values ranged from 2110 to 2950. The stocking density 9 bird/m² group ranked the highest due to the lowest MSE value. The

MSE values in the current study suggested the stocking density 9 bird/m² depict better assessment for the growth of the stocking densities. Many authors^[11,16,17] found similar results using the models outlined in this study.

Body weight and growth rates are economically important features for broiler productions. The present study provides information about the effect of stocking density on some growth function parameters in broilers. In this study, the stocking density of 9 bird/m² seemed to be appropriate for describing the broiler growth of the stocking densities according to the asymptotic or mature weight, determination of coefficients and the mean square error values.

REFERENCES

1. Durmani Kuhl H, Porter T, López S, Kebreab E, Strathe AB, Dumas A, Dijkstra J, France J: A review of mathematical functions for the analysis of growth in poultry. *World Poultry Sci J*, 66 (2): 227-240, 2010.
2. Sarica M, Erensayin C: Etlik piliç yetiştiriciliği. In, Türkoğlu M, Sarica M (Eds): Tavukçuluk Bilimi. s.239-263, Bey Ofset Matmacılık, Ankara, 2009.
3. Şekeroğlu A, Berberoğlu E, Demir E, Sarica M: Etlik piliçlerde yerleşim sıklığının performansa ve atlık nem düzeyine etkisi. 1. Doğu Anadolu Kanatlı Sempozyumu, 21-24 Mayıs, Van, s.11-16, 2001.
4. El-Deek AA, Al-Harathi MA: Responses of modern broiler chicks to stocking density, green tea, commercial multi enzymes and their interactions on productive performance, carcass characteristics, liver composition and plasma constituents. *Int J Poult Sci*, 3 (10): 635-645, 2004.
5. Türkyılmaz MK: The effect of stocking density on stress reaction in broiler chickens during summer. *Turk J Vet Anim Sci*, 32 (1): 31-36, 2008.
6. Skomrucha I, Muchacka R, Osnówka-Czajka SE, Herbul E: Response of broiler chickens from three genetic groups to different stocking densities. *Ann Anim Sci*, 9 (2): 175-184, 2009.
7. Shanawany MM: Broiler performance under high stocking densities. *Br Poult Sci*, 29 (1): 43-52, 1988.
8. Feddes JJR, Emmanuel EJ, Zuidhof MJ: Broiler performance, body weight variance, feed and water intake, and carcass quality at different stocking densities. *Poult Sci*, 81 (6): 774-779, 2002.
9. Porter T, Kebreab E, Darmani Kuhl H, Lopez S, Strathe AB, France J: Flexible alternatives to the Gompertz equation for describing growth with age in turkey hens. *Poult Sci*, 89 (2): 371-378, 2010.
10. Goto T, Goto N, Shiraishi J, Bungo T, Tsudzuki M: Comparison of growth curves of four breeds of Japanese native chicken, onaga-dori, tosa-jidori, ukokkei and hinai-dori. *J Anim Vet Adv*, 9 (9): 1362-1365, 2010.
11. Ahmadi H, Mottaghtalab M: Hyperbolic models as a new powerful tool to describe broiler growth kinetics. *Poult Sci*, 86 (11): 2461-2465, 2007.

- 12. Fitzhugh HA:** Analysis of growth curves and strategies for altering their shape, *J Anim Sci*, 42 (4): 1036-1051, 1976.
- 13. Mignon-Grasteau S, Beaumont C, Le Bihan-Duval E, Poivey JP, Rochambeau de H, Ricard FH:** Genetic parameters of growth curve parameters in male and female chickens. *Br Poult Sci*, 40 (1): 44-51, 1999.
- 14. Rogers SR, Pesti GM, Marks HL:** Comparison of three nonlinear regression models for describing broiler growth curves. *Growth*, 51 (2): 229-239, 1987.
- 15. SAS:** Version 8.01 SAS Institute Inc., Cary, NC. 1999.
- 16. Santos AL, Sakomura NK, Freitas ER, Fortes CMS, Carrilho ENVM:** Comparison of free range broiler chicken strains raised in confined or semi-confined systems. *Braz J Poultry Sci*, 7 (2): 85-92, 2005.
- 17. Roush WB, Dozier WA, Branton SL:** Comparison of Gompertz and neural networks models of broiler growth. *Poult Sci*, 85 (4): 794-797, 2006.
- 18. Mignon-Grasteau S, Piles M, Varona L, de Rochambeau H, Poivey JP, Blasco A, Beaumont C:** Genetic analysis of growth curve parameters for male and female chickens resulting from selection on shape of growth curve. *J Anim Sci*, 78 (10): 2515-2524, 2000.
- 19. Norris D, Ngambi JW, Benyi K, Makgahlela ML, Shimelis HA, Nesamvuni EA:** Analysis of growth curves of indigenous male Venda and Naked Neck chickens. *S Afr J Anim Sci*, 37 (1): 21-26, 2007.
- 20. Yakupoglu C, Atil H:** Comparison of growth curve models on broilers growth curve 1: parameters estimation. *Online J Biol Sci*, 1 (7): 680-681, 2001.
- 21. Marcato SM, Sakomura NK, Kawauchi IM, Barbosa NAA, Freitas EC:** Growth of body parts of two broiler chicken strains. *EPC2006 XII European Poultry Conference, 10-14 September, Verona - Italy*, <http://www.cabi.org/animalscience/Uploads/File/AnimalScience/additionalFiles/WPSAVerona/10522.pdf>, 2006.
- 22. Goliomytis M, Panopoulou E, Rogdakis E:** Growth curves for body weight and major component parts, feed consumption, and mortality of male broiler chickens raised to maturity. *Poult Sci*, 82 (7): 1061-1068, 2003.
- 23. Taylor CS, Fitzhugh HA:** Genetic relationships between mature weight and time taken to mature within a breed. *J Anim Sci*, 33 (4): 726-731, 1971.
- 24. Brown JE, Fitzhugh HA, Cartwright TC:** A comparison of nonlinear models for describing weight age relationships in cattle. *J Anim Sci*, 42 (4): 810-818, 1976.

Effects of Dietary Inclusion of Plant Extract Mixture and Copper into Layer Diets on Egg Yield and Quality, Yolk Cholesterol and Fatty Acid Composition ^[1]

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Summary

This study was conducted to determine the effects of plant extract mixture (PEM) inclusion at different levels and copper into diets of hens on performance, egg quality traits, yolk and serum cholesterol content and yolk fatty acid composition. A total of 192 Lohmann white layers, 38 wks of age, were used in this study. The experiment was carried out on a control and 7 treatments groups (basal diet plus 200 mg/kg copper (CuSO₄·5H₂O), 500 mg/kg PEM, 500 mg/kg PEM + 200 mg/kg copper, 750 mg/kg PEM, 750 mg/kg PEM + 200 mg/kg copper, 1000 mg/kg PEM + 200 mg/kg copper). PEM, copper and PEM plus copper combinations did not affect performance parameters. Increasing level of PEM increased quadratically shell stiffness and linearly shell thickness, but did not affect other egg traits. Treatments had no a significant effect on cholesterol and triglycerid concentrations of egg and serum. While supplemented PEM and copper did not affect fatty acid level of yolk, PEM plus copper combination increased oleic acid and total MUFA levels. Results obtained from present study showed that supplementation of a mixed herbal product containing *Origanum vulgare* (dried leaf), *Thymus vulgaris* (dried leaf), thyme oil, origanum oil, garlic oil, anise oil and fennel oil to diets of laying hens can be beneficial to improve egg quality traits especially such as shell stiffness and thickness.

Keywords: Plant extract mixture, Copper, Performance, Egg quality, Cholesterol, Fatty acid composition of yolk

Yumurtacı Tavuk Rasyonlarına Farklı Seviyelerde Bitki Ekstraktı ve Bakır İlavesinin Yumurta Verimi ve Kalitesi, Yumurta Sarısı Kolesterolü ve Yağ Asidi Kompozisyonuna Etkileri

Özet

Bu çalışma, yumurtacı tavuk rasyonlarına ilave edilen farklı seviyelerde bitki ekstrakt karışımının ve bakırın performans, yumurta kalitesi, yumurta sarısı ve serum kolesterol içeriği ile yumurta sarısı yağ asidi kompozisyonu üzerine etkisini belirlemek amacıyla yürütülmüştür. Araştırmada; 38 haftalık yaşta 192 adet Lohmann beyaz yumurtacı tavuk kullanılmıştır. Deneme, bir kontrol ve 7 muamele (200 mg/kg bakır, 500 mg/kg bitki ekstrakt karışımı, 500 mg/kg bitki ekstrakt karışımı + 200 mg/kg bakır, 750 mg/kg bitki ekstrakt karışımı, 750 mg/kg bitki ekstrakt karışımı + 200 mg/kg bakır, 1000 mg/kg bitki ekstrakt karışımı, 1000 mg/kg bitki ekstrakt karışımı + 200 mg/kg bakır) grubundan oluşmuştur. Bitki ekstrakt karışımı, bakır ve bakır ile bitki ekstrakt karışımının performans parametrelerini etkilemediği tespit edilmiştir. Bitki ekstrakt karışımının artan seviyesi kırılma mukavemetini ve kabuk kalınlığını artırmaya rağmen diğer yumurta kalite kriterlerin de önemli bir etkiye sahip olmadığı saptanmıştır. Muamele grupları arasında yumurta sarısı kolesterol ve trigliserid oranı ile serum kolesterol ve trigliserid oranları bakımından önemli bir farklılığın olmadığı tespit edilmiştir. Bitki ekstrakt karışımı ile bakır kombinasyonu yumurta sarısında oleik asit ve toplam MUFA seviyesini artırırken diğer uygulamaların yumurta sarısı yağ asidi oranı üzerine etkisinin olmadığı belirlenmiştir. Sonuç olarak; yumurtacı tavuk rasyonlarına kekik, kekik otu, kekik yağı, sarımsak yağı, anason ve rezene yağından oluşan bitki ekstrakt karışımı ilavesinin yumurta kalite kriterlerini özellikle kabuk kalınlığı ve kırılma mukavemetini artırmada faydalı olabileceği söylenebilir.

Anahtar sözcükler: Bitki ekstrakt karışımı, Bakır, Performans, Yumurta kalitesi, Kolesterol, Yumurta sarısı yağ asit kompozisyonu

INTRODUCTION

It is known that a raised concentration of serum LDL-cholesterol increases the risk of coronary heart disease^[1]. It is also known that the intake of egg cholesterol contributes to an elevation of the level of serum is considered relevant to find ways to produce eggs low cholesterol^[2,3]. Nutritionist have been able to alter the diet of laying hens so that their eggs contain less cholesterol than the standard eggs. Results obtained from different studies have successfully demonstrated that natural products^[4] and copper^[5] could offer solutions to the health problem and cholesterol reduction. Natural products, essential extracts or oil derived from herbs and spices, exhibit hypocholesterolemic effect^[6]. The hypocholesterolemic effect of essential oils has been reported for chickens^[6-9]. It was reported that the pure components of essential oils inhibit hepatic 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase activity which is a key regulatory enzyme in cholesterol synthesis^[10].

Copper (Cu) is an essential mineral which serves as a cofactor in many enzyme system²¹. Research evidences demonstrated that pharmacological levels of Cu (>250 mg/kg diet) caused changes in 17 beta-estradiol and enzymes involved in carbohydrate, lipid and amino acid metabolism in mature laying hens and suggested that copper supplements can affect reproductive physiology and lipid metabolism^[11-13]. Kim et al.^[14] indicated that reduced glutathione by addition of copper may play a major role in cholesterol homeostasis. Also, Iduwu et al.^[15] reported that copper regulates cholesterol biosynthesis by reducing hepatic glutathione concentrations which is known to regulate cholesterol biosynthesis through the stimulation of the enzyme 3-hydroxy-3-methyl glutaryl coenzyme A (HMG-CoA) reductase, a key regulatory enzyme in cholesterol synthesis. Pesti and Bakalli^[13] obtained a reduction in egg yolk cholesterol concentration and an increase in egg production when 125 and 250 mg/kg dietary levels of Cu were fed on White Leghorn hens. Skrivan et al.^[16] suggested that copper supplementation at 200 mg/kg level in broiler diet had beneficial effects on growth performance. The maximum dietary tolerable level of copper for poultry was set at 300 mg kg⁻¹ according to NRC^[17]. Chiou et al.^[18] reported that Layer diets containing 200 mg Cu per kg gave a positive response on the performance, liver function and enzyme activities but high dietary copper dosage (400, 600 or 800 mg kg⁻¹) gave a progressively negative response.

The objective of this study was to examine the dietary effect of the single or combination feeding of plant extracts mixture including *Origanum vulgare* (dried leaf), *Thymus vulgaris* (dried leaf), thyme oil, origanum oil, garlic oil, anise oil and fennel oil at different levels and copper on laying performance, egg quality, yolk fatty acid composition, yolk and serum cholesterol and triglyceride content.

MATERIAL and METHODS

This experiment was conducted by the researchers based on protocols by Atatürk University Ethical Commission Report (No: 24.04.2009/5/54).

A total of 192 Lohmann white layers, 38 weeks of age, were randomly allocated eight groups, each formed 6 replicate cages as subgroups, comprising of four hens. Experimental diets were (1) a standard commercial layer diet (Control), (2) a diet including 200 mg/kg copper (CuSO₄·5H₂O), (3) a diet including 500 mg/kg plant extracts mixture, (4) a diet including 500 mg/kg plant extracts mixture + 200 mg/kg copper, (5) a diet including 750 mg/kg plant extracts, (6) a diet including 750 mg/kg plant extracts mixture + 200 mg/kg copper, (7) a diet including 1.000 mg/kg plant extracts and (8) a diet including 1.000 mg/kg plant extracts mixture + 200 mg/kg copper for 14 weeks including adaptation period of 2 weeks. Diets were formulated according to NRC^[17] and chemically analyzed by the AOAC^[19] (Table 1). A commercial blend of PEM Fitococci[®] contained 463 g/kg active ingredients *, basically including *Origanum vulgare* (dried leaf), *Thymus vulgaris* (dried leaf), thyme oil, origanum oil, garlic oil, anise oil and fennel oil. PEM and Cupric sulfate (CuSO₄·5H₂O) as a copper source were provided from Farmavet International Inc., Kocaeli, 41400, Turkey. During the experimental period, hens were fed ad libitum once daily at 08:30 h and water was available all the times. Hens were subjected to a 16L:7D cycle.

Feed intake and egg production were recorded daily, egg weight was measured bi-weekly, and body weight was measured at the end of experimental period. Feed conversion ratio (FCR) was expressed as kilogram of feed consumed per kilogram of egg produced. To evaluate egg quality parameters 12 egg samples were randomly picked up from each experimental group every month. Before determination of egg quality parameters such as shape index, shell strength, shell thickness, albumen index, yolk index, yolk color (Yolk Colour Fan, the CIE standard colorimetric system, F. Hoffman-La Roche Ltd., Basel, Switzerland), and Haugh unit, samples were stored for 24 h in room temperature. They were assessed according to the method of Kaya et al.^[20].

At the end of the experiment period, blood and egg samples were taken from the each replicate in order to determine the ratio of triglyceride and cholesterol. Triglyceride and cholesterol ratios of blood and egg samples were assessed according to the method of Kaya and Macit^[21].

Sample of 5 eggs was randomly collected from each

* Active ingredients given by Farmavet International Inc: 1,8-Cineole (0.24%), Allilene (0.24%), Allilene (0.12%), Alpha-Pinene (0.12%), Alpha Terpineol (0.70%), Borneol (0.18%), Caffeic-Acid (2.28%), Camphene (0.08%), Carvacrol (4.48%), Eugenol (0.12%), Geraniol (1.04%), Limonene (0.56%), Linalool (0.96%), Myrcene (0.18%), P-Cymene (2.38%), Phenol (0.86%), Polyphenol (6.00%), Tannin (12.9%), Rosmarinic-Acid (7.60%), Terpinen-4-Ol (0.06%), Ursolic Acid (1.92%), Tymol (3.26%)

experimental group for fatty acid (FA) analysis at the end of the trial. The FA profile was determined with gas chromatograph (HP Agilent Technologies 6890N, Inc. Headquarters Santa Clara, US). Method identified by Yüksel et al.^[22] was used to determine FA levels of yolk samples.

The experimental data were statistically analysed to the general linear models procedure of SPSS^[23]. Polynomial contrast was constructed to determine the effect of the

feeding level of PEM in the diets. The effects of the dietary treatments on response variables were declared to be significant at $P < .05$.

RESULTS

The effects of single feeding of PEM or Cu and combination of copper and PEM on the performance traits are presented in Table 2. Dietary additives did not have significant effect on final body weight and body weight change of hens. Similarly, feeding as the single or combination of plant extract mixture at different levels and copper did not affect the egg weight, egg production, feed intake, feed conversion ratio and broken egg rate.

The egg quality traits of hens fed a diet supplemented with PEM and Cu or combination of copper and PEM are shown in Table 3. Shape index, shell weight, yolk index, albumen index and haugh unit were not affected by addition of PEM ($P > .05$). But PEM supplementation affected shell stiffness and shell thickness ($P < .05$). Shell stiffness was quadratically increased by increasing level of PEM. On the other hand, a significant linear increase was found in shell thickness when PEM at 500, 750 and 1000 mg/kg levels was added to layer diets. The supplementation of copper had no significant effect on egg quality traits except for yolk color. Egg quality traits were not influenced by feeding of combination of PEM and copper.

The effect of inclusion of PEM at 500, 750 and 1000 mg/kg levels into diet on ratios of cholesterol and triglyceride as a percentage of total lipids in serum and egg yolk is presented in Table 4. Cholesterol and triglyceride concentrations of yolk and serum were not affected by dietary treatments.

Table 5 shows the dietary effects of PEM, 200 ppm

Table 1. Ingredients and chemical composition of the experimental basal diet

Tablo 1. Bazal rasyonun bileşimi ve kimyasal kompozisyonu

Ingredients	(%)	Chemical Composition (Analyzed on Dry Matter Basis)	
Corn	52.81	Dry matter (%)	89.47
Soybean meal	18.13	Crude protein (%)	16.50
Wheat	6.00	Crude fiber (%)	4.49
Full fat Soybean oil	1.65	Crude ash (%)	11.70
Sunflower meal	7.50	Ether extract (%)	4.88
Corn gluten meal	2.04	Lysine (%)	0.7
Soybean oil	1.60	Methionine (%)	0.33
Ground limestone	6.82	Calcium (%)	3.4
Salt	0.30	Total phosphorus (%)	0.7
Di-calcium phosphate 18	2.65	ME [*] kcal/kg	2720
DL-Methionine 99%	0.15		
L-Lysine	0.10		
Vit-Min	0.25		

Each two kilogram of vitamin- mineral premix contained 12.000 IU of retinyl palmitate, 2.500 IU of cholecalciferol, 30 IU of DL- α -tocopheryl acetate, 34 mg of phylloquinone, 3 mg of thiamin, 6 mg of riboflavin, 30 mg of nicotin amid, 10 mg of Ca-D-pantothenate, 5 mg of pyridoxine, 15 mg of cobalamin, 1 mg of folic acid, 50 mg of D-Biotin, 300 mg of choline chloride, 50 mg of ascorbic acid, 80 mg of manganese, 60 mg of iron, 60 mg of zinc, 5 mg of copper, 2 mg of iodine, 5 mg of cobalt, 150 mg of selenium^{*} Metabolizable Energy Calculated

Table 2. Effects of plant extract mixture and copper on laying performance parameters

Tablo 2. Bitki ekstrakt karışımı ve bakırın performans parametreleri üzerine etkisi

PEM mg/kg	Cu ppm	IBW (g)	FBW (g)	BWC (g)	EW (g)	EP (%)	FI (g/d)	FCR ¹	DER (%)
0	0	1627.27	1760.00	132.83	67.54	89.02	131.85	2.20	0.03
	200	1560.83	1705.83	145.00	66.72	85.68	123.35	2.17	0.27
500	0	1525.63	1650.63	125.00	65.96	90.29	128.20	2.16	0.13
	200	1593.96	1668.96	75.00	65.89	90.03	130.12	2.20	0.17
750	0	1622.08	1723.33	101.25	67.64	87.54	128.44	2.19	0.20
	200	1576.04	1681.67	105.63	64.56	90.39	128.40	2.18	0.23
1000	0	1597.92	1698.13	100.21	66.22	90.78	127.87	2.14	0.07
	200	1587.71	1670.00	82.29	64.12	85.75	125.24	2.32	0.40
SEM		29.95	30.55	21.93	1.13	2.27	2.41	0.24	0.11
PEM		0.55	0.12	0.16	0.39	0.65	0.73	0.54	0.83
Cu		0.53	0.23	0.41	0.06	0.37	0.18	0.25	0.06
PEM*Cu		0.14	0.66	0.50	0.56	0.33	0.17	0.59	0.47

IBW= Initial body weight, FBW= Final body weight, BWC= Body weight change, EW= Egg weight, EP= Egg production rate, FI= Feed intake, FCR= Feed conversion ratio (kg feed:kg egg), DER= Damage egg rate, PEM= plant extracts mixture, SEM= Standard error of mean, Cu= Copper

Table 3. Effects of plant extract mixture and copper on egg quality parameters**Tablo 3.** Bitki ekstrakt karışımı ve bakırın yumurta kalite parametreleri üzerine etkisi

PEM mg/kg	0		500		750		1000		SEM	PEM / Polinomial	Cu	PEM*Cu
Cu mg/kg	0	200	0	200	0	200	0	200				
SI (%)	74.22	74.44	73.89	73.83	73.42	73.61	74.17	74.94	0.65	0.38	0.54	0.93
SS (kg/cm ²)	0.84	1.03	1.14	1.20	0.95	0.91	0.90	0.84	0.07	0.01/(Q)	0.44	0.26
ST (mm×10 ⁻²)	0.39	0.40	0.39	0.40	0.39	0.40	0.43	0.42	0.01	0.00/(L)	0.49	0.58
SW (g)	8.55	8.37	8.32	8.80	8.22	8.22	8.37	8.17	0.17	0.18	0.84	0.20
YC	8.83	8.39	8.44	8.11	8.33	7.50	8.17	7.78	0.23	2.02	0.00	0.70
YI (%)	42.71	42.44	43.09	43.74	43.15	43.58	43.20	42.50	0.54	0.35	0.95	0.58
AI (%)	8.32	8.44	8.44	8.90	8.85	7.93	8.20	8.86	0.39	0.87	0.76	0.20
HU	82.01	82.65	83.03	84.52	84.20	80.17	80.83	85.04	1.66	0.77	0.62	0.11

SI= Shape index, SS= Shell strenght, ST= Shell thickness, SW= Shell weight, YC= Yolk color, YI= Yolk index, AI= Albumen index, HU= Haugh unit, PEM= Plant extract mixture, SEM= Standart error mean, Cu= Copper, L= Linear effect of PEM supplementation; Q= Quadratic effect of PEM supplementation

Table 4. Effects of plant extract mixture on cholesterol and trigliserid level of egg and serum (%)**Tablo 4.** Bitki ekstrakt karışımı ve bakırın yumurta sarısı ve serum kolesterol ile trigliserid oranları üzerine etkisi (%)

PEM mg/kg		0		500		750		1000		SEM	PEM	Cu	PEM*Cu
Cu mg/kg		0	200	0	200	0	200	0	200				
Egg yolk	Chol	18.84	18.19	18.25	20.99	17.77	18.95	17.45	16.22	1.26	0.19	0.57	0.39
	Trig	60.54	56.00	56.33	60.35	58.52	61.33	61.29	59.03	1.72	0.57	1.0	0.06
Serum	Chol	24.35	23.52	23.33	22.83	23.33	23.12	23.34	23.00	0.65	0.54	0.32	0.97
	Trig	43.87	45.17	45.36	45.26	42.63	44.12	44.15	43.09	0.92	0.15	0.53	0.47

Chol= Cholesterol, Trig= Trigliserid, PEM= Plant extract mixture, SEM= Standart error mean, Cu= Copper

copper and combination of PEM and copper on fatty acid composition of yolk. While supplemented PEM and copper did not affect fatty acid level in yolk, PEM with copper combination increased oleic acid and total monounsaturated fatty acid (MUFA) level ($P < .05$).

DISCUSSION

Present study, significant differences were observed among the groups with respect to performance parameters at the end of the 14-week experimental period. A similar observation was reported by Bozkurt et al.^[9] who determined that body weight was not affected by addition of an essential oil combination to the layer diets. Arpasova et al.^[24] concluded that body weight was not influenced diet including 0.25 ml/kg thyme essential oil in the laying hens. But Çabuk et al.^[25] found that essential oils had the positive effects on body weight of laying hens. In accordance with the present findings, Orhan and Eren^[26] reported that the addition of essential oil mixture including *Origanum vulgare* (leaf), *Thymus vulgaris* (leaf), thyme oil, origanum oil, garlic oil, anise oil and fennel oil into laying hen diets had no significant effect on egg weight, egg production, feed conversion ratio, feed consumption. In agreement with the present study, Aksu and Bozkurt^[27] found that body weight, feed intake, and feed efficiency of broilers were not affected by the adding of essential oils at the level of

1.000 mg/kg. Similarly, Köksal and Küçükarsan^[28] reported that supplementation of essential oil into basal diet at level of 0.75 g/kg did not have significant effect on performance including body weight, body weight gain, feed intake and feed conversion ratio in broilers. Specific effects of the essential oils on laying performance have not received much attention because poultry may not acutely respond to flavor^[29]. In contrast to our results, a study has observed that the addition of essential oil mixture into a layer diet has positive effect on egg weight and egg production rate^[30]. Chandra et al.^[31] reported that the supplementation of dietary copper at different levels (50, 75, 100, 125, 150 and 175 mg Cu per kg diet) into layer diets had no significant influence on performance parameters. Kaya and Macit^[21] reported that egg weight, egg production and feed consumption decreased with 200 mg/kg Cu sulfate supplementation but feed conversion ratio and cracked egg were not affected. Pekel and Alp^[32] postulated that supplementation with 250 mg/kg of Cu sulfate improved egg production but decreased egg weight and feed intake.

Except for shell stiffness and shell thickness, none of the egg quality traits were affected by PEM. In contrast to our study, other researchers have observed that the addition of essential oil mixture to layer diet did not have significant effect on shell strenght^[9,26,30]. Similarly Nasiroleslami and Torki^[33] reported that adding 300 mg/kg fennel essential oil to laying hen diet improved shell thickness. In contrast

Table 5. Effects of plant extract mixture and Cu on fatty acid composition of egg yolk (%)**Tablo 5.** Yumurta tavuk rasyonlarına bitki ekstrakt karışımı ve bakır ilavesinin yumurta sarısı yağ asidi kompozisyonu üzerine etkisi (%)

PEM mg/kg	0		500		750		1000		SEM	PEM	Cu	PEM*Cu
Cu mg/kg	0	200	0	200	0	200	0	200				
C14:0	0.28	0.28	0.27	0.32	0.27	0.27	0.23	0.26	0.02	0.08	0.17	0.49
C14:1	0.01	0.01	0.02	0.03	0.01	0.02	0.02	0.02	0.01	0.28	0.22	0.89
C15:0	0.03	0.04	0.03	0.04	0.02	0.02	0.02	0.02	0.01	0.12	0.90	0.38
C16:0	25.93	25.78	25.45	26.58	26.10	26.04	25.19	25.93	0.45	0.66	0.20	0.41
C16:1	2.16	2.34	2.33	2.72	2.44	2.63	2.28	2.35	0.20	0.37	0.15	0.88
C17:0	0.18	0.17	0.18	0.17	0.16	0.15	0.18	0.16	0.01	0.367	0.10	0.98
C17:1	0.08	0.08	0.08	0.08	0.08	0.08	0.07	0.08	0.01	0.68	0.58	0.56
C18:0	9.63	9.57	9.33	9.11	8.95	8.80	9.16	9.58	0.30	0.13	0.99	0.71
C18:1t (n-9)	0.06	0.06	0.06	0.07	0.06	0.02	0.06	0.05	0.02	0.38	0.36	0.43
C18:1c (n-9)	37.95 ^b	38.27 ^{ab}	37.98 ^b	36.02 ^c	37.68 ^b	39.05 ^a	37.95 ^b	38.46 ^{ab}	0.55	0.07	0.88	0.03
C18:2t (n-6)	1.50	1.59	1.60	1.60	1.62	1.76	1.60	1.59	0.06	0.16	0.18	0.57
C18:2c (n-6)	16.97	16.57	17.21	17.79	17.25	16.02	17.48	16.54	0.60	0.50	0.25	0.46
C18.3 (n-6)	0.12	0.11	0.12	0.11	0.10	0.07	0.09	0.10	0.02	0.13	0.34	0.51
C18.3 (n-3)	0.40	0.42	0.44	0.48	0.45	0.39	0.45	0.40	0.03	0.45	0.60	0.33
C20:1	0.19	0.22	0.20	0.19	0.22	0.21	0.20	0.21	0.01	0.50	0.51	0.47
C20:2	0.19	0.20	0.21	0.21	0.21	0.19	0.19	0.20	0.01	0.61	0.87	0.65
C20:3 (n-6)	0.18	0.19	0.19	0.19	0.18	0.17	0.14	0.17	0.02	0.14	0.40	0.56
C20:4	1.89	1.90	1.90	1.84	1.80	1.79	1.94	1.85	0.05	0.13	0.28	0.74
C24:1	0.71	0.73	0.69	0.65	0.71	0.69	0.76	0.68	0.02	0.18	0.13	0.17
Σ SFA	36.05	35.83	35.26	36.21	35.51	35.26	34.78	35.96	0.47	0.56	0.22	0.29
Σ UFA	62.43	62.69	63.03	62.01	62.79	63.08	63.19	62.69	0.45	0.67	0.45	0.41
Σ MUFA	41.17 ^b	41.71 ^{ab}	41.36 ^b	39.77 ^c	41.19 ^b	42.70 ^a	41.30 ^b	41.83 ^{ab}	0.50	0.06	0.49	0.03
Σ PUFA	21.27	20.99	21.66	22.24	21.60	20.38	21.89	20.86	0.66	0.48	0.30	0.51
Σ (ω-6)	20.67	20.36	21.01	21.54	20.94	19.79	21.25	20.26	0.62	0.49	0.28	0.52
Σ (ω-3)	0.41	0.42	0.44	0.49	0.45	0.39	0.45	0.40	0.03	0.37	0.62	0.29

C14:0= Miristic Acid, C14:1= Miristoleic Acid, C16:0= Palmitic Acid, C16:1= Palmitoleic Acid, C17:0= Heptadecanoic Acid, C17:1= Heptadecenoic Acid, C18:0= Stearic Acid, C18:1t (n-9)= Elaidic Acid, C18:1c (n-9)= Oleic Acid, C18:2t (n-6)= Linoleic Acid, C18:2c(n-6)= Linoleic Acid, C18.3(n-6)= γ-Linolenic Acid, C18.3(n-3)= α-Linolenic Acid, C20:1= Gadoleic Acid, C20:2= Eikosadienoic Acid, C20:3 (n-6)= Eikosatrienoic Acid, C20:4= Arashidonic Acid, C24:1= Nervonic Acid, Σ SFA= Total saturated fatty acid, Σ UFA= Total unsaturated fatty acid, Σ MUFA= Total Monounsaturated Fatty Acid, Σ PUFA= Total unsaturated fatty acid, Σ (ω-6)= Total omega-6, Σ (ω-3)= Total Omega-3, PEM= Plant extract mixture, SEM= Standart error mean, Cu= Copper

to present study, there have been some researches who reported the unbeneficial effects of essential oil mixture on shell thickness in laying hens [9,30,34]. Information acquired from literature showed that herbal extracts used in poultry nutrition caused feed intake and feed efficiency improvement [35,36] because of increased production of digestive enzymes [37]. Increase in shell thickness may lead to higher nutrient retention and nutrient availability through the intestines during shell formation in this study. Yolk color was decreased by supplementation of 200 mg/kg copper to laying hen diets. In contrast to our results, Kaya and Macit [21] observed that yolk color was not affected by addition of 200 mg/kg copper to layer diets.

The effects of supplemental treatments on cholesterol and triglyceride ratios of yolk and serum were not significant in the present study. In accordance with the present findings,

some researchers who reported that cholesterol and triglyceride concentrations of serum were not affected by addition of 24 mg/kg essential oil mixture [9] or 300 mg essential oils of fennel per kg diet [33]. A different observation was reported by Bölükbaşı et al. [38] who determined that the addition of 200 ppm thyme essential oil to the layer diet resulted in remarkable reduction in serum cholesterol and triglyceride ratios. Bölükbaşı et al. [39] showed that adding essential oils of thyme (200 mg/kg) into layer diet decreased triglyceride ratios of yolk but not effected cholesterol ratios of yolk. The results of cholesterol and triglyceride ratios of egg and serum were supported by Kaya and Macit [21] who founded that the supplementation of 200 ppm copper did not have a significant effect on cholesterol and triglyceride ratios of egg and serum. On the contrary to the result of this experiment, in previous studies, egg yolk cholesterol in laying hens was decreased

by dietary Cu supplementation^[5,13,40,41]. Also Abaza et al.^[42] stated that Cu supplementation at 0 to 100 and 200 mg/kg levels in the laying quail diets decreased cholesterol and triglyceride concentrations of serum.

Inclusion of PEM with copper combination into the diets of laying hens increased oleic acid and total mono-unsaturated fatty acid (MUFA) level. The increase in MUFA observed in the eggs from layer fed the combination of PEM and copper diets was mainly contributed by oleic acid (C18:1c [n-9]). Recently, MUFA have been a subject of interest due to their possible hypolipidemic and anti-thrombotic effects^[43]. Ansari et al.^[44] reported that fatty acid level of yolk in layer was not effected by 250 ppm copper. Maurice and Lightsey^[45] postulated that adding of 250 ppm copper into layer diets did not effect fatty acid level of yolk except for arashidonic acid level.

The differences between present study and other experiments may be due to differences in the laying period, age and genotype of hen and the combination, and kind of used extract and levels of Cu and extract in the diets of laying hens. In previous years, effects of supplementation of PEM together with Cu as combination in laying hens diets have not been studied. Therefore there have not been yet any literature to compare the results related to performance and some egg quality traits containing cholesterol and fatty acid levels obtained from present study with findings reported by other researchers.

Results from present study showed that addition of plant extract mixture containing leaves of *Origanum vulgare* (leaf), *Thymus vulgaris* (leaf), thyme oil, origanum oil, garlic oil, anise oil and fennel oil into the laying hen diets can be beneficial to improve the shell stiffness and thickness from egg quality traits. In addition, feeding of laying hens with diet containing combination of plant extract mixture and copper increased MUFA level of yolk, but did not decrease the cholesterol level of egg yolk in laying hens.

In conclusion, combination of 750 mg/kg PEM and Cu may be used in the diets of 38 wks of age laying hens for 3-month experimental period, but more experiment on the supplementation of extracts and Cu combinations into diets of laying hens should be conducted by researchers related to this subject.

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REFERENCES

1. Gray J, Griffin B: Eggs and dietary cholesterol dispelling the myth. *Brit Nutr Foundation, Nutrition Bulletin*, 34, 66-70, 2009.
2. Donalc I: Eating eggs and nutrition. *World Poult*, 16, 24-25, 1999.
3. El Bagir NM, Hama AY, Hamed RM, Abd El Rahim AG, Baynen AC: Lipid composition of egg yolk and serum in laying hens fed diets containing blac cumin (*Nigella sativa*). *Int J Poult Sci*, 5, 574-578, 2006.
4. Mottaghitaleb M, Taraz Z: Garlic powder as blood serum and egg yolk cholesterol lowering agent. *J Poult Sci*, 41, 50-57, 2004.
5. Al Ankari A, Najib H, Al Hozab A: Yolk and serum cholesterol and production traits, as affected by incorporating a supraoptimal amount of copper in the diet of the leghorn hens. *Br Poult Sci*, 39 (3): 393-397, 1998.
6. Craig WJ: Health-promoting properties of common herbs. *Am J Clin Nutr*, 70 (suppl): 491-499, 1999.
7. Case GL, HE L, Mo H, Elson CE: Induction of geranyl pyrophosphate pyrophosphatase activity by cholesterol suppressive isoprenoids. *Lipids*, 30, 357-359, 1995.
8. Bölükbaşı ŞC, Ürüsan H, Erhan MK, Kızıltunç A: Effect of dietary supplementation with bergamot oil (*Citrus bergamia*) on performance and serum metabolic profile of hens, egg quality and yolk fatty acid composition during the late laying period. *Arch Geflügelkd*, 74, 172-177, 2010.
9. Bozkurt M, Tokuşoğlu Ö, Küçükylmaz K, Akşit H, Çabuk M, Çatlı AU, Seyrek K, Çınar M: Effects of dietary mannan oligosaccharide and herbal essential oil blend supplementation on performance and oxidative stability of eggs and liver in laying hens. *Ital J Anim Sci*, 11(e41): 223-229, 2012.
10. Crowell PL: Prevention and therapy of cancer by dietary mono-terpenes. *J Nutr*, 129, 775-778, 1999.
11. Pearce J, Jackson N, Stevenson MH: The effect of dietary concentration on copper sulphate on the laying domestic fowl: effects on some aspects of lipid, carbohydrate and amino acid metabolism. *Br Poult Sci*, 24, 337-348, 1983.
12. Bakalli RI, Pesti GM, Ragland WL, Konjufca W: Dietary copper in excess of nutritional requirements reduces plasma and breast muscle cholesterol in chickens. *Poult Sci*, 74, 360-365, 1995.
13. Pesti GM, Bakalli RI: Studies on the effect of feeding cupric sulfate pentahydrate to laying hens on egg cholesterol content. *Poult Sci*, 77, 1540-1545, 1998.
14. Kim S, Chao PY, Allen GDA: Inhibition of elevated hepatic glutathione abolishes copper deficiency cholesterolemia. *FASEB J*, 6, 2467-2471, 1992.
15. Idowu OMO, Kuye OA, Oladele-Ojo VO, Eruvbetine D: The effects of dietary intake and dietary concentration of organic and inorganic copper supplements on egg and plasma cholesterol levels. *Xth European Symposium on the Quality of Eggs and Egg Products Doorwerth, The Netherlands*, 23-26 May, pp.401-409, 2005.
16. Skrivan M, Skrivanova V, Marounek M, Tumova E, Wolf J: Influence of dietary fat source and copper supplementation on broiler performance, fatty acid profile of meat and depot fat, and on cholesterol content in meat. *Br Poult Sci*, 41, 608-614, 2000.
17. National Research Council: Nutrient Requirements of Poultry. 9th Revised ed., National Academy Press, Washington, DC, 1994.
18. Chiou PW, Chen KL, Yu B: Toxicity, tissue accumulation and residue in egg and excreta of copper in laying hens. *Anim Feed Sci Technol*, 67, 49-60, 1997.
19. AOAC: Official Methods of Analysis. Vol 1. 15th ed., Arlington, VA., 1990.
20. Kaya H, Çelebi Ş, Macit M, Geyikoğlu F: The effects of raw and physical processed common vetch seed (*Vicia sativa*) on laying performance, egg quality, metabolic parameters and liver histopathology of laying hens. *Asian-Aust J Anim Sci*, 24, 1425-1434, 2011.
21. Kaya H, Macit M: Effect of inclusion of garlic (*Allium sativum*) powder at different levels and copper into diets of hens on performance, egg quality traits and yolk cholesterol content. *Inter J Poult Sci*, 11, 114-119, 2012.
22. Yüksel S, Yanar M, Aksu MI, Kopuzlu S, Kaban G, Sezgin E, Oz F: Effects of different finishing systems on carcass traits, fatty acid composition, and beef quality characteristics of young Eastern Anatolian Red bulls. *Trop Anim Health Prod*, 44, 1521-1528, 2012.
23. SPSS: SPSS for Windows Release 10.0, SPSS Inc, 1999.
24. Arpasova H, Hascık P, Bujko J: Influence of plant essential oils on selected parameters of the performance of laying hens. *J Cent Eur Agric*, 11, 11-18, 2010.
25. Çabuk M, Bozkurt M, Alçiçek A, Çatlı AU, Başer KHC: Effect of a

dietary essential oil mixture on performance of laying hens in the summer season. *S Afr J Anim Sci*, 36, 215-221, 2006.

26. Orhan F, Eren M: Effect of herbal mixture supplementation to fish oiled layer diets on lipid oxidation of egg yolk, hen performance and egg quality. *Ankara Univ Vet Fak Derg*, 58, 33-39, 2011.

27. Aksu T, Bozkurt AS: Effect of dietary essential oils and/or humic acids on broiler performance, microbial population of intestinal content and antibody titres in the summer season. *Kafkas Univ Vet Fak Derg*, 15 (2): 185-190, 2009.

28. Köksal BH, Küçükersan MK: Effects of humate and vegetable extract mixture supplementation to diets on growth performance, some immunity and serum biochemistry parameters in broiler chickens. *Kafkas Univ Vet Fak Derg*, 18 (1): 103-108, 2012.

29. Al-Kassie GAM: Influence of two plant extracts derived from thyme and cinnamon on Broiler performance. *Pakistan Vet J*, 29 (4): 169-173, 2009.

30. Bozkurt M, Küçükyılmaz K, Pamukçu M, Çabuk M, Alçiçek A, Çatlı AU: Long-term effects of dietary supplementation with an essential oil mixture on the growth and laying performance of two layer strains. *Ital J Anim Sci*, 11(e5): 23-28, 2012.

31. Chandra D, Shrivastava HP, Mandal AB, Tyagi PK, Singh DP: Effect of feeding different levels of dietary copper on production performance, egg quality traits and egg yolk cholesterol content in CARI-Red layers. *Indian J Poult Sci*, 46, 79-82, 2011.

32. Pekel AY, Alp M: Effects of different dietary copper sources on laying hen performance and egg yolk cholesterol. *J Appl Poult Res*, 20, 506-513, 2011.

33. Nasiroleslami M, Torki M: Including essential oils of fennel (*Foeniculum Vulgare*) and ginger (*Zingiber Officinale*) to diet and evaluating performance of laying hens, white blood cell count and egg quality characteristics. *Adv Environ Biol*, 4, 341-345, 2010.

34. Varel VH: Livestock manure odor abatement with plant-derived oils and nitrogen conservation with urease inhibitors: A review. *J Anim Sci*, 80, 1-7, 2002.

35. Williams P, Losa R: The use of essential oils and their compounds in

poultry nutrition. *World Poult*, 17, 14-15, 2001.

36. Jamroz D, Wiertelcki T, Houszka M, Kamel C: Influence of diet type on the inclusion of plant origin active substances on morphological and histochemical characteristics of the stomach and jejunum walls in chicken. *J Anim Phys Anim Nut*, 90, 255-268, 2006.

37. Köksal BH, Küçükersan MK: Yumurta tavuğu rasyonlarına ilave edilen humat ve bitki ekstraktı karışımının performans ile bazı kan parametrelerine etkileri. *Ankara Üniv Vet Fak Derg*, 59, 121-128, 2012.

38. Bölükbaşı ŞC, Erhan MK, Kaynar Ö: The Effect of feeding thyme, sage and rosemary on laying hen performance, cholesterol and some proteins ratio of egg yolk and *Escherichia coli* count in feces. *Arch Geflügelkd*, 72, 231-237, 2008.

39. Bölükbaşı ŞC, Erhan MK, Kaynar Ö: Effect of dietary thyme oil on laying hens performance, cholesterol ratio of egg yolk and *Escherichia coli* concentration in feces. *3rd Joint Meeting of the Network of Universities and Research Institutions of Animal Science of the South Eastern European Countries, Thessaloniki 10-12 February*, 2007.

40. Balevi T, Coskun B: Effects of dietary copper on production and egg cholesterol content in laying hens. *Br Poult Sci*, 5, 530-534, 2004.

41. Idowu OMO, Laniyan TF, Kuye OA, Oladele-Ojo VO, Eruvbetine D: Effect of copper salts on performance, cholesterol, residues in liver, eggs and excreta of laying hens. *Arch Zootechn*, 55, 327-338, 2006.

42. Abaza IM, Ezzat W, Shoeib MS, El-Zaiat AA, Hassan II: Effects of copper sulfate on productive, reproductive performance and blood constituents of laying Japanese quail fed optimal and sub-optimal protein. *Inter J Poult Sci*, 8, 80-89, 2009.

43. Botsoglou NA, Yannakopoulos AL, Fletouris DJ, Tserveni-Goussi AC, Psomas I: Yolk fatty acid composition and cholesterol content in response to level and form of dietary flaxseed. *J Agr Food Chem*, 46, 4652-4656, 1998.

44. Ansari R, Azarbayejani A, Ansari S, Asgari S, Gheisari A: Production of egg enriched with omega-3 fatty acids in laying hens. *Arya J*, 1, 242-246, 2006.

45. Maurice D, Lightsey SF: Cholesterol ve fatty acids in eggs from hens fed high levels of copper. *Poult Sci*, 79 (Suppl. 1): 75 (Abstr.), 2000.

The Statistical Analysis of Some Volumetric Measurements in the Japanese Quails' Head with Different Feather Color: A Computed Tomography Study

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Summary

The aim of this study was to contribute to quail head morphology and the formation of basic data sources for comparative measurement in the quail head. In this study, 60 Japanese quail (*Coturnix japonica*) heads, which were white, brown and wild type were used. Four different anatomical structures of the quail heads, varying in feather color, were measured manually by two independent radiologists. These anatomical structures were head volume (HV), brain volume (BV), parietooccipital air space volume (POAV) and calvarial bone volume (CBV). According to computed tomography (CT) measurements, the quail head with white feather color had the smallest volumetrical values. This condition is thought to be caused by genetic differences.

Keywords: *Computed Tomography, Japanese quail, Morphometry*

Farklı Tüy Renklerine Sahip Japon Bildircinlerinin Başlarında Bazı Hacimsel Ölçülerin İstatistiksel Analizi: Bilgisayarlı Tomografi Çalışması

Özet

Bu çalışmanın amacı bildircin başlarında karşılaştırmalı ölçümler için temel veri kaynağı oluşturmak ve bildircin başı morfolojisine katkı sağlamaktır. Çalışmada beyaz, kahverengi ve yabani tip olmak üzere toplam 60 Japon bildircini başı kullanıldı. Farklı renklere göre bildircin başlarının dört farklı anatomik yapısı iki radyolog tarafından ölçüldü. Bu yapılar kafa hacmi, cavum cranii hacmi - beyin hacmi, parietooccipital hava alanı hacmi ve kafa kemikleri hacmi şeklindeydi. Bilgisayarlı Tomografi ölçülerine göre, beyaz tüy rengine sahip bildircin başının en küçük hacimsel değerlere sahip olduğu belirlendi. Bu duruma genetik farklılıkların neden olduğu düşünülmektedir.

Anahtar sözcükler: *Bilgisayarlı Tomografi, Japon Bildircini, Morfometri*

INTRODUCTION

The quail (*Coturnix japonica*) is a species categorized under Phasianidae family [1,2]. The quail is widely used as an *in vivo* [3-6] in physiological, pathological, toxicological and anatomical studies owing to its lower feed consumption, early sexual maturity and fast growth. In recent years, the increase in consumption of meat and eggs of the quail has also strengthened its economic aspect [1,2,7].

feature. Differences in the Quail feather color are attributed to mutations in the color [8]. The feather colors are white [9], brown, yellow [9,10], wild type (gray) [11] and roux [10].

In terms of anatomical model preparation, computed tomography (CT) is a quite useful method for reliable topographical patterns [12]. CT is used for antropomorphometrical studies for creating macroscopic and microscopic models and revealing the phenotypic differences quickly in

Quail feather color is considered as a race or genetical



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small animals^[13-17]. The detailed imaging of structures that make up the skull and the minimum degree of overlapping images provides a great advantage to the CT compared to other radiological methods^[18,19]. One of the advantages of the CT is the short-term applicability^[20]. Using CT, high resolution images are obtained from cross-sections of head, and subsequently anatomical or pathological data could be evaluated^[21-24]. Three-dimensional images of anatomical structures must be accurately obtained for the zoo-morphological requirements, and functional, developmental, comparative studies^[25]. Additionally, the development of high resolution imaging techniques such as magnetic resonance imaging (MRI) and CT facilitated to create a small animal model and anatomical evaluations^[26-30]. Volumetric measurements of small organisms or non-living structures can be evaluated via the three-dimensional images obtained from CT^[25]. Three-dimensional imaging method of animal materials can be gathered into two broad categories: the first one is based on reconstruction from serial section images and the second one is based on whole - volume imaging^[25]. A volume image obtained from CT consists of a stack of reconstructed cross sections around the axis of rotation^[31]. Therefore, the organism could easily be evaluated in terms of the development, growth and structural differences owing to obtained images^[25]. In the next step, some morphometric and volumetric measurements are practised by using these images^[20].

Roentgen and CT are diagnosis methods which are frequently referred to in veterinary practice^[32,33]. CT has just begun to be used in the field of avian medicine^[20]. Therefore, the evaluation and interpretation of images are possible only by becoming familiar with a good level of macro - anatomy of the region and the cross - sectional anatomy^[33].

Turbinates, sinuses, nasal cavity, and other anatomical structures can be monitored by using CT in detail in the bird head^[20]. In previous studies, anatomical CT images were excessively employed in imaging the head, nasal - paranasal sinuses, cranial cavity, nasolacrimal duct system, and turbinates have been analyzed in terms of and its three dimensional reconstructed models^[24,34-39] in the differences animal specieses (reptile, sea turtle, donkey, rodentia, dog and cat^[32-36,39]). However, there was no a study which reveals the anatomical data of the quail head by using CT in the literature.

The aim of this study was to contribute to quail head morphology and the formation of basic data sources for comparative measurement in the quail head using CT. Additionally, in this study, using the above-mentioned characteristics of CT was purposed to determined both status of imaging quality and parameters of method in quail head and to identify whether or not the difference term some volumetrical measurements (see materials and methods section) among quail heads, varying in feather color.

MATERIAL and METHODS

In this study 60 quail heads were used. A total of 20 of them were white, 20 of them were brown and 20 of them were wild type. Quails were supplied from Quail Unit in the Education, Research and Application Farm of Kafkas University. Quails were six weeks, female and average 120-160 g (white: 130 ± 5.3 g, brown: 148 ± 7.6 g, wild type: 140 ± 8.2 g) live weight. Animals were grown under the same live conditions (light intensity, feeding, the cage state, water etc.). Quails were killed by slaughtering for protect to wastable quilty of carcase and their heads were referred to the Research and Application Hospital Radiology Unit of Kafkas University to obtain CT images. To scan the heads 64 sliced CT (0.05 mm section) device (Aquilion 64®, Toshiba Medical Systems, 2011, Zoetermeer. The Netherlands) was used.

Four different anatomical structures of each group were measured manually by two independent radiologists with more than five-years experience. Radiologists were defined as observer 1 and observer 2. These anatomical structures were head volume (HV), brain volume (BV) (Fig. 1-D), parietooccipital air space volume (diverticula from recessus tympanicus dorsalis invade the occipital, parietal, prootic and squamosal bones^[40]) (POAV) (Fig. 1-A, Fig. 1-B, Fig. 2-F) and calvarial bone volume (CBV). Aquarius iNtuition Edition ver. 4. 4. 6. software was used for measurements.

Different tissues have different densities in CT, so four different density ranges were used to calculate four different tissue volumes. To calculate the POAV from -1.000 to -850 Hounsfield unit (HU), HV 0-1000 HU, BV 0-150 HU and CBV 200-1000 HU ranges were^[41] setted. Both 2D axial and coronal plane images were used to calculate all the volumes. In addition to 2D images, measurements were verified also on 3D volume rendered images. For the volume unit, cm³ were used and all the data were recorded carefully.

To calculate the BV and POAV region growing tool (Fig. 1-A, Fig. 1-D), CBV and HV single click tool (Fig. 1-C) was used. Because BV and POAV are spread to area, but CBV and HV are not.

Four different anatomical structure of three kind of quail heads were measured by two radiologists and many different parameters obtained. To process the findings easily, we named the variables as: head volume of observer 1 (HV1), head volume of observer 2 (HV2), brain volume of observer 1 (BV1), brain volume of observer 2 (BV2), parietooccipital air space volume of observer 1 (POAV1), parietooccipital air space volume of observer 2 (POAV2), calvarial bone volume of observer 1 (CBV1), calvarial bone volume of observer 2 (CBV2).

Statistical Analysis

Statistical analysis of the study was performed using

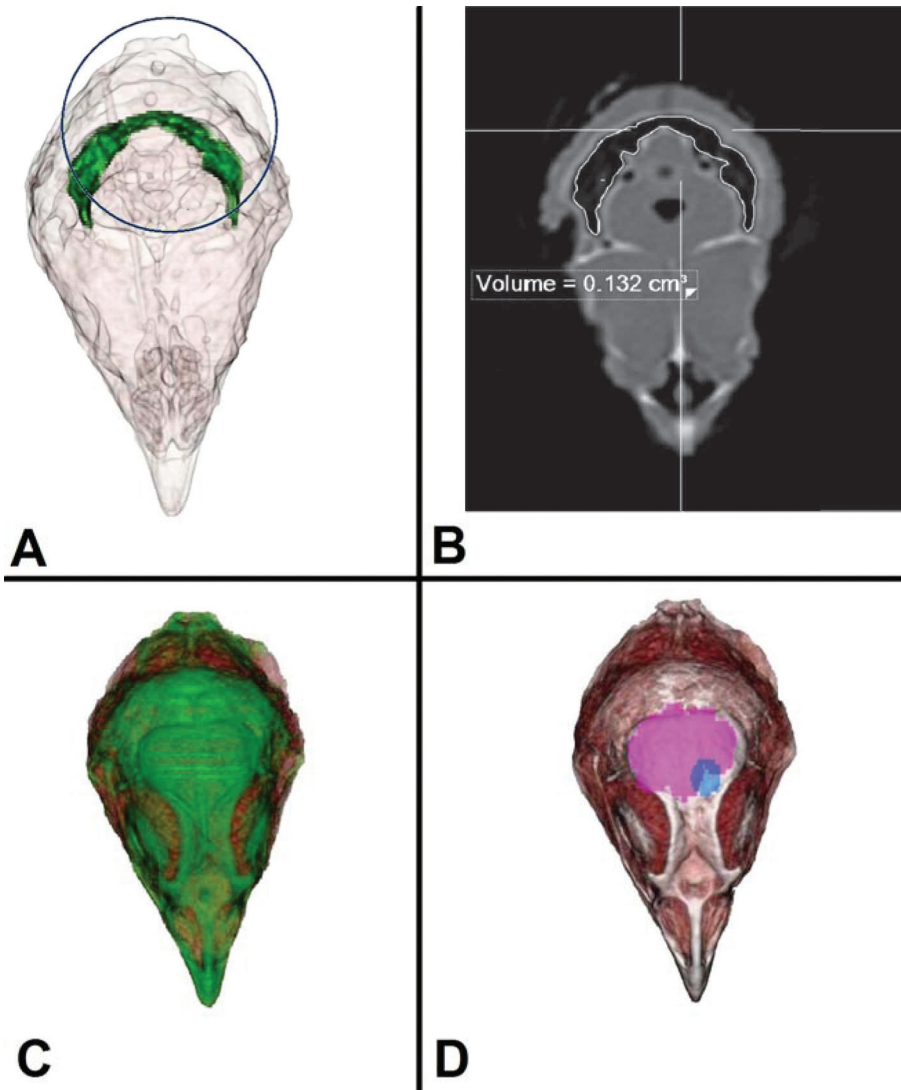


Fig 1. A- 3D volume rendered dorso-ventral aspect image of parietooccipital air space volume of quail head measured by region growing tool, B- 2D axial slice of parietooccipital air space volume of quail head measured by region growing tool, C- 3D volume rendered dorso-ventral aspect image of head volume of quail measured by single click tool, D- 3D volume rendered dorso-ventral aspect image of brain volume of quail head measured by region growing tool

Şekil 1. A- Bildircin başındaki parietooccipital hava alanı hacminin bölge büyütme aracı kullanılarak ölçümünün dorso-ventral yönden 3B görüntüsü, B- Bildircin başındaki parietooccipital hava alanı hacminin bölge büyütme aracı kullanılarak ölçümünün 2B eksen kesit görüntüsü, C- Bildircin kafa hacminin tek tık aracı kullanılarak ölçümünün dorso-ventral yönden 3B görüntüsü, D- Bildircin başındaki beyin hacminin bölge büyütme aracı kullanılarak ölçümünün dorso-ventral yönden 3B görüntüsü

Fig 2. Dorso - ventral view of transversal section through of the quail head. A- brain, B- cerebellum, C- occipital bone, D- oculus, E- rostrum, F- parietooccipital air space

Şekil 2. Bildircin kafasının transversal kesitin dorso - ventral görünüşü. A- beyin, B- beyincik, C- occipital kemik, D- göz, E- gaga, F- parietooccipital hava alanı



Statistical Package for Social Sciences (SPSS) version 17.0 software package. Continuous variables were expressed as arithmetical mean \pm standard deviation.

Paired sample t test was used to determine the differences of the measurements of BV, HV, POAV and CBV between observer 1 and observer 2 regardless of the

quail types and the groups were dependent according to observers.

Kruskal Wallis test used to calculate the differences of BV, POAV, CBV and HV of white, brown and wild type subgroups for observer 1 and 2 independently. Firstly three groups were analyzed, secondly white-brown, white-wild

and brown-wild groups were analyzed in pairs to find out the differences between the groups.

RESULTS

According to the findings of the study, average values of the BV, POAV, CBV and HV of the quails (n:60) were 0.43, 0.23, 1.93 and 6.64 cm³, respectively.

According to the observers, the average results obtained from the study showed in *Table 1*. In this table, it seems that values of observer 1 are a little more than the values of observer 2.

Table 2 indicated that each volume of three groups of quail heads showed significant difference between each other, except BV2. Moreover, the same table showed significant difference between white and brown quail head measurements except BV2 and CBV2. Additionally, *Table 2* showed significant difference between wild type and brown quail head measurements except POAV1, BV2 and POAV2, and significant difference between white and wild type quail head measurements except BV2 and POAV2 volumes ($P<0.05$). This results indicated that approximately all of the four measurements of the three kind of quail heads were different from each other.

Paired samples t test demonstrates that there were no significant difference in CBV and HV measurements and there were significant difference in BV and POAV between the observers ($P<0.05$).

DISCUSSION

One should consider the limitations of this study before using these data. We have firstly reported the volumetric measurements of head in the quail by using CT. Thus, we were not able to discuss the variation of morphometrical properties of the bird head. While this study provides new morphometrical data for four volumetric measurement in the quail head, volumetric morphometrical values for other species, such as the chicken, goose, and pigeon, are also needed. When we performed this study, there was not a method established in the birds. Therefore, we could not compare in terms of method.

According to the findings of the study, average values of the BV, POAV, CBV and HV of the quail (n:60) were 0.43, 0.23, 1.93 and 6.64 cm³, respectively. According to this situation, BV was forming %6.47 of HV, POAV was forming %3.46 of HV, and CBV was forming %29.07 of HV.

BV and POAV differences were due to the subjective results of the region growing tool and method. CBV and HV analogy were due to the single click method based on our results. Single click is a useful tool that allows measuring the volume by clicking only once to calculate the volume. However, region growing is time consuming and relatively difficult. In this regard, the results of single click are more objective than region growing method.

In this study, feather color having the race property for quails was primary exit point. Therefore, the heads of quails

Table 1. Shows all the data of each groups based on observers

Table 1. Araştırmacılar göre grupların verilerinin tamamı görünmektedir

Measurement	Observer 1			Observer 2		
	White	Brown	Wild	White	Brown	Wild
BV (region growing)	0.45±0.07	0.56±0.07	0.50±0.05	0.36±0.67	0.37±0.05	0.34±0.10
POAV (region growing)	0.21±0.07	0.33±0.07	0.29±0.04	0.15±0.05	0.19±0.06	0.20±0.19
CBV (single click)	1.56±0.27	1.79±0.32	2.49±0.29	1.63±0.26	1.79±0.32	2.35±0.36
HV (single click)	5.92±0.41	6.60±0.65	7.50±0.65	5.88±0.34	6.50±0.81	7.45±0.68

The unit of the mean values and the standard deviations are cm³

Table 2. Kruskal Wallis Test of the white, brown and wild type subgroups (W-White, B-Brown, WT-Wild Type)

Table 2. Beyaz, kahverengi ve yabani tipin Kruskal Wallis Testi (W-Beyaz, B-Kahverengi, WT-Yabani Tip)

Test	HV1	BV1	POAV1	CBV1	HV2	BV2	POAV2	CBV2
Chi-Square (W, B&WT)	*33.75	*22.1	*19.71	*36.83	*32.76	2.84	*7.07	*26.63
Chi-Square (W&B)	*11.71	*18.62	*15.81	*6.4	*10.11	0.27	*6.94	3.05
Chi-Square (WT&B)	*13.14	*9.51	3.59	*22.55	*11.35	2.64	3.64	*16.15
Chi-Square (W&WT)	*26.56	*5.10	*10.36	*27.27	*27.83	1.32	0.05	*20.91

* Means within a row are significantly different ($P<0.05$)

in three different feather colors were used. There are many quails with different feather colors in the world. Some of them are white, brown, roux, yellow and gray or wild type^[8-11]. In this study, different races of quails were used to compare morphological and morphometric differences. Thus, both a method for subsequent studies and a data base were formed.

CT measurements shows that the wild type quail has the biggest head and then brown and white quails were ranking lower, respectively. The brown quail has the biggest cranial cavity volume and then it is followed by wild type and white had smallest. Moreover, the wild type quail has the biggest head bone volume and then brown and white quails have respectively. According to these results, quail race with white feather color had the smallest volumetrical values. Although more comprehensive studies are needed to determine the cause of the above mentioned situation, we could speculate that the applied selection pressure in the quail unit and genetic differences can be effective.

When data of head morphometry in the quails are evaluated study material is provided from single sex and equal number of members. Female quails constituted the animal material of this study. Hence, the data obtained from female quails was analyzed.

The lack of previous CT based studies in the avian literature hampered the comparisons. For this reason, the results that we have reported here have potential to contribute not only to the assessments of species but also the comparative and topographic anatomy of birds.

In conclusion, this study showed us that CT can be used for morphological and morphometric studies in quail head, additionally birds and differences term some volumetrical measurements among quail heads, varying in feather color. Furthermore, we concluded that more comprehensive studies are necessary to determine the exact relationship between feather color (or race) and some morphometric measurements in the Japanese quails.

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REFERENCES

1. **İde T:** Laboratuvar Hayvanları Biliminin Temel İlkeleri. Medipres Yayınları, Özkan Matbaacılık, Ankara, 2003.
2. **Sarıca M, Camcı Ö, Selçuk E:** Bildircin, Sülün, Keklik, Etçi Güvercin, Beç Tavuğu ve Devekuşu Yetiştiriciliği. No: 4, 3. Baskı, Samsun, 2003.
3. **Fitzgerald CT:** The Coturnix Quail Anatomy and Histology. Iowa State University Press, Ames Iowa, 1970.
4. **Howes JR, Ivey WD:** Coturnix quail for avian research. Feedstuffs, May 12, 1-2, 1961.
5. **McLelland J:** A Colour Atlas of Avian Anatomy. Wolfe Publishing Ltd.

England, 1990.

6. **Sreeranjini AR, Iyyangar MP, Pramodkumar D:** Histological study on the fibrous architecture of kidney and ureter of japanese quail (*Coturnix coturnix japonica*). *J Vet Anat Sci*, 6 (2): 107-110, 2010.
7. **Taşbaş M, Hazıroğlu RM, Çakır A, Özer M:** Denizli horozunun solunum sisteminin morfolojisi I. Cavitas nasalis. *Ankara Univ Vet Fak Derg*, 41, 63-80, 1994.
8. **Cneg KM, Kimura M:** Mutations and major variants in japanese quail. Chapter 13. Crawford RD (Ed): Poultry Breeding and Genetics. pp.33-362, Elsevier, Amsterdam, 1990.
9. **Truax RE, Johnson WE:** Genetics of plumage color mutants is japanese quail. *Poultry Sci*, 58, 1-9, 1979.
10. **Nadeau NJ:** The Evolutionary Genetics of Sexually Selected Plumage Colour Traits in the Galliform Birds, Clare Collage, 2006.
11. **Yıldız MA, Kesici T:** Japon bildircinlarda (*Coturnix japonica*) sarı ve lekeli beyaz tüy rengini belirleyen genler arasındaki genetik ilişkinin incelenmesi. *Lalahan Hay Arast Enst Derg*, 37 (2): 84-90, 1997.
12. **Elliott RC:** A comparative computed tomography study of canine laparoscopic abdominal anatomy pre- and post insufflation. Pretoria Univ, 2011.
13. **Turner CH, Sun Q, Schrieffer J, Pitner N, Price R, Boussein ML, Rosen CJ, Donahue LR, Shultz KL, Beamer WG:** Congenic mice reveal sex-specific genetic regulation of femoral structure and strength. *Calcif Tissue Int*, 73, 297-303, 2003.
14. **Silha JV, Mishra S, Rosen CJ, Beamer WG, Turner RT, Powell DR, Murphy LJ:** Perturbations in bone formation and resorption in insulin-like growth factor binding protein-3 transgenic mice. *J Bone Miner Res*, 18, 1834-1841, 2003.
15. **Moverare S, Venken K, Eriksson AL, Andersson N, Skrtic S, Wergedal J, Mohan S, Salmon P, Bouillon R, Gustafsson JA, Vanderschueren D, Ohlsson C:** Differential effects on bone of estrogen receptor alpha and androgen receptor activation in orchidectomized adult male mice. *Proc Natl Acad Sci, USA*, 100, 13573-13578, 2003.
16. **Paulus MJ, Gleason SS, Easterly ME, Foltz CJ:** A review of high-resolution X-ray computed tomography and other imaging modalities for small animal research. *Lab Animal*, 30, 36-45, 2001.
17. **Kindlmann GL, Weinstein DM, Jones GM, Johnson CR, Capecchi MR, Keller C:** Practical vessel imaging by computed tomography in live transgenic mouse models for human tumors. *Molec Imag*, 4, 417-424, 2005.
18. **Barbee DD, Allen JR, Gavin PR:** Computed tomography in horses: Tech-nique. *Vet Radiol Ultrasoun*, 28, 144-151, 1987.
19. **Gibbs C:** Dental imaging. In, Baker GJ, Easley J (Eds): Equine Dentistry. pp. 139-169, Philadelphia, WB Saund, 1999.
20. **Forbes NA:** Advanced Imaging Diagnostics in Avian Veterinary Practice. pp. 34 -35, Parrots, 2011.
21. **Regedon S, Franco A, Garin JM, Robina A, Lignereux Y:** Computed tomographic determination of the cranial volume of the dog applied to racial and sexual differentiation. *Acta Anat*, 142, 347-350, 1991.
22. **Regedon S, Lignereux Y, Garin JM, Martin A, Robina A:** Volumetry of the cranial cavity of the pointer: Computed tomographic determination. *Rev Med Vet*, 142, 907-910, 1991.
23. **Regedon S, Franco A, Lignereux Y, Garin JM, Robina A, Martin A:** Skull volume in Pekinese dogs. Tomodensitometry and sex-linked difference. *Rev Med Vet*, 143, 745-748, 1992.
24. **Robina A, Regedon S, Guillen MT, Lignereux Y:** Utilization of computed tomography for the determination of the volume of the cranial cavity of the Galgo hound. *Acta Anat*, 140, 108-111, 1991.
25. **Metscher BD:** MicroCT for comparative morphology: simple staining methods allow high-contrast 3D imaging of diverse non-mineralized animal tissues. *BMC Physiol*, 9, 11, 2009.
26. **Hirsch J:** Imaging and biological function in health and disease. *J Clin Invest*, 111, 1440-1443, 2003.
27. **Weissleder R, Mahmood U:** Molecular Imaging. *Radiol*, 219, 316-333, 2001.

- 28. Kiessling F, Heilmann M, Vosseler S, Lichy M, Krix M, Fink C, Kiessling I, Steinbauer H, Schad L, Fusenig NE, Delorme S:** T1 dynamic - weighted monitoring of vascularization in human carcinoma heterotransplants by magnetic resonance imaging. *Int J Cancer*, 104, 798, 2003.
- 29. Krix M, Kiessling F, Vosseler S, Farhan N, Mueller MM, Bohlen P, Fusenig NE, Delorme S:** Sensitive noninvasive monitoring of tumor perfusion during antiangiogenic therapy by intermittent bolus - contrast power Doppler sonography. *Cancer Res*, 63, 8264-8270, 2003.
- 30. Liao ZX, Travis EL, Tucker SL:** Damage and morbidity from pneumonitis after irradiation of partial volumes of mouse lung. *Int J Radiol Oncology Biol Physiol*, 32, 1359-1370, 1995.
- 31. Kalender WA:** Computed Tomography: Fundamentals, System Technology, Image Quality, Applications. Erlangen, Publicis Corporate Publishing, 2005.
- 32. Arencibia A, Rivero MA, Miguel De I, Contreras S, Cabrero A, Oros J:** Computed tomographic anatomy of the head of the loggerhead sea turtle (*Caretta caretta*). *Res Vet Sci*, 81, 165-169, 2006.
- 33. Banzato T, Selleri P, Veladiano IA, Martin A, Zanetti E, Zotti A:** Comparative evaluation of the cadaveric, radiographic and computed tomographic anatomy of the heads of green iguana (*Iguana iguana*), common tegu (*Tupinambis merianae*) and bearded dragon (*Pogona vitticeps*). *BMC Vet Res*, 8, 53, 2012.
- 34. El-Gendy SA, Alsafy MAM:** Nasal and paranasal sinuses of the donkey: Gross anatomy and computed tomography. *J Vet Anat*, 25-41, 2010.
- 35. Phillips JE, Ji L, Rivelli MA, Chapman RW, Corboz MR:** Three-dimensional analysis of rodent paranasal sinus cavities from X-ray computed tomography (CT) scans. *Canada J Vet Res*, 73, 205-211, 2009.
- 36. García-Real I, Kass PH, Sturges BK, Wisner ER:** Morphometric analysis of the cranial cavity and caudal cranial fossa in the dog: A computed tomographic study. *Vet Radiol Ultrasound*, 45 (1): 38-45, 2004.
- 37. Onar V, Kahvecioğlu KO, Çebi V:** Computed tomographic analysis of the cranial cavity and neurocranium in the German shepherd dog (Alsatian) puppies. *Vet Archiv*, 72, 57-66, 2002.
- 38. Onuk B, Kabak M, Sahin B, Ince NG, Selcuk MB:** Kazda (Anser anser domesticus) cavitas nasale, septum nasi ve concha'ların hacim ve hacim oranlarının fiziksel kesit ve bilgisayarlı tomografi görüntülerinden karşılaştırmalı olarak hesaplanması. *VII. Ulusal Veteriner Anatomi Kongresi*, Antalya, 27-29 Ekim 2011.
- 39. Schlueter C, Budras KD, Ludewig E, Mayrhofer E, Koenig HE, Walter A, Oectering GU:** CT and anatomical study of the relationship between head conformation and the nasolacrimal drainage system, *J Feline Med Surg*, 11, 891-900, 2009.
- 40. Baumel JJ, King SA, Breasile JE, Evans HE, Berge JCV:** Nomina Anatomica Avium. 2nd ed., Prepared by the International Committee on avian Anatomical Nomenclature, A Committee of The World Association of Veterinary Anatomists. Nuttall Ornithological Club. Cambridge, Massachusetts, 1993.
- 41. Hofer M:** CT Teaching Manual: A Systematic Approach to CT Reading. Third ed., Thieme Medical Publishers, 2008.

The Anatomy of the Cardiac Veins in Storks (*Ciconia ciconia*)

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Summary

This study was carried out to describe the origin, course and ramifications of the cardiac veins in storks. For this purpose, coloured latex injection method was applied on heart of nine adult storks. The results indicate that venous drainage of the heart was provided by the *v. cardiaca sinistra*, the *v. cardiaca media*, the *vv. cardiae dextrae* and the *vv. cardiae minimae*. The coronary sinus was not found in all samples. The *v. cardiaca sinistra* and *v. cardiaca media* were opened into the *v. cava cranialis sinistra*. The *v. cardiaca sinistra* was the largest vein providing to venous drainage of heart which consisted of two main parts *pars interventricularis* and *v. cardiaca circumflexa sinistra*. The *v. cardiaca circumflexa sinistra* was collected to venous blood of the ventriculus sinister and atrium sinistrum by means of several vessels. One branch of the *v. cardiaca circumflexa sinistra* coming from ventriculus sinister was much larger than rest and drained big part of ventriculus sinister. The *v. cardiaca media* was a single vein. The *vv. cardiae dextrae* were opened directly into the right atrium. The *vv. cardiae minimae* were collected venous blood from the wall of atrium dextrum and interventricular septum and emptied into the right chambers of heart. The venous blood of the interventricular septum was collected by the *vv. septales* of *pars interventricularis* and *v. cardiaca media*. It was concluded that circulation of cardiac veins in stork closely resembles that of rodent and is unlike both fowl and ostrich patterns.

Keywords: Anatomy, Cardiac veins, Heart, Stork

Leyleklerde (*Ciconia ciconia*) Kalp Venlerinin Anatomisi

Özet

Bu çalışmada, leylekte koroner venlerin başlangıç, seyir ve dallanmaları incelendi. Bu amaçla dokuz adet yetişkin leyleğe renklendirilmiş latex verildi. Kalbin venöz drenajının *v. cardiaca sinistra*, *v. cardiaca media*, *vv. cardiae dextrae* ve *vv. cardiae minimae* tarafından sağlandığı tespit edildi. Sinus coronarius'un bulunmadığı belirlendi. *V. cardiaca sinistra* ve *v. cardiaca media*'nin *v. cava cranialis sinistra*'ya açıldığı görüldü. *V. cardiaca sinistra*'nın kalbin venöz kanını toplayan en güçlü ven olduğu tespit edildi. Bu damarın, *pars interventricularis* ve *v. cardiaca circumflexa sinistra* olarak iki bölümden oluştuğu görüldü. *V. cardiaca circumflexa sinistra*'nın bir çok damar aracılığı ile ventriculus sinister ve atrium sinistrum'un venöz kanını topladığı görüldü. Ventriculus sinister'den çıkan *v. cardiaca circumflexa sinistra*'nın bir dalının diğer dallardan güçlü olduğu ve ventriculus sinister'in büyük bir bölümünü diranajını sağladığı tespit edildi. *V. cardiaca media*'nin tek olarak şekillendiği belirlendi. *Vv. cardiae dextrae*'nin doğrudan atrium dextrum'a açıldığı belirlendi. *V. cardiae minimae*'nin atrium dextrum duvarı ve septum interventriculare'nin venöz kanını topladıkları ve kalbin sağ boşluklarına açıldığı görüldü. Septum interventriculare'nin drenajını *v. cardiaca media* ile *pars interventricularis*'in *vv. septales*'inin yaptığı tespit edildi. Sonuç olarak leyleğin koroner ven dolaşımı tavuk ve devekuşundan ziyade kemirgenlere daha çok benzediği görüldü.

Anahtar sözcükler: Anatomi, Kalp venleri, Kalp, Leylek

INTRODUCTION

The venous drainage of the heart is provided by the *v. cardiaca sinistra*, the *v. cardiaca media*, the *vv. cardiae dextrae* and the *v. cardiae minimae* in birds^[1,2]. The *v. cardiaca sinistra* of birds are formed by the *pars interventricularis* and the *pars basilaris*^[1-3]

The *pars interventricularis* is started near the apex of the heart and run in the paraconal interventricular groove. The *pars interventricularis* at the coronary groove border continues as the *pars basilaris*, which lies on the base of the heart, between the conus arteriosus and bulbus aortae

and opened into right atrium ^[1-4]. However, the *pars basilaris* is not found in some samples of fowl ^[1].

Vena cardiaca circumflexa sinistra is begun by the fine superficial tributaries from the proximal part of the left aspect of the ventriculus sinister. Then, it runs in the coronary groove and opened into atrium dextrum ^[1,3]. *V. cardiaca media* was begun just above the apex of the heart. After it was received many ventral and dorsal tributaries from the adjacent wall of the ventriculus sinister and ventriculus dexter, led to right atrium ^[1-4]. *Vv. cardiaca dextrae* are consisted by veins draining the ventriculus dexter and led to the atrium dextrum. *Venae cardiaca minimae* are found constantly in the wall of atrium dextrum and the interventricular septum ^[1].

Generally, the coronary sinus collecting of principal cardiac veins is a large blood vessel or dilation of right atrium ^[5]. Although the coronary sinus is emphasized to exist in ostrich ^[3,6], some authors state that domestic birds ^[1,4] have not a coronary sinus.

Birds are the most diverse group of animals rather than mammals which are alive today ^[7]. Although venous drainage of heart has been well documented in mammalian, there is no study about the venous drainage of bird's heart, except fowl (*Gallus domesticus*) ^[1,8-10] and ostrich ^[3]. Therefore, this study aimed to reveal the conformation and branching of the cardiac veins in stork (*Ciconia ciconia*) which is wild and a migrant bird.

MATERIAL and METHODS

Nine adult storks of both sexes and weighing between 2.5 and 3 kg were utilized for the study. The animals used in the study were obtained from Mustafa Kemal University and Adana Veterinary Control and Research Institute. Animals which were could not survive due to badly injured or suspected various diseases were used. All experimental procedures were approved by Adana Veterinary Control and Research Institute's Animal Experiments Local Ethics Committee (Approval no. 02.02.2012/15).

Coloured latex injection was applied in order to describe the coronary veins as suggested by literatures ^[11,12].

Coelom of the storks were opened right just after death. The heart was removed from the coelom. Plastic catheters were inserted into the *v. cava caudalis*. All cardiac vessels and chambers were washed with 0.9% saline (NaCl). After, two *v. cava cranialis* were clamped. After then, blue (Setacolor TM, Cobalt blue, num. 20; PEBEO Cedex, France) colored latex (Rubber latex TM; MERCAN, Istanbul, Turkey) was injected into the *v. cava caudalis*. The hearts were then immersed in 10% formalin for 3 days. After solidification of latex solution, the cardiac veins were investigated macroscopically using a dissection microscope (Nikon SMZ-2T; Nikon Corp., Tokyo, Japan). Photographs were

taken digitally and stored on a personal computer. The vessels were measured with a digital caliper (DV892 calipers, Tecnotest corp., Germany) sensitivity of 0.1% mm.

Nomina Anatomica Avium ^[2] was used for anatomical nomenclature.

RESULTS

The veins draining the heart of the stork were the *v. cardiaca sinistra*, the *v. cardiaca media*, the *vv. cordicae dextrae* and the *vv. cardicae minimae*. The coronary sinus was absent in all of the studied samples.

The *vena cardiaca sinistra* (Figs. 1,2/1): In all the samples, the *v. cardiaca sinistra* was the largest vein carrying the venous blood of the heart and emptied into the *v. cava cranialis sinistra*. During its course, the *v. cardiaca sinistra* was named as the *pars interventricularis* (Figs. 1,3/2) in the paraconal interventricular groove and the *v. cardiaca circumflexa sinistra* (Figs.1,2/3) in the coronary groove.

The *pars interventricularis* (Figs. 1,3/2): The *pars interventricularis* began at the level of the apex of the heart and anastomosed with the *v. cardiaca media* (Fig. 3/→) at the same level. The *pars interventricularis* was formed by the 5-6 *vv. ventriculares* (Fig. 1/4) coming from the ventriculus sinister and 4-5 *vv. ventriculares* (Fig. 1/5) from ventriculus dexter. The terminal branches of the *pars interventricularis* of *v. cardiaca sinistra* anastomosed with the branches of *vv. cordicae dextrae* (Fig. 3/→). Along its course, the *pars interventricularis* also received 6-7 small *vv. septales* (Fig. 1/6) from interventricular septum. A strong *v. septalis* draining the proximal half to a third of the interventricular septum was also joined the *pars interventricularis* in three samples. Just before continuing as *v. cardiaca circumflexa sinistra*, the *pars interventricularis* also received the *v. conalis* (Fig. 1/7) draining the left part of conus arteriosus and proximal portion of the ventriculus dexter. The terminal branches of *v. conalis* were found to anastomose with the *v. conalis* (Fig. 1/→) of *vv. cordicae dextrae* in the four of the investigated hearts.

The *vena cardiaca circumflexa sinistra* (Figs. 1,2/3): The *pars interventricularis* coursed in the coronary groove as the *v. cardiaca circumflexa sinistra*. It was entirely covered by the auricula sinistra. Along its course, the *v. cardiaca circumflexa sinistra* received 3-4 thin *vv. ventriculares* (Figs. 1,2/8) starting from the proximal part of the ventriculus sinister. In all hearts, it was noted that one of the *vv. ventriculares* coming from the ventriculus sinister was the most distinct one. In all samples, this vein (Figs. 1,2/9) started from the apex of the heart and flowed towards the base of the heart on the ventriculus sinister, parallel to the *v. cardiaca media*. Along its course, it collected several subbranches from the wall of ventriculus sinister. Its

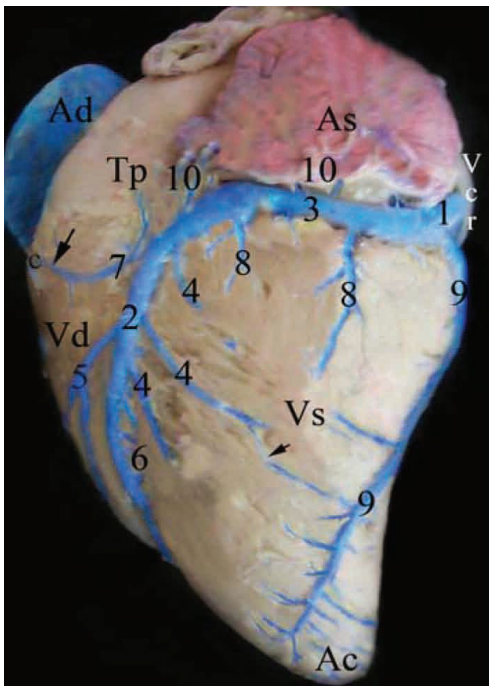


Fig 1. The *pars interventricularis*, the *v. cardiaca circumflexa sinistra* and their branches (cranial surface)

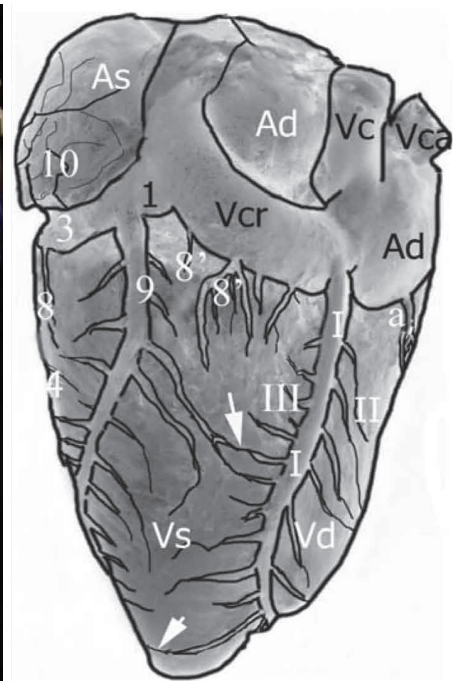
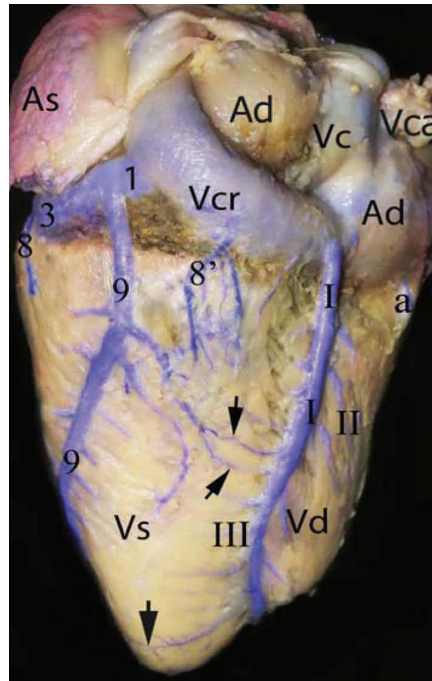
Şekil 1. *Pars interventricularis* ile *v. cardiaca circumflexa sinistra*'nın dalları (cranial yüz)

1: *v. cardiaca sinistra*, 2: *v. pars interventricularis*, 3: *v. cardiaca circumflexa sinistra*, 4-5: The *vv. ventriculares* of *pars interventricularis*, 6: The *vv. septales* of *pars interventricularis*, 7: The *v. conalis* of *pars interventricularis*, 8: The *vv. ventriculares* of *v. cardiaca circumflexa sinistra*, 9: The strong vein of *v. cardiaca circumflexa sinistra*, 10: The *vv. atriales* of *v. cardiaca circumflexa sinistra*, c: The *v. conalis* of *v. cardiaca circumflexa sinistra*, Tp: truncus pulmonalis, As: atrium sinistrum, Ad: atrium dextrum, Vd: ventriculus dexter, Vs: ventriculus sinister, AC: apex cordis, Vcr: *v. cava cranialis sinistra* and arrows: anastomose

Fig 2. The *v. cardiaca sinistra* and the *v. cardiaca media* and their branches (caudal surface)

Şekil 2. *V. cardiaca sinistra* ve *v. cardiaca media*'nin dalları (caudal yüz)

1: *v. cardiaca sinistra*, 3 *v. cardiaca circumflexa sinistra*, 8: The *vv. ventriculares* of *v. cardiaca circumflexa sinistra* 8': The *vv. ventriculares* of *v. cava cranialis sinistra*, 9: The strong vein of *v. cardiaca circumflexa sinistra*, 10: The *vv. atriales* of *v. cardiaca circumflexa sinistra*, I: *v. cardiaca media*, II,III: The *vv. ventriculares* of *v. cardiaca media*, a: The *vv. ventriculares* of *vv. cardiaca dextrae*, As: atrium sinistrum, Ad: Atrium dextrum, Vd: ventriculus dexter, Vs: ventriculus sinister, Vcr: *v. cava cranialis sinistra*, Vca: *v. cava cranialis dextra*, Vc: *v. cava caudalis* and arrows: anastomose



terminal branches were also seen to reach the papillary muscle in all samples. In one heart, it was also larger than the *v. cardiaca media*, according to its subbranches and diameter. Its branches also anastomosed with the *pars interventricularis* (Fig. 1/→) and *vv. ventriculares* of *v. cardiaca media* (Fig. 2/→). In five samples, 2 or 3 fine *vv. ventriculares* (Fig. 2/8') which were located on the proximal of ventriculus sinister were also seen to open directly into *v. cava cranialis sinistra*. The *v. cardiaca circumflexa sinistra* also received dorsally directed 5-6 thin *vv. atriales* (Figs. 1,2/10) from the wall of atrium sinistrum. The *v. cardiaca circumflexa sinistra* led into *v. cava cranialis sinistra*.

The *venae cardiaca media* (Fig. 1/I): The *v. cardiaca media* was formed by the branches stemming from the apex of the heart and ascending towards the base of the heart in the subsinuosal interventricular groove. It was opened into the *v. cava cranialis sinistra*. The *v. cardiaca media* was thinner than the *v. cardiaca sinistra* and a single vein. Six or seven *vv. ventriculares* (Fig. 3/II) from the ventriculus dexter and 5-7 *vv. ventriculares* (Fig. 2/III) from the ventriculus sinister joined to the *v. cardiaca media*. The *v. cardiaca media* was also carried to the *vv. septales* (Fig. 4/IV) from the interventricular septum. These *vv. septales* were smaller and fewer in number than those opening to the *pars interventricularis*.

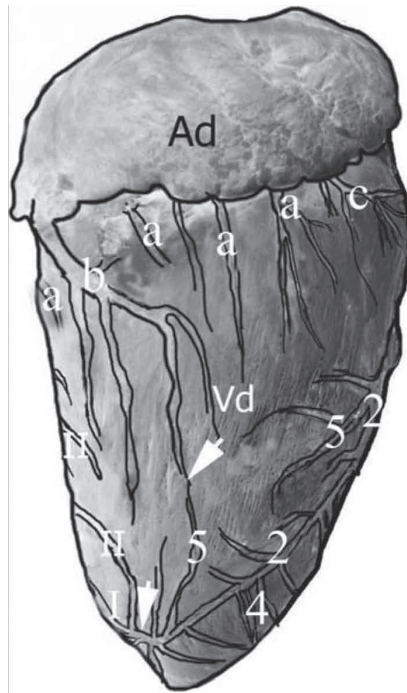
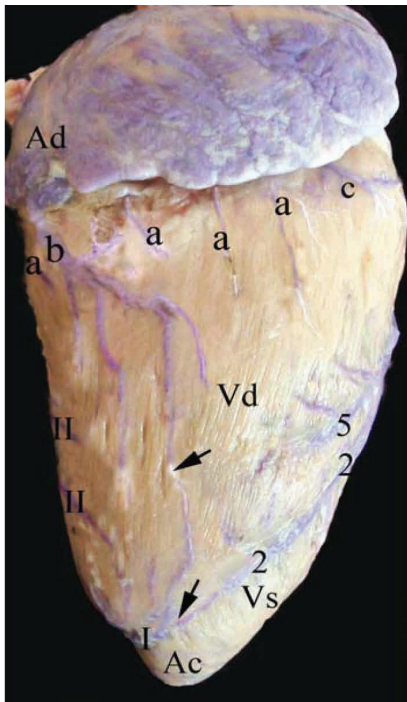


Fig 3. Vv. cardiacae dextrae (atrial surface)

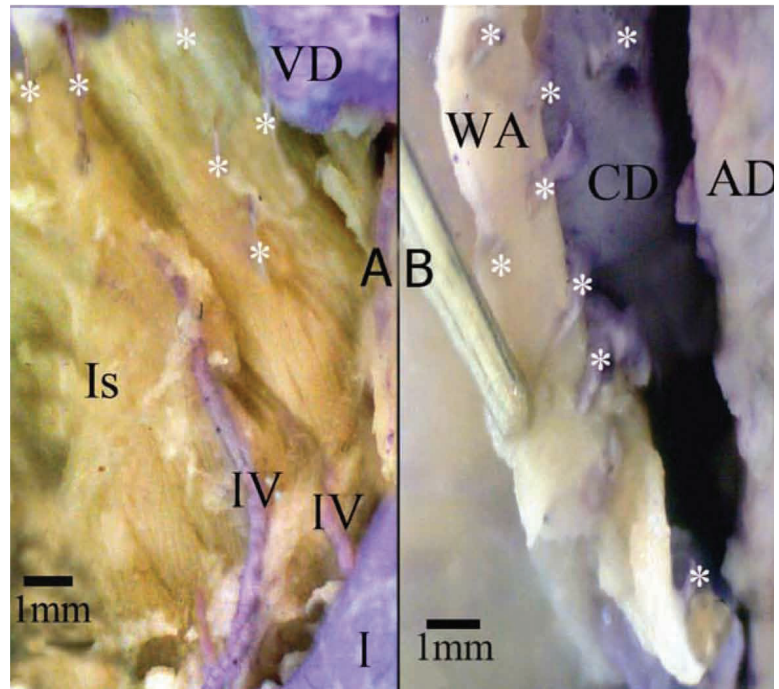
Şekil 3. Vv. cardiacae dextrae (atrial yüz)

I: v. cardiaca media, II: The vv. ventriculares of v. cardiaca media, a: The vv. ventriculares of vv. cardiacae dextrae, b: The strong vein of vv. cardiacae dextrae, c: v. conalis, 2: pars interventricularis, 5: The vv. ventriculares of pars interventricularis, Ad: Atrium dextrum, Vd: ventriculus dexter, AC: apex cordis and arrows: anastomose

Fig 4. The vv. septales and the vv. cardiacae minimae in the interventricular septum (A), The vv. cardiacae minimae on the wall of atrium dextrum (B)

Şekil 4. Septum interventriculares'te vv. septales ve vv. cardiacae minimae (A), Atrium dextrum duvarında vv. cardiacae minimae (B)

I: v. cardiaca media, IV: The vv. septales of v. cardiaca media, Is: interventricular septum, VD: The ventriculus dexter, CD: The cavity of atrium dextrum, AD: atrium dextrum, Aw: The wall of atrium dextrum and *: vv. cardiacae minimae



The venae cardiacae dextrae (Fig. 3/a,b,c): The vv. cardiacae dextrae consisted of 4-5 vv. ventriculares, v. conalis and 3-4 fine veins draining the right atrioventricular valve. The vv. ventriculares were joined to the vv. cardiacae dextrae at the distal border of the proximal third of the ventriculus sinister. After their origin, the vv. ventriculares run toward base of heart and emptied directly into right atrium (Fig. 3/a). In six specimens, one of vv. ventriculares was longer than the others. This vein (Fig. 3/b) originated from the 1/3 distal part of the ventriculus dexter and collected venous blood from the distal and middle part of wall of ventriculus

dexter. In addition to vv. ventriculares, another vessel involved in the formation of the vv. cardiacae dextrae was v. conalis. The v. conalis (Fig. 3/c) was formed by the thin branches stemming from the right part of conus arteriosus and the proximal portion of ventriculus dexter.

The venae cardiacae minimae (Fig. 4/*): The vv. cardiacae minimae were identified only on the right half of the heart. They seemed to play a role in the venous drainage of the interventricular septum and the wall of atrium dextrum. These veins were opened into the cavity of the

ventriculus dexter (Fig. 4-A/*) and the atrium dextrum (Fig. 4-B/*).

During their course, the *v. cardiaca circumflexa sinistra*, the *v. cardiaca media*, a thick subbranch of *v. cardiaca circumflexa sinistra* and the *pars interventricularis* were seen to course subepicardially. However, the thin branches of *v. cardiaca sinistra* and *v. cardiaca media* were situated intramyocardially. Although the *vv. ventriculares* of *vv. cardiacae dextrae* were located intramyocardially, the some thin branches belonging to *vv. cardiacae dextrae* were found subepicardially.

DISCUSSION

The *vena cava cranialis sinistra* is usually absent in carnivore, equine, ruminant [5]. However, this vein could be constant observed in birds [1-4], rat [13], beaver [14], mice [15], rabbit [12,16,17], marsupial [18]. In fowl, the *v. cava cranialis sinistra* is also termed the left precaval vein [1]. In the present study, the *v. cava cranialis sinistra* collected a great amount of venous blood of the heart, as reported in rabbits [12,16] beaver [14], mice [15] but does not correspond to the finding of Lindsay [1] who stated that the cardiac veins generally empty directly into the atrium dextrum in the fowl.

The coronary sinus is a tubular dilatation of the right atrium or it is the rudiment of the right sinus horn of the embryonic heart in some domestic animals [5,19] and humans [19]. Some researchers are reported that a great amount of venous blood is collected by coronary sinus, domestic animals [5,20], roe deer [21], porcupines [11] and human [19]. However, the coronary sinus was absent in stork, as observed in the domestic birds [1,4] and rat [13], beaver [14], mice [15], rabbit [12,16,17].

As indicated in the birds [1-3,22], the *v. cardiaca sinistra*, the *v. cardiaca media*, the *vv. cardiacae dextrae* and the *vv. cardiacae minimae* were identified in the stork heart.

In birds [1-3] *v. cardiaca sinistra* is constituted by two main veins, the *pars interventricularis* and the *pars basilaris*. In most fowl [1,22] and ostrich [3], after the *pars interventricularis* reaches to the junction of the paraconal interventricular groove and coronary groove, it advances caudally as the *pars basilaris* in the fat-filled space between the atrium sinistrum and truncus pulmonalis and aorta, and empties into the left recess of the atrium sinistrum. These arrangement and distribution are also reported in marsupial [21]. However, the *pars basilaris* was not found in our samples. The *pars interventricularis* was continued as the *v. cardiaca circumflexa sinistra* in coronary groove in all samples. Same finding is reported by Nickel et al. [5], Yoldas and Nur [12], Yadm and Gad [17], Aksoy et al. [20], Kabak and Onuk [21], Beşoluk and Tıpırdamaz [23] in mammalian and by Lindsay [1] and Kaupp [22] in some fowl.

The *pars interventricularis* of *v. cardiaca sinistra* is single vein that beginning at the level of the notch of cardiac apex in fowl [1]. This vein is a double vein in 13 percent of ostrich [3]. The one of them lies paraconal interventricular groove. The other one runs caudally to it on the wall of the ventriculus sinister. However, in the present study, the *pars interventricularis* was observed to be a single vein all samples. On the other hand, Lindsay [1] is reported that in 6 percent of fowl the *pars interventricularis* is not found.

In mammalian [5], two branches of paraconal interventricular vein is confirmed to be distributed to the wall of the ventriculus sinister that are called the left proximal collateral vein and the left distal collateral vein. In the investigated samples, there was not found the similar these veins. But, numerous fine branches of the *pars interventricularis* were observed instead of the two branches, as in birds [1,3], rabbit [12], beaver [14] and mice [15].

Along its course, *v. cardiaca circumflexa sinistra* collects multiple the *vv. atriales* and *vv. ventriculares* from the atrium sinistrum and ventriculus sinister as indicated by the literature [1-4,22], respectively. One of the most interesting findings of the present study was that a vein of *vv. ventriculares* which was stronger than the others was found in samples. We thought that this vein was the same as left marginal ventricular vein in mammalian [5,12,14,17,20,21,23], according to its course.

In fowl, the *v. cardiaca media* is a prominent venous channel, which independently enters into the atrium dextrum [1]. In our samples, it opened into the *v. cava cranialis sinistra* as reported in rabbit [12,16], beaver [14], marsupial [15]. In birds [1,3,4], the *v. cardiaca media* is the strongest vein of the cardiac veins. Moreover, Bezuidenhout [3] reported that the *v. cardiaca media* is as a double vein in ostrich. In this study, the *v. cardiaca media* was weaker than the *v. cardiaca sinistra* and a single vein. This situation is similar to finding for rabbit [12], beaver [14] and marsupial [15]. Furthermore, in birds [1,3,4], rabbit [12,16], beaver [14] and marsupial [15], along its course, it was collected fine branches from the both ventricles, as in stork

In birds, the *vv. cardiacae dextrae* (*ventrales*) are constituted with the *vv. atriales*, *vv. ventriculares* and *vv. conalis* [1-3]. These veins are formed to the *v. cardiaca circumflexa dextra* that drains directly into the atrium dextrum in birds [1,3]. In the present study, the *vv. cardiacae dextrae* were constituted of a few *vv. ventriculares* opened into atrium dextrum separately. Thus, the *v. cardiaca circumflexa dextra* was not formed in stork, as in some fowl [1], beavers [14], mice [15] rabbits [17], marsupial [18]. In six specimens, one of veins collecting venous blood of the ventriculus dexter was stronger than the others. This vein was similar to vessel named as the right marginal ventricular vein in mammalian [5,12,20,21,23]. This strong vein has been also reported in mice [15] and marsupial [18].

In many mammals ^[5,12,14-17,19-21,23], main cardiac veins which are the great cardiac vein, middle cardiac vein, right cardiac veins and their secondary branches are well-defined in terms of the course they follow and the place they drain and that are present in a regular and specialised way, though they may display some variations. Unlike the mammals, as reported in the resources on birds ^[1,3,4,22] as well as in our samples, the size, number, distribution and origins of these secondary branches were observed to differ from sample to sample. For this reason, the identification of the secondary branches, except for these two strong veins, were not undertaken in our samples.

The *venae cardiacae minimae* carrying venous blood from myocardium to the heart chambers opened into the atrium dextrum ^[12,16] or to all of the heart chambers in mammalian ^[5,18,20]. In our samples, these veins were found constantly in the atrium dextrum and the which is similar to the findings reported by literature ^[1,3,12,16].

The venous drainage of the interventricular septum was provided mostly by the *vv. septales* of both the *pars interventricularis* of *v. cardiaca sinistra* and the *v. cardiaca media* in birds ^[1,3]. Although the *vv. cardiacae minimae* were made a small contribution to the venous drainage of the interventricular septum by means of the few thin, the *vv. septales* were primarily responsible for septal drainage in stork, which was similar to the cases in the fowl¹ and mammalian ^[12,20].

In conclusion, the venous drainage of the heart was consisted of the *v. cardiaca sinistra*, the *v. cardiaca media*, the *vv. cardiacae dextrae* and the *vv. cardiacae minimae*. The *v. cardiaca sinistra* was the largest vessel that drained of the heart. The origin and course of *v. cardiaca sinistra* and the *v. cardiaca media* of stork basically resembled the great cardiac vein and middle cardiac vein of the mammalian, respectively. One of the most interesting finding of the present study was that the two strong veins found in all samples, called the left marginal ventricular vein and right marginal ventricular vein, especially present in the mammalian. Because *v. cardiaca circumflexa dextra* was absent, veins formed the *vv. cardiacae dextrae* opened into atrium dextrum separately. The *vv. cardiacae minimae* emptied into both the atrium dextrum and the ventriculus dexter. There was no the coronary sinus.

REFERENCES

1. Lindsay FEF: The cardiac veins of *Gallus domesticus*. *J Anat*, 101 (3): 555-568, 1967.
2. Nomina Anatomica Avium (NAA): Second Edition. Publ. Nuttall Ornithological Club, No. 23. Cambridge, Mass, 1993.
3. Bezuidenhout AJ: The coronary circulation of the heart of the ostrich (*Struthio camelus*). *J Anat*, 138 (3): 385-397, 1984.
4. Nickel RA, Schummer A, Seiferle E: Anatomy of the Domestic Birds. Verlag Paul Parey, Berlin-Hamburg, 1977.
5. Nickel R, Schummer A, Seiferle E: The Anatomy of the Domestic Animals. Vol 3. The Circulatory System, the Skin and the Cutaneous Organs of the Domestic Mammals. Verlag Paul Parey, Berlin, 1981.
6. Yoldas A: A Macroanatomic Investigation on the Heart and Coronary Arteries of the Heart of the Ostrich. Thesis. Selcuk University, Institute of Medical Sciences. Turkey, Konya, 2007.
7. Charles S, Ahlquist JE: Phylogeny and Classification of Birds. New Haven, Yale University Press, 1990.
8. Neugebauer LA: De venis avium. *Nova Acta Acad. Caesar. Leop. Carol*, 13 (1): 521-697, 1845.
9. Kern A: Das Vogelherz. *Morph. Jb*, 56, 264-315, 1926.
10. Uchiyama T: Thebesian veins and sinuses of the hen's heart. *Morph. Jb*, 60, 296-322, 1929.
11. Atalar Ö, Yılmaz S, Dinç G, Özdemir D: The venous drainage of the heart in porcupines (*Hystrix cristata*). *Anat Histol Embryol*, 33, 233-235, 2004.
12. Yoldas A, Nur İH: The distribution of the cardiac veins in the New Zealand white rabbits (*Oryctolagus cuniculus*). *Iranian Journal of Veterinary Research*, 13 (3): 227-233, 2012.
13. Walker WF, Homberger DG: Anatomy and Dissection of the Rat. 3rd ed., pp.49-50, England, W.H. Freeman and Co., 1998.
14. Bisailon A: Gross anatomy of the cardiac blood vessels in North American beaver. *Anat Anz*, 150, 248-258, 1981.
15. Ciszek B, Skubiszewska D, Ratajska A: The anatomy of the cardiac veins in mice. *J Anat*, 211, 53-63, 2007.
16. Bahar S, Tipirdamaz S, Eken E: The distribution of the cardiac veins in Angora rabbits. *Anat Histol Embryol*, 36, 250-254, 2007.
17. Yadm ZA, Gad MR: Origin, course and distribution of the venae cordis in the rabbit and goat (comparative study). *Vet Med J*, 40, 1-8, 1992.
18. Dowd DA: The coronary vessels in the heart of a marsupial (*Trichosurus vulpecula*). *Am J Anat*, 140, 47-56, 1974.
19. Maric I, Bobinac D, Petkovic, M Dujmović: Tributaries of the human and canine coronary sinus. *Acta Anat (Basel)*, 156, 61-69, 1996.
20. Aksoy G, Özmen E, Kürtül İ, Özcan S, Karadağ H: The venous drainage of the heart in the Tuj sheep. *Kafkas Univ Vet Fak Derg*, 15 (2): 279-286, 2009.
21. Kabak M, Onuk B: Macroanatomic investigations on the venous drainage of the heart in Roe Deer (*Capreolus capreolus*). *Kafkas Univ Vet Fak Derg*, 18 (6): 957-963, 2012.
22. Kaupp BF: Anatomy of the Fowl. P.245. Philadelphia and London, W.B. Saunders, 1918.
23. Besoluk K, Tipirdamaz S: Comparative macroanatomic investigations of the venous drainage of the heart in Akkaraman sheep and Angora goats. *Anat Histol Embryol*, 30, 249-252, 2001.

Cirit Atlarında Vücut Ölçüleri ^{[1][2]}

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Özet

Bu araştırmada, Erzurum ilinde cirit kulüplerinde bulunan erkek Arap atlarının, vücut ölçüleri ve cirit oyunlarındaki koşu süreleri belirlenmiştir. Bu parametreler üzerine, atların performanslarına göre oluşturulan grupların, yaşı ve vücut kondisyon skorunun (VKS) etkisi araştırılmıştır. Ayrıca koşu süreleri ile vücut ölçüleri arasındaki fenotipik korelasyonlar da hesaplanmıştır. Cidago yüksekliği (154.19 ± 0.33), sağrı yüksekliği (154.27 ± 0.35) ve göğüs derinliği (67.01 ± 0.25) ortalama değerleri, diğer bazı at ırkları ve özellikle Arap atları için bildirilen değerlerden yüksek; sağrı uzunluğu (47.51 ± 0.25), ön göğüs genişliği (38.41 ± 0.21), göğüs çevresi (157.53 ± 0.58), baş uzunluğu (54.75 ± 0.20) düşük; ön incik çevresi (19.40 ± 0.14) ölçüsü ise benzer veya yüksek bulunmuştur. Atların yarış performansı ve cirit atı sahiplerinin tercihinine göre yapılan sınıflandırmada, en iyi cirit atlarının göğüs derinliği ve sağrı uzunluğunun daha yüksek olduğu belirlenmiştir. Atların yaşı arttıkça, ön göğüs genişliği ve ön incik çevresi ölçüsünde artış gözlenmiştir. VKS 3 ve 4 olan atlar arasında vücut ölçülerinde farklılık gözlenmemiştir. Vücut ölçüleri arasında önemli düzeyde fenotipik korelasyonlar bulunurken, koşu süresi ile incelenen vücut ölçüleri arasında ilişki tespit edilmemiştir. Koşu süreleri bakımından, VKS 3 olan atların (4.71 ± 0.080) VKS 4 olan atlardan (4.96 ± 0.048) daha hızlı koştuğu belirlenmiştir.

Anahtar sözcükler: At, Cirit, Vücut ölçüleri, Koşu süreleri, Vücut kondisyon skoru

Body Sizes of the Javelin Horses

Summary

In this research, the running times and body sizes of the male Arabian horses in Javelin Clubs of the Erzurum province were determined. The effects of the performance based groupings of horses, age and body condition score (BCS) were investigated on these parameters. The phenotypic correlations between the running times and body sizes were calculated. The average values of the withers height (154.19 ± 0.33), the height at the rump (154.27 ± 0.35) and the depth of the chest (67.01 ± 0.25) were found higher than those reported for some horse races, especially for the Arabian horses; the rump length (47.51 ± 0.25), width of the front chest (38.41 ± 0.21), chest circumference (157.53 ± 0.58) and the length of the head (54.75 ± 0.20) were found lower, and the front cannon bone circumference size (19.40 ± 0.14) was found similar or higher. Horses having higher chest depth and rump length were determined to be preferred for javelin sport in the classification made according to the running performance of the horses and the preferences of the breeders. Increases in the width of the front chest and the front cannon bone circumference were observed as the age of horses advanced. No differences in body sizes were observed between BCS 3 and 4 horses. While there were many significant correlations between the body sizes, no relationship was detected between the examined body sizes and the running times. When investigated according to the running times, it was determined that BCS 3 horses (4.71 ± 0.080) were running faster than BCS 4 horses (4.96 ± 0.048).

Keywords: Horse, Javelin, Body sizes, Run time, Body condition score

GİRİŞ

Türklerin en eski oyunlarından olan cirit ^[1] yakın zamana kadar Anadolu'nun birçok bölgesinde oynanmaktaydı. Ancak günümüzde sadece Erzurum, Uşak, Manisa, Sivas, Bayburt, Erzincan, Kars ve Malatya illerinde oynanmaktadır ^[2].

Eski yıllarda cirit atları Erzurum'da yetiştirilirken, günümüzde atlı cirit sporunda kullanılacak atlar, genellikle at yarışlarında başarılı olamayan veya çeşitli nedenlerle yarıştan çıkarılan erkek Arap atları arasından seçilmektedir ^[3].



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Atlarda tanımlayıcı bir ırk özelliği olan vücut ölçüleri, vücut yapısının uyumluluğunun değerlendirilmesinde [4] veya atların normal ve anormal gelişimin karşılaştırılmasında kullanılmaktadır [5]. Yarış atlarında vücut ölçüleri ile performansları arasında genetik korelasyon vardır. Atların seçiminde veya fiyatlarının belirlenmesinde de vücut ölçülerinden faydalanılabilir [6].

Atlarda vücut konformasyonu; kullanım yönü, genetik yapı ve beslenme durumuna göre şekillenmektedir [7,8]. Cirit atlarında, aniden hızlanma, durma ve dönme hareketleri düz koşu yapan yarış atlarına göre farklıdır [9]. Yarış atı, binek atı veya çekim atlarında vücut bölgelerinin gelişimi farklı olabilmektedir [10].

Farklı araştırmacılar tarafından Arap atları [4,6,11-18], farklı at ırkları [19-25] ve Türkiye’de yetiştirilen yerli atlardan alınan vücut ölçü değerleri [26-29] ile ilgili araştırmalar yürütülmüştür. Atlarda koşu hızı üzerine etkili faktörler üzerine çalışmalar da yapılmıştır [30-32]. Ancak cirit atlarında, vücut ölçüleri ile ilgili herhangi bir çalışmaya rastlanılmamıştır.

Erzurum bölgesinde yapılan bu çalışmada, cirit atlarının vücut ölçüleri ve cirit oyunlarındaki koşu süreleri belirlenerek, bu parametreler üzerine; anket bilgilerine ve cirit oyunlarında alınan derecelere göre oluşturulan skor gruplarının, yaşın ve vücut kondisyon skorunun etkisi araştırılmıştır.

MATERYAL ve METOT

Araştırma Erzurum merkezde 13 spor kulübüne kayıtlı, 94 erkek Arap atına ait veriler kullanılarak gerçekleştirilmiştir. Cirit atlarının büyük çoğunluğunun (%91.26) Arap atı olması ve cirit oyunlarında erkek atların (%100) tercih edilmesi nedeniyle, araştırma erkek Arap atları üzerinde yürütülmüştür. Cirit müsabakaları sırasında 40 adet ata ait düz koşu süresi ölçülmüştür.

Vücut ölçüleri, atlar beton ya da düz bir zemin üzerine getirilerek normal bir pozisyonda durmaları temin edildikten sonra alınmıştır. Vücut ölçülerinden cidago yüksekliği, sağrı yüksekliği, göğüs derinliği, ölçü bastonu ile (Hauptner®), sağrı uzunluğu ve ön göğüs genişliği ölçü pergeli (Elmark®) ile, sırt uzunluğu, beden uzunluğu, göğüs çevresi, boyun kalınlığı, baş uzunluğu, ön incik çevresi, carpal eklemin çevresi ve tarsal eklemin çevresi ölçü şeridi ile ölçülmüştür. Cirit yarışlarında atların koşu sürelerini belirlemek için tripot ile sabitlenmiş video kamera (Sony DCR-SR58®) kullanılmıştır. Atların koşu süreleri kronometre ile belirlenmiştir.

Aşağıda belirtildiği şekilde skor, yaş ve VKS grupları oluşturulmuştur.

Skor grupları: Cirit atı sahiplerinden elde edilen bilgiler ve Geleneksel Spor Dalları Federasyonunun düzenlemiş olduğu müsabakalarda alınan takım derecelerine göre

atları 3 gruba ayrılmıştır. Skor grupları belirlenirken üç farklı değerlendirmenin puan toplamaları dikkate alınmıştır: 1. değerlendirmede “kulübünüzdeki atlardan 6 tanesini iyiden kötüye sıralayınız” sorusuna verilen cevaba göre 1. sıradaki ata 6 puan, 6. sıradaki ata 1 puan verilmiştir. 2. değerlendirmede, “kulüp dışı atlardan 6 tanesinin isimlerini iyiden kötüye sıralayınız” sorusuna verilen cevaba göre 1. sıradaki ata 6 puan, 6. sıradaki ata 1 puan verilmiştir. Her atın aldığı toplam puan hesaplanarak 6’lı puanlamaya çevrilmiştir. 3. değerlendirmede ise yapılan resmi müsabakalar sunucunda birinci takımın bütün atları 3 puan, ikinci takım 2.5 puan, üçüncü takım 2 puan, dördüncü takım 1.5 puan, beşinci takım 1 puan alırken, sonuncu takımın bütün atları da 0.5’er puan almıştır. Bu üç değerlendirmeye ait puanların toplamı ile skor grupları oluşturulmuştur. Cirit atları, iyi performanstan, zayıf performansa sıralanarak, skor gruplarında toplam puanı 7-15 arası olan cirit atları 1. grupta (iyi), puanı 4-6.5 arası olanlar 2. grupta (orta) ve puanı 0-3.5 arası olanlar ise 3. grupta (zayıf) yer almıştır.

Yaş grupları: 4-5, 6-7, 8-9 ve 10 yaş ve üzeri yaştaki atlar yaş gruplarını oluşturmuştur.

Vücut kondisyon skoru grupları: Çalışmada cirit atlarına ait VKS, Carroll ve Huntington [33] tarafından bildirilen Avustralya sistemine göre 0 puan ile 5 puan arasında skorlama yapılarak belirlenmiştir. Cirit oyunlarında kullanılan atların VKS değerleri 3 ve 4 olarak tespit edilmiştir. VKS 3 olan atlar grup 3, VKS değeri 4 olan atlar ise grup 4 olarak tanımlanmıştır.

Cirit oyunları sırasında, atların rakip oyuncudan kaçtığı veya rakip oyuncuya yetişmek için kısa mesafede hızlanarak koştukları alan olan, cirit sahasındaki atış alanı çizgisi ile orta çizgi arasındaki mesafede (51 metre) 40 adet ata ait koşu süreleri ölçülmüştür. Koşu süreleri her cirit atına ait en az 10 kayıtlın ortalamaları alınarak değerlendirilmiştir.

Veriler skor grubu, yaş grubu, VKS grupları şeklinde tanımlanmıştır. Vücut ölçüleri ve koşu sürelerine etkili faktörler aşağıdaki matematik modelle değerlendirilmiştir.

$$Y_{ijkl} = \mu + a_i + b_j + c_k + e_{ijkl}$$

Bu modelde yer alan terimlerden;

Y_{ijkl} = i. skor grubu, j. yaş grubu, k. vücut kondisyon skoru gurubundaki 1. hayvana ait fenotipik değeri, μ = Populasyon ortalamasını, a_i = Skor etkisini (1, 2, 3), b_j = Yaşın etkisini (4-5, 6-7, 8-9, 10<), c_k = Vücut kondisyon skorunun etkisini (3, 4), e_{ijkl} = Şansa bağlı hatayı temsil etmektedir.

Verilerin istatistiksel analizi için SPSS 10.0 paket program [34] kullanılmıştır. Alt grup ortalamalarının karşılaştırılmasında Duncan çoklu karşılaştırma testi [35] uygulanmıştır. İncelenen özellikler arasındaki ilişkilerin belirlenmesinde parametrik doğrusal korelasyon tekniğinden (Pearson Korelasyonu) yararlanılmıştır [35].

BULGULAR

Erzurum ilindeki cirit atalarının farklı skor, yaş ve VKS gruplarına göre incelenen vücut ölçüleri ve koşu sürelerine ait ortalamaları ile çoklu karşılaştırma testi sonuçları *Tablo 1* ve *Tablo 2*'de, koşu süresi ve vücut ölçüleri arasındaki fenotipik korelasyonlar ise *Tablo 3*'te verilmiştir.

TARTIŞMA ve SONUÇ

Vücut Ölçüleri: Çalışmada atlarda cidago yüksekliği için bulunan ortalama değer (154.19 ± 0.33 cm); Andolusian atları [25] için belirtilen değerle benzerlik gösterirken, İspanyol [15], İran [6,16], Mısır Arap atları [4], Türkiye'de yetiştirilen Arap atları [13,17,18] ve yerli atlarda [26-29] tespit edilen değer-

lerden daha yüksek bulunmuştur. Ayrıca İngiliz aygırları [13], Noriker [20], İspanyol Andolusian [22], Finn [23] ve Murgese [19]. Mangalarga Marchador [21], Bardigrano atlarının [24] cidago yüksekliği bu çalışmadaki cirit atlarında tespit edilen değerden daha düşüktür. Bu durum cirit atlarının seçiminde daha yüksek cidagolu Arap atlarının tercih edilmesinin bir sonucudur. Adımların genişliği ve sırtın sağlamlığına yardım ettiğinden, hızlı yürüyüşlü binek atlarda uzun ve yüksek cidago istenir. Cidago alçak olduğunda, eyerin kayma ihtimali de çoktur [36]. Yaşın cidago yüksekliği üzerine etkisi; Van, Kars yerli [26,28], Ankara ve İzmir hipodromlarındaki Arap atlarında [13] bu araştırma bulgularına paralel olarak önemsiz ($P > 0.05$) bulunurken, Murgese atlarında [19] çok önemli, İngiliz atlarında [13], Çifteler, Anadolu, Karacabey, Mahmudiye ve Sultansuyu Tarım İşletmesi Arap atlarında [11,17,18], İran Arap atlarında [6] ise ista-

Tablo 1. Farklı skor, yaş ve VKS gruplarındaki cirit atlarına ait bazı vücut ölçüleri (cm)

Table 1. Body sizes of javelin horses at different score, age, and VKS groups (cm)

Grup ¹	Skor			Yaş				Vücut Kondisyon Skoru		Genel Ortalama
	1	2	3	4-5	6-7	8-9	10<	3	4	
N	32	27	35	29	29	22	14	33	61	
CY	154.58±0.57	154.35±0.58	153.65±0.50	155.17±0.55	154.04±0.53	154.35±0.63	153.21±0.80	154.04±0.56	154.34±0.38	154.19±0.33
SY	154.61±0.60	154.61±0.62	153.59±0.53	155.19±0.58 ^a	153.97±0.56 ^{ab}	155.09±0.66 ^a	152.84±0.84 ^b	154.51±0.59	154.04±0.40	154.27±0.35
SU	48.05±0.42 ^a	47.82±0.43 ^a	46.67±0.37 ^b	47.39±0.40	47.43±0.39	47.79±0.46	47.45±0.59	47.43±0.41	47.60±0.28	47.51±0.25
SIU	80.71±1.12	82.98±1.15	82.66±0.98	82.18±1.07	82.24±1.05	82.73±1.24	81.31±1.57	81.44±1.10	82.79±0.75	82.12±0.66
GD	67.96±0.43 ^a	66.56±0.44 ^b	66.50±0.38 ^b	67.44±0.41	67.57±0.40	66.41±0.48	66.61±0.61	66.99±0.43	67.03±0.29	67.01±0.25
GC	157.88±0.99	157.59±1.01	157.13±0.87	157.35±0.95	158.05±0.92	156.98±1.09	157.75±1.39	156.78±0.97	158.28±0.66	157.53±0.58
GG	38.62±0.36	38.68±0.37	37.91±0.31	37.90±0.34 ^b	38.00±0.33 ^b	38.53±0.40 ^{ab}	39.20±0.50 ^a	38.42±0.35	38.39±0.24	38.41±0.21
BEU	149.83±0.10	149.07±1.03	147.43±0.88	149.19±0.96	147.79±0.93	149.74±1.14	148.39±1.41	149.25±0.98	148.30±0.67	148.78±0.59
BK	84.35±1.05	83.64±1.08	81.51±0.93	84.26±1.01 ^{ab}	82.43±0.98 ^{ab}	81.11±1.17 ^b	84.86±1.48 ^a	83.68±1.04	82.66±0.70	82.17±0.62
BAU	55.16±0.35 ^a	54.01±0.36 ^b	55.07±0.31 ^a	55.04±0.33	54.84±0.32	54.22±0.38	54.89±0.49	54.69±0.34	54.80±0.23	54.75±0.20
IC	19.35±0.24	19.34±0.24	19.52±0.21	19.14±0.23 ^b	19.30±0.22 ^b	19.20±0.26 ^b	19.97±0.34 ^a	19.30±0.24	19.51±0.16	19.40±0.14
CC	30.64±0.31 ^{ab}	31.26±0.32 ^a	30.36±0.27 ^b	30.62±0.30	30.56±0.29	31.23±0.34	30.62±0.43	30.68±0.30	30.83±0.21	30.76±0.18
TC	40.96±0.40	40.77±0.41	40.34±0.35	41.08±0.38	40.62±0.37	40.63±0.44	40.42±0.56	40.75±0.39	40.63±0.26	40.69±0.23
BU/CY	0.97	0.97	0.96	0.96	0.96	0.97	0.97	0.97	0.96	0.96

^{a,b,c} Aynı satırda farklı harfleri taşıyan ortalamalar arası farklılıklar önemlidir ($P < 0.05$), ¹: Skor: 1= İyi, 2= Orta, 3= Zayıf, Vücut Kondisyon Skoru: 3= VKS 3, 4= VKS 4 olan ataları belirtmektedir, CY: Cidago yüksekliği, SY: Sağrı yüksekliği, SU: Sağrı uzunluğu, SIU: Sırt uzunluğu, GD: Göğüs derinliği, GC: Göğüs çevresi, GG: Ön göğüs genişliği, BEU: Beden uzunluğu, BK: Boyun kalınlığı, BAU: Baş uzunluğu, IC: Ön incik çevresi, CC: Carpal eklem çevre uzunluğu, TC: Tarsal eklem çevre uzunluğu, BU/CY: Beden uzunluğunun cidago yüksekliğine oranı

Tablo 2. Farklı skor, yaş ve VKS gruplarındaki cirit atlarına ait koşu süreleri (sn)

Table 2. Running time of javelin horses at different score, age, and VKS groups (sec)

Özellikler	N	Gruplar	X	SxX	(P<) ¹
Skor	14	1	4.70 ^b	0.067	*
	11	2	4.97 ^a	0.060	
	15	3	4.84 ^b	0.101	
Yaş	12	4-5	4.74	0.087	ÖS
	11	6-7	4.90	0.062	
	9	8-9	4.77	0.071	
	8	10<	4.93	0.098	
VKS	13	3	4.71	0.080	**
	27	4	4.96	0.048	

¹ ÖS: $P > 0.05$, * $P < 0.05$, ** $P < 0.01$, ^{a,b}: Aynı sütunda farklı harfleri taşıyan ortalamalar arası farklılıklar önemlidir ($P < 0.05$)

tistiksel olarak önemli olduğu saptanmıştır. Araştırmada, yaşın cidago yüksekliği üzerine etkisinin önemsiz çıkmasının sebebi bu çalışmada incelenen cirit atlarının gelişimini tamamlamış (4 yaş ve üzeri) olmasından kaynaklanmıştır.

Sağrı yüksekliği için bulunan ortalama (154.27 ± 0.35 cm) değer; Finn atlarında [23] bildirilen değerlere benzer olarak bulunurken, Mısır [4], İzmir Şirinyer ve Ankara 75. Yıl Hipodromundaki Arap aygırlarında [13], Anadolu, Karacabey, Sultansuyu, Mahmudiye Tarım İşletmelerinde yetiştirilen Arap atları [18], Mangalarga Marchador atları [21] ve bazı yerli atlarda [26-28] bildirilen değerlerden yüksek olarak saptanmış, TJK bünyesindeki İzmir Şirinyer ve Ankara 75. Yıl Hipodromundaki İngiliz aygırı [13] ve Noriker atlarından [20] ise düşük bulunmuştur. Bu sonuçlara göre Cirit oyunlarında sağrı yüksekliği daha fazla olan Arap atların tercih

Tablo 3. Koşu süresi ve vücut ölçüleri arasındaki fenotipik korelasyonlar**Table 3.** Phenotypic correlations between measurements and running time

Özellikler	KS	CY	SY	SU	SIU	GD	GC	GG	BEU	BK	BAU	IC	CC
CY	-0.23												
SY	-0.30	0.70**											
SU	-0.11	0.11	0.17										
SIU	-0.08	0.25	0.39*	0.02									
GD	-0.30	0.30	0.23	-0.04	0.39*								
GC	-0.05	0.17	0.19	0.24*	0.45**	0.38*							
GG	-0.14	0.08	0.10	0.35*	0.09	-0.10	0.13						
BEU	-0.21	0.48**	0.62**	0.47**	0.5**	0.43**	0.56**	0.24					
BK	-0.07	0.23	0.36*	0.18	0.02	0.24	0.30	0.18	0.22				
BAU	0.06	0.28	-0.01	-0.05	-0.18	0.25	0.14	0.22	0.20	0.18			
IC	-0.25	0.21	0.15	0.17	0.17	0.46**	0.32*	0.02	0.19	0.35*	0.26		
CC	-0.30	0.28	0.19	0.27	0.22	0.46**	0.41**	0.11	0.41**	0.17	0.15	0.73**	
TC	-0.17	0.23	0.14	0.10	0.42**	0.54**	0.34*	0.17	0.50**	0.28	0.18	0.34*	0.50**

** P<0.01, * P<0.05, CY: Cidago yüksekliği, SY: Sağrı yüksekliği, SU: Sağrı uzunluğu, SIU: Sırt uzunluğu, GD: Göğüs derinliği, GC: Göğüs çevresi, GG: Ön göğüs genişliği, BEU: Beden uzunluğu, BK: Boyun kalınlığı, BAU: Baş uzunluğu, IC: Ön incik çevresi, CC: Carpal eklem çevre uzunluğu, TC: Tarsal eklem çevre uzunluğu, KS: Koşu süresi

edildiği anlaşılmaktadır. Sağrı yüksekliğine yaştan etkisi Van yerli atlarında [28], Anadolu, Karacabey, Mahmudiye Tarım İşletmelerinde yetiştirilen Arap atlarında [18] elde edilen, araştırma bulgularına benzer olarak önemli (P<0.05) bulunurken, Ankara ve İzmir hipodromlarında yapılan çalışmada Arap (3-4 yaş) ve İngiliz (3-5 yaş) atlarında ise önemsiz (P>0.05) bulunmuştur [13]. Alarların [28] belirttiği gibi 10 yaş ve üzeri atlarda yaştan ilerlemesine bağlı olarak sağrı yüksekliğinde düşüş gözlenmiştir.

Sağrı uzunluğu için tesbit edilen 47.51±0.25 cm değer İspanyol Arap atları [15] için bildirilen değere benzer, Mangalarga Marchador atlarında [21] bildirilenden ise düşüktür. Cirit atlarında sağrı uzunluğu farklı skor gruplarında istatistik olarak önemli (P<0.05) farklılık göstermiştir. En uzun sağrı atların birinci ve ikinci skor grubunda bulunduğu belirlenmiştir. Sağrının uzun olması üzerindeki kasların uzun olmasını ve hayvanın daha hızlı koşmasını sağlar [10]. Cirit oyunlarında uzun sağrı atlar tercih edilmektedir.

Göğüs derinliği için bulunan ortalama değer (67.01±0.25 cm) bazı yerli [26-29] ve İspanyol Andolisan [22] atlarında bildirilen değerlerden yüksek olarak bulunurken; Sultansuyu ve Mahmudiye Tarım İşletmelerinde yetiştirilen Arap atlarında [18] bildirilen değerlere benzer, Anadolu, Karacabey Tarım İşletmesi Arap atları [18], Mısır'daki Arap aygırları [4] ve Noriker atlarında [20] bildirilen değerlerden ise düşük olduğu tespit edilmiştir. Skor grupları incelendiğinde göğüs derinliğinin, en iyi atların bulunduğu birinci skor grubunda daha yüksek olduğu tespit edilmiştir. Akciğer ve kalbin gelişme derecesi, göğüs boşluğunun büyüklüğü ile ilişkili olduğu için göğüsün gelişme derecesi önemlidir [7,11]. Cirit atlarının seçiminde göğüs derinliği gelişmiş atlar tercih edilebilir. Göğüs derinliğine yaştan etkisi Van yerli atlarında [28] elde edilen sonuçta benzerlik göstererek önemsiz (P>0.05) bulunurken, Kars yerli [26] (P<0.05) ve farklı tarım işletmelerinde yetiştirilen Arap atlarında [18], istatistiksel olarak önemli (P<0.01) olduğu belirtilmiştir.

Çalışmada 157.53±0.58 cm olarak tesbit edilen göğüs çevresi ölçüsü, bazı yerli atlarda [26,28] bildirilen değerlerden daha yüksek, Anadolu, Karacabey, Mahmudiye [18] ve Sultansuyu [17,18] Tarım İşletmelerinde yetiştirilen Arap atlarında, Mısır'da yetiştirilen [4] ve yaşı büyük olan Arap aygırlarında [11], Noriker [20], Van yerli [27], Murgese [19] ve Bardigrano atlarında [24] bildirilen değerden ise daha düşük bulunmuştur. Göğüs çevresi bakımından bu çalışmadaki skor, yaş ve VKS gruplarında istatistik farklılık önemsizdir (P>0.05). Arap atlarında (3-4 yaşlı) [14], Kars yöresi yerli atlarda [26] ve Sultansuyu Tarım İşletmesi Arap atlarında [17] yaştan etkisinin önemli (P<0.05) olduğu bildirilmiştir. Göğüs çevresinin genişliği, içindeki organların gelişme durumu ve büyüklüğü hayvanların sağlığı ve verimleri üzerine önemli derecede etkilidir [7].

Ortalama ön göğüs genişliği değeri (38.41±0.21); Kars yerli atlarına [26] benzer tespit edilirken; İspanyol Arap [15] ve Türk Rahvan atlarından [15] yüksek olarak bulunmuş, Mısır Arap aygırlarından [4], Noriker [20], Mangalarga Marchador [21] ve Van yerli atlarından [27] düşük bulunmuştur. Cirit atlarında ön göğüs genişliği incelendiğinde yaş gruplarındaki fark istatistiksel olarak önemli (P<0.05) bulunmuştur. On ve üzeri yaştaki atlarda ön göğüs genişliğinin daha yüksek olduğu tespit edilmiştir.

Beden uzunluğu için ortalama değer (148.78±0.59 cm), Mısır'da yetiştirilen Arap aygırlarına [4], İspanyol Arap atlarına [15] benzerlik gösterirken, yerli atlar [26-29] için bildirilen değerlerden daha yüksek bulunmuştur. Diğer yandan bu çalışmada beden uzunluğu için bildirilen değerin İran Arap atları [16], İzmir Şirinyer ve Ankara 75. Yıl hipodromundaki Arap aygırları [13], farklı tarım işletmelerinde yetiştirilen Arap atları [18], İngiliz aygırları [3], Noriker [20], Mangalarga Marchador [21], İspanyol Andolisan [22] ve Finn atları [23] için bildirilen değerden ise düşük olduğu saptanmıştır. Beden uzunluğu ortalamaları arasındaki farklılıklar yaş, ırk, bölge ve işletme farklılıklarından kaynaklanabilir. Cirit

atlarında skor grupları arasında beden uzunluğu bakımından fark gözlenmemesine ($P>0.05$) rağmen cirit atlarının beden uzunlunun yerli atlar hariç birçok Arap atı ve farklı at ırklarından düşük olması, cirit atı seçiminde beden uzunluğu fazla olan atların tercih edilmediğini göstermektedir. Cirit atlarında beden uzunluğunun cidago yüksekliğine oranı ortalama 0.96 bulunmuştur bu değer farklı tarım işletmelerinde yetiştirilen Arap atları^[18] için bildirilen değerlerden (1.01-1.04) düşüktür. Bu durum cirit atlarında cidago yüksekliği fazla, beden uzunluğu daha az olan Arap atlarının tercih edildiğini göstermektedir. İncelenen beden uzunluğu değerleri için skor, yaş ve VKS gruplarında istatistiksel olarak farklılık belirlenmemiştir ($P>0.05$). Yaşın beden uzunluğu üzerine etkisi, bu araştırma bulgularına paralel olarak; Van yerli atları^[28], Arap ve İngiliz atlarında^[13] istatistiksel olarak önemsiz ($P>0.05$) bulunurken, Altındere^[27] ve Sultansuyu^[12,17] tarım işletmelerinde yetiştirilen atlarda önemli ($P<0.05$) farklı tarım işletmelerinde yetiştirilen Arap atlarında^[18] çok önemli ($P<0.01$) bulunmuştur. Safkan Arap atlarının 4 yaşında büyümelerini tamamladıkları belirtilmiştir^[18]. Yaşın etkisinin önemsiz çıkması araştırmada yer alan atların 4 yaş üzeri olmasından kaynaklanabilir.

Atlarda boyun kalınlığı ortalama 82.17 ± 0.62 cm'dir. Boyun kalınlığı yaş gruplarında istatistiksel olarak farklı bulunmuştur ($P<0.05$). Boyun kalınlığı 10 ve üzeri yaştaki atlarda en yüksek değeri almıştır. Boyun kalınlığı incelendiğinde skor ve VKS gruplarında istatistiksel farklılık saptanmamıştır ($P>0.05$). Boyunun kendisi kuru, kasları iyi gelişmiş ve yağlanmamış olmalıdır, bu boyun şekli bütün verim yönleri için elverişli sayılmaktadır^[36].

Baş uzunluğu (54.75 ± 0.20 cm) için ortalama değer; İspanyol Arap atları^[15], Kars yöresindeki yerli atlarında bildirilen değerlere^[26] benzer; İran Arap atları^[6], Noriker atları^[20], Mangalarga Marchador atları^[21], Uzunyayla atları ve Türk Rahvan atında^[29] bildirilen değerden ise düşük olduğu tespit edilmiştir. Başın yapısı hayvanların ırkı, verim ve hizmet yönüne göre değişir. Cirit atlarında baş uzunluğunun, skor gruplarında istatistiksel olarak farklı olduğu tespit edilmiştir ($P<0.05$). Baş uzunluğunun, 1. skor grubunda en yüksek değere sahip olduğu belirlenmiştir. Kars yöresindeki değişik yaş ve cinsiyetteki Türk yerli atlarda yapılan çalışmada baş uzunluğu üzerine yaşın etkisinin istatistiksel olarak önemli ($P<0.05$) olduğu belirlenmiştir^[32]. Ancak bu çalışmada, farklı yaştaki atların baş uzunlukları arasında istatistiksel olarak fark bulunmamıştır ($P>0.05$). Başın yapısı yarış atlarında genelde asil yani küçük, zarif yapılı ve kuru; ağır atlarda ise iri, kaba ve etlidir^[7].

Bu çalışmada ön incik çevresi için tespit edilen değer (19.40 ± 0.14 cm); Safkan Arap atı için bildirilen bazı değerlere benzer^[11,15,17,18], bazı değerlerden^[4,17] ise yüksektir. Bu durum cirit atlarında ön incik çevresi bakımından değişikliğin olmadığını göstermektedir. Diğer yandan, bu çalışmada ön incik çevresi içi bulunan değer, yerli atlar için bildirilen değerlerden yüksek iken^[26-29], bazı yabancı at ırkları^[19,20,22,24]

için bildirilen değerlerden ise düşüktür. Ön inciğin kısa ve geniş olması dayanıklılığı artıracığından aranılan özelliiktir^[36]. Yaş gruplarında ön incik çevresi incelendiğinde istatistiksel farklılık önemli bulunmuş olup ($P<0.05$), 10 ve üzeri yaştaki atların en kalın inciğe sahip olduğu saptanmıştır. Bu araştırma bulgularına paralel olarak; Çifteler^[11] ve Sultansuyu^[17] Tarım İşletmesinde yetiştirilen Türk Arap atlarında yaşın etkisinin önemli ($P<0.05$) olduğunu bildirilmiştir. Diğer yandan bu araştırma bulgularından farklı olarak Kars yöresi yerli atlarda yapılan çalışmada ön incik çevresi üzerine yaşın etkisinin önemsiz ($P>0.05$) olduğu belirlenmiştir^[26].

Koşu Süreleri: Atların 51 metrelik mesafeyi ortalama 4.83 ± 0.048 sn'de koştuğu tespit edilmiştir. En çok tercih edilen atların bulunduğu 1.skor grubundaki atların koşu süreleri 3. skor grubundakilere benzer, 2. skor grubundakilerden ise düşüktür. Atların koşu hızı, atın yaşı, eğitmeni, binicisi, hava şartları, hipodromun durumu, taşınan ağırlık ve atın sağlık durumu gibi birçok çevresel faktöre bağlı olabilir^[31]. Koşu süreleri arasında varyasyonun düşük olması nedeniyle hayvan sayısının veya koşu mesafesinin artırılması isabetli olacaktır. At yarışlarında yarış hızının müsabakada ya da sezonda elde ettiği başarıyı ifade ettiği belirtilmektedir^[32]. Cirit oyunlarında atların kısa sürede hızlanarak rakip cirit oyuncusundan kaçması veya yakalaması için koşu hızı önemlidir. Pilarski ve ark.^[30] yaşın, atların hızı üzerine etkisinin olduğunu belirtmiş ancak bu çalışmada farklı yaş gruplarında koşu süreleri bakımından farklılık istatistiksel olarak önemsiz ($P>0.05$) çıkmıştır.

Vücut kondisyon skoru 3 olan atların hızı, VKS 4 olan atlardan yüksek çıkmıştır ($P>0.01$). Avustralya sistemine göre de VKS 3 olan atların performansları yüksektir^[37]. Cirit atlarının dengeli bir şekilde beslenerek cirit sezonu öncesi ve sonrası VKS'nun 4 yerine 3'te tutulması daha yüksek performans sağlayabilir.

Fenotipik Korelasyonlar: Cirit atlarından alınan vücut ölçüleri arasındaki fenotipik korelasyon değerlerinden cidago yüksekliği ile sağrı yüksekliği, beden uzunluğu ve boyun uzunluğu arasında istatistiksel olarak önemli veya çok önemli pozitif ilişki bulunmuştur. Benzer şekilde Antalya^[14] yapmış olduğu çalışmada cidago yüksekliği ile sağrı yüksekliği (0.59) arasında çok önemli ilişkinin olduğunu bildirmiştir.

Cidago yüksekliği ile göğüs derinliği (0.302), göğüs genişliği (0.077) ve göğüs çevresi (0.171) arasında istatistik olarak bir ilişki bulunmamasına rağmen Bayram ve ark.^[27] ile Kırmızıbayrak ve ark.^[26] söz konusu vücut ölçüleri ile cidago yüksekliği arasında önemli ve çok önemli derecelerde ilişkilerin bulunduğunu saptamışlardır. Cidago yüksekliği ile incik çevresi (0.212) arasında belirlenen korelasyon istatistik olarak önemli bulunmazken, Antalya^[14], Bayram ve ark.^[27], Suantama ve ark.^[23] ve Sadek ve ark.^[4] önemli ve çok önemli derecede korelasyonlar tespit etmişlerdir.

Yapılan çalışmada sağrı yüksekliği ile beden uzunluğu (0.620) arasında istatistiksel olarak çok önemli ($P<0.01$) düzeyde pozitif korelasyon bulunmuştur. Kırmızıbayrak ve ark.^[26] ile Suontama ve ark.^[23] bu çalışmaya benzer olarak bu ilişkiyi istatistiksel olarak çok önemli bulmuşlardır. Sağrı yüksekliği ile göğüs genişliği (0.101) arasındaki ilişki Bayram ve ark.^[27] bulgularına benzer olarak istatistiksel olarak önemsiz bulunmuştur.

Antalyalı^[14] ile Sadek ve ark.^[4] bu çalışmaya benzer olarak beden uzunluğu ile göğüs çevresi arasındaki korelasyonu istatistiksel olarak önemli bulmuşlardır.

İncik çevresi ile göğüs derinliği (0.458) arasında çok önemli ($P<0.01$), boyun kalınlığı (0.346) ve göğüs çevresi (0.324) arasında önemli ($P<0.01$) pozitif bir korelasyon olduğu belirlenmiştir. Antalyalı^[14] incik çevresi ile göğüs çevresi (0.075) arasındaki ilişkiyi istatistiksel olarak önemsiz bulmuştur. Bayram ve ark.^[27] incik çevresinin göğüs derinliği (0.11) ve göğüs çevresi (0.28) arasındaki ilişkiyi önemli ve çok önemli olduğunu belirtmişlerdir. Suontama ve ark.^[23] incik çevresi ve göğüs çevresi (0.58) arasındaki korelasyonu istatistiksel olarak çok önemli düzeyde bulmuştur. İncik çevresi ile carpal eklem çevre uzunlukları arasındaki yüksek dereceli pozitif ilişki (0.74) cirit atı seçiminde incik çevresi ölçüsü yanında carpal eklem çevre ölçüsünün de dikkate alınabileceğini göstermektedir.

Sonuç olarak cirit atları seçiminde yüksek cidago ve sağrılı, beden uzunluğu daha kısa olan Arap atları tercih edilmektedir. Cirit oyunlarında performansı iyi olan atlarda, göğüs derinliği ve sağrı uzunluğu daha fazladır. Cirit atı seçiminde bu ölçüler dikkate alınmalıdır. Cirit oyunlarında atların VKS'sinin 3 olması, performanslarını olumlu etkileyebilir. Cirit atlarında vücut ölçüleri arasındaki fenotipik korelasyonlar, bu karakterlerin biri bakımından üstün değerli bireylerin seçilmesi ile seçilen bireylerde ikinci karakterin ne durumda olacağı hakkında fikir vermesi açısından önemlidir.

KAYNAKLAR

1. Karaca Ş: Türk ve İslam aleminde at ve ata binicilik. *Yüksek Lisans Tezi*, Selçuk Üniv. Sosyal Bil. Enst., 1998.
2. Anonim: <http://www.gelenekselfedgovtr/>, Erişim tarihi: 08.01.2010.
3. Aka ST: Atlı cirit sporcularının sosyal beceri düzeylerinin çeşitli değişkenlere göre incelenmesi. *Yüksek Lisans Tezi*, Karadeniz Teknik Üniv. Sosyal Bil. Enst., 2010.
4. Sadek M, Al-Aboud A, Ashmawy A: Factor analysis of body measurements in Arabian horses. *J Anim Breed Genet*, 123 (6): 369-377, 2006.
5. Anderson T, McIlwraith C: Longitudinal development of equine conformation from weanling to age 3 years in the Thoroughbred. *Equine Vet J*, 36 (7): 563-570, 2004.
6. Gharahveysi S, Khashan N, Gerami A, Torshizi RV: Estimation of genetic parameters on conformation traits of the Iranian Arab horses population. *Pakistan J Biol Sci*, 11 (2): 280-284, 2008.
7. Akçapınar H, Özbeyaz C: Hayvan Yetiştiriciliği Temel Bilgileri. Kariyer Matbaacılık, Ankara, 1999.
8. Akçapınar H, Atasoy F, Gücüyener Ö: Atlarda Eksteriör ve Önemi, *Ulusal Atçılık Sempozyumu* 18-25 Eylül, Ankara, Özet Kitabı, s. 34-36, 2005.
9. David WF: Physical Conditioning of Horses. *ANSI-3915 Division of Agricultural Sciences and Natural Resources Oklahoma State University*, <http://pods.dasn.okstate.edu/docushare/dsweb/Get/Document-2062/ANSI-3915web.pdf>, Accessed: 08.01.2012.

pdf, Accessed: 08.01.2012.

10. Özbeyaz C, Akçapınar H: At Yetiştiriciliği Ders Notları. Ankara Üniv. Veteriner Fakültesi, Ankara, 2003.
11. Doğan İ, Akcan A, Koç M: Safkan erkek ve dişi Arap taylarda önemli beden ölçülerinin incelenmesi. *Türk J Vet Anim Sci*, 26, 55-60, 2002.
12. Özdemir B: Sultansuyu Tarım İşletmesi'nde yetiştirilen Arap atların dölvörümü, yaşama gücü ve beden ölçüleri. *Doktora Tezi*, Uludağ Üniv. Sağlık Bil. Enst., 1998.
13. Yıldırım İG: Atlarda genel vücut yapısının morfometrik yöntemlerle incelenmesi. *Yüksek Lisans Tezi*, Adnan Menderes Üniv. Sağlık Bil. Enst., 2007.
14. Antalyalı A: Türk safkan Arap atlarında bazı vücut özelliklerinin fenotipik ve genotipik parametreleri. *Doktora Tezi*, Ankara Üniv. Sağlık Bil. Enst., 2008.
15. Cervantes I, Baumung R, Molina A, Druml T, Gutiérrez J, Sölkner J: Size and shape analysis of morphofunctional traits in the Spanish Arab horse. *Livest Sci*, 125 (1): 43-49, 2009.
16. Gharahveysi S, Manafiazar, G., Mirhabibi, S: Genetic correlation of conformation and race performance traits in Iranian Arab horse population. *Global Veterinaria*, 4 (5): 510-514, 2010.
17. Kaygısız A, Orhan H, Vanlı Y, Güler A, Gökdere MA: Sultansuyu Tarım İşletmesi'nde yetiştirilen Türkiye Arap atlarının vücut ölçülerine ait fenotipik ve genetik parametre tahminleri. *İğdır Üniv Fen Bil Enst Derg*, 1 (1): 69-74, 2011.
18. Gücüyener Hacan Ö, Akçapınar H: Farklı hararlarda yetiştirilen safkan Türk Arap atlarında bazı fenotipik ve genetik parametreler. I Vücut ölçüleri ve kalıtım dereceleri. *Lalahan Hay Araşt Enst Derg*, 51 (2): 55-70, 2011.
19. Dario C, Carnicella D, Dario M, Bufano G: Morphological evolution and heritability estimates for some biometric traits in the Murgesse horse breed. *Genet Mol Res*, 5 (2): 309-314, 2006.
20. Druml T, Baumung R, Sölkner J: Morphological analysis and effect of selection for conformation in the Noriker draught horse population. *Livest Sci*, 115 (2-3): 118-128, 2008.
21. Pinto LFB, Almeida FQ, Quirino CR, De Azevedo PCN, Cabral GC, Santos EM: Evaluation of the sexual dimorphism in Mangalarga Marchador horses using discriminant analysis. *Livest Sci*, 119 (1-3): 161-166, 2008.
22. Gómez M, Valera M, Molina A, Gutiérrez J, Goyache F: Assessment of inbreeding depression for body measurements in Spanish Purebred (Andalusian) horses. *Livest Sci*, 122 (2-3): 149-155, 2009.
23. Suontama M, Saastamoinen M, Ojala M: Estimates of non-genetic effects and genetic parameters for body measures and subjectively scored traits in Finnish trotters. *Livest Sci*, 124 (1-3): 205-209, 2009.
24. Sabbioni A, Beretti V, Zanon A, Pagani G, Filippini S, Superchi P: Morphological evolution of Bardigiano horse. *Ital J Anim Sci*, 4 (2s): 412-414, 2010.
25. Munoz A, Trigo P, Riber C, Malonda V, Castejon F: A study of serum insulin-like growth factor type 1 (IGF-1) concentrations in resting untrained Andalusian horses: influence of age and gender. *Veterinary Medicine (Prague)*, 56 (5): 231-242, 2011.
26. Kırmızıbayrak T, Aksoy A, Tilki M, Saatçı M: Kars yöresi Türk yerli atlarının morfolojik özelliklerinin incelenmesi. *Kafkas Univ Vet Fak Derg*, 10 (1): 69-72, 2004.
27. Bayram D, Öztürk Y, Küçük M: Van yöresinde yetiştirilen atlarda fenotipik özellikler. *YYU Vet Fak Derg*, 16 (1): 85-88, 2005.
28. Alarslan E: Van ili Erçiş ilçesi Ulupamir köyündeki geleneksel at yetiştiriciliğinin yapısal özellikleri. *Yüksek Lisans Tezi*, Yüzüncü Yıl Üniv. Fen Bil. Enst., 2009.
29. Caglayan T, Inal S, Garip M, Coskun B, Inal F, Gunlu A: The determination of situation and breed characteristics of Turkish rahvan horse in Turkey. *J Anim Vet Adv*, 9 (4): 674-680, 2010.
30. Pilarski W, Swiezynski K, Kobryn H, Kobrynczuk F, Radomski L: Attempt to qualify the horse's sport performance on the basis of biometric criteria. *Annals of Warsaw Agricultural University SGGW AR Veterinary Medicine*, 18, 13-18, 1993.
31. Ekiz B, Kocak Ö, Yılmaz A: Phenotypic and genetic parameter estimates for racing traits of thoroughbred horses in Turkey. *Arch Tierzucht*, 48, 121-129, 2005.
32. Gür E: Atlarda yarış öncesi dönemde kalbin gelişimi, maksimal frekansı ve koşu hızı ile atlatık potansiyeli arasındaki ilişkinin incelenmesi. *Doktora Tezi*, İstanbul Üniv. Sağlık Bil. Enst., 2010.
33. Carroll C, Huntington P: Body condition scoring and weight estimation of horses. *Equine Vet J*, 20 (1): 41-45, 1988.
34. SPSS: SPSS for Windows Release 10.0. 1997.
35. Yıldız N, Akbulut Ö, Bircan H: İstatistiğe Giriş. 3. Baskı, Aktif Yay., İstanbul, 2002.
36. Yarkın İ: Atçılık. Ankara Üniv. Zir. Fak. Yayınları, Yayın No:40. 1962.
37. Garlinghouse SE, Burrill MJ: Relationship of body condition score to completion rate during 160 km endurance races. *Equine Vet J*, 31 (30s): 591-595, 1999.

Effect of Varieties on Potential Nutritive Value of Pistachio Hulls

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Summary

The aim of this study was to determine the potential nutritive value of pistachio hulls obtained from six different varieties using chemical composition and *in vitro* gas production technique. *In vitro* gas productions of pistachio hulls were determined at 0, 3, 6, 12, 24, 48, 72 and 96 h incubation times and their gas production kinetics were described. There is considerable variation among pistachio hulls obtained from six different varieties in terms of chemical composition and *in vitro* gas production, metabolisable energy (ME) and organic matter digestibility (OMD). Dry matter (DM) contents of pistachio hulls ranged 26.45 to 29.25%. Ash content of pistachio hulls ranged from 8.50 to 19.86%. The crude protein (CP) contents of pistachio hulls ranged 7.27 to 14.99%. The hulls from Kırmızı, Ohadi and Keten Gömleği (KG) had significantly higher CP contents than the others. The NDF and ADF contents of pistachio hulls ranged from 18.25 to 22.49% and 14.32 to 18.29% respectively. The pistachio hulls from Ohadi and Uzun had significantly higher NDF and ADF contents than the others. The CT contents of pistachio hulls ranged from 2.03 to 2.63%. The hulls from Beyaz Ben (BB) had significantly higher CT content than that of KG. The gas production rate ranged from 0.071 to 0.107%. The gas production rate of pistachio hulls from Sultani and KG were significantly higher than the others. The potential gas production of pistachio hulls from BB and Ohadi were significantly higher than the others. The ME and OMD contents of pistachio hulls ranged from 9.76 to 11.05 MJ/kg DM and 69.00 to 74.50% respectively. The ME contents of pistachio hulls from BB and Ohadi were significantly higher than those of Kırmızı, Uzun and KG whereas the OMD contents of pistachio hulls from Ohadi were significantly higher than those of pistachio hulls for Kırmızı, Sultani, Uzun and KG. In conclusion, chemical characterization with CP, OMD and ME suggests that the pistachio hulls had moderate level of CP concentration and was quite digestible therefore it can be said that pistachio hulls will provide feedstuffs of an acceptable quality for ruminant animals.

Keywords: Chemical composition, Condensed tannin, Digestibility, Nutritive value, *In vitro* gas production, Pistachio hulls

Varyetenin Antep Fıstığı Dış Kabuğunun Potansiyel Besleme Değerine Etkisi

Özet

Bu çalışmanın amacı, farklı varyetelerden elde edilen Antep fıstığının dış kabuğunun potansiyel besleme değerini kimyasal kompozisyonu ve *in vitro* gaz üretim tekniğini kullanarak belirlemektir. *In vitro* gaz ölçümleri fermentasyonun başlamasından sonra 3, 6, 12, 24, 48, 72 ve 96 saatlerinde yapılmıştır. Farklı varyetelerden elde edilen Antep fıstığı kabuğunun kompozisyonları, *in vitro* gaz üretimi, metabolik enerji içeriklerinde ve organik madde sindirim dereceleri arasında önemli varyasyonlar vardır. Antep fıstığı kabuklarının kuru madde içeriği %26.45 - %29.25 arasında değişmiştir. Kül içeriği ise %8.50 - %19.86 arasında değişmiştir. Antep fıstığı kabuklarının ham protein içeriği %7.27 - %14.99 arasında değişmiştir. Kırmızı, Ohadi ve keten gömleği varyeteleri en yüksek protein içeriğine sahip olmuştur. Antep fıstığı kabukların NDF ve ADF içerikleri sırasıyla %18.25 - %22.49 ve %14.32 - %18.29 arasında değişmiştir. Ohadi ve Uzun varyeteleri en yüksek NDF ve ADF içeriğine sahip olmuştur. Antep fıstığı kabukların kondense tanen içerikleri %2.03 - %2.63 arasında değişmiş olup Beyaz Ben varyetesi en yüksek tanen içeriğine sahip olmuştur. Antep fıstığı kabukların gaz üretim hızı %0.071 ile %0.107 arasında değişmiş olup en yüksek gaz üretim hızına Sultani ve Keten Gömleği varyeteleri sahip olmuşlardır. Antep fıstığı kabukların potansiyel gaz üretim miktarları 65.92 ile 73.46 ml arasında değişmiş olup en yüksek gaz üretim miktarına Beyaz Ben ve Ohadi varyeteleri sahip olmuştur. Antep fıstığı kabukların metabolik enerji ve organik madde sindirim dereceleri sırasıyla 9.76 ile 11.05 MJ/kg KM, %69.00 - %74.50 arasında değişmiştir. Beyaz Ben ve Ohadi varyetelerinden elde edilen kabukların metabolik enerji içerikleri Kırmızı, Sultani, Uzun ve Keten Gömleği varyetelerinden elde edilen kabukların metabolik enerjisi içeriğinden yüksek olmasına rağmen Ohadi varyetesiinden elde edilen Kırmızı, Sultani, Uzun ve Keten Gömleği varyetelerinden elde edilen kabuklarının OMD değerinden yüksek bulunmuştur. Sonuç olarak, kimyasal olarak ham protein, OMD ve ME bakımından yapılan karakterizasyon Antep fıstığı kabuklarının orta seviyede ham protein içerdiği ve oldukça sindirilebilir olduğundan dolayı ruminantlar için kabul edilebilir kalitede yem sağlayacağı söylenebilir.

Anahtar sözcükler: Kimyasal kompozisyon, Kondense tanen, Sindirim derecesi, Besleme değeri, *In vitro* gaz üretimi, Fıstık dış kabuğu



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INTRODUCTION

After processing of fruits, vegetables, crops and nuts, considerable amount agricultural byproduct become available and used to meet the energy and protein requirements of livestock animal in most parts of the world especially during the dry season when food shortage occurs^[1]. There is about 2.212.229 hectares of pistachio garden in Turkey and annual dry pistachio production is 128000 tones^[2]. Pistachio fruit is well known for its oleaginous and edible seed. The hull of pistachio is constituted by epicarp and mesocarp which adheres tightly the hard inner shell until the pistachio nut is ripe^[3]. De-hulling of pistachio after harvest produces considerable amount hulls which are dried or ensiled for storage. Several researches showed that dried or ensiled pistachio by product can be used ruminant diets to some extent^[1-6]. Although there is considerable amount pistachio hull production in Turkey there is no information about chemical composition, digestibility, and metabolic energy contents of pistachio hulls obtained from different varieties. Chemical composition, in combination with *in vitro* gas production, OMD and ME content were widely used to determine the potential nutritive value of feedstuffs which are previously limited or un-investigated^[7-10]. The aim of this study was to determine the potential nutritive value of pistachio hulls obtained from six different varieties using chemical composition and *in vitro* gas production technique.

MATERIAL and METHODS

This experiment was conducted in the laboratory of Department of Animal Science, Faculty of Agriculture, University of Kahramanmaraş Sutcu Imam, Kahramanmaraş, Turkey. Studies performed using *in vitro* experimental model was approved by the Animal Experimentation Ethics Committee of University of Kahramanmaraş Sutcu Imam, Faculty of Agriculture (Protocol No: 2013/01). Pistachio hulls obtained from six different varieties (Beyaz Ben, Kırmızı, Ohadi, Sultani, Uzun and Keten Gömleği) were shade dried and taken to laboratory and milled in a hammer mill through a 1 mm sieve for subsequent analysis.

Dry matter (DM) contents of pistachio hulls was determined by drying the samples at 105°C overnight and ash by igniting the samples in muffle furnace at 525°C for 8 h. Nitrogen (N) contents of pistachio hulls was measured by the Kjeldahl method^[11]. Crude protein contents of pistachio hulls were calculated as N X 6.25. Neutral detergent fiber (NDF) contents of pistachio hulls was determined by the method van Soest and Wine^[12] and ADF contents of pistachio hulls were determined by the method of van Soest^[13]. Condensed tannin contents of pistachio hulls were determined by butanol-HCl method as described by Makkar *et al.*^[14]. All chemical analysis is carried out in duplicate.

Pistachio hulls milled through a 1 mm sieve were incubated *in vitro* rumen fluid in calibrated glass syringes following the procedures of Menke *et al.*^[15].

Rumen fluid was obtained from one year old and approximately 50 kg of three fistulated Awassi ram after one week adjustment period of a diet. Fistulated Awassi ram fed twice daily with a diet containing alfalfa hay (60%) and concentrate (40%). Rumen fluid was collected before morning feeding and squeezed through four layers of cheesecloth. The rumen fluid was flushed with CO₂. The rumen fluid was added to buffered mineral solution in the ratio of 1:2 respectively. Approximately 0.200 gram dry weight of Pistachio hulls samples was weighed in triplicate into calibrated glass syringes of 100 ml. The syringes were prewarmed at 39°C before the injection of 30 ml rumen fluid-buffer mixture into each syringe followed by incubation in a water bath at 39°C. Gas production was recorded at 3, 6, 12, 24, 48, 72 and 96 h after incubation and corrected for blank incubation. Cumulative gas production data of Pistachio hulls were fitted to non-linear exponential model as: $Y = A(1 - \exp^{-ct})$ ^[16].

Where Y is gas production at time 't', A is the potential gas production (ml/200 mg DM), c is the gas production rate constant (h⁻¹) and t is the incubation time (h).

ME (MJ/kg DM) values of pistachio hulls were estimated using equation of Menke *et al.*^[15] as follows:

$$ME \text{ (MJ/kg DM)} = 2.20 + 0.136 \text{ GP} + 0.057 \text{ CP},$$

where, GP = 24 h net gas production (ml/0.200 g); CP = Crude protein (%).

Organic matter digestibility (%) values of pistachio hulls were calculated using equation of Menke *et al.*^[14] as follows:

$$OMD \text{ (\%)} = 14.88 + 0.889\text{GP} + 0.45\text{CP} + 0.0651 \text{ XA}$$

where XA: ash content (%).

One-way analysis of variance (ANOVA) was carried out to determine the effect of variety on the chemical composition, gas production kinetics, ME and OMD of pistachio hulls. Significance between individual means was identified using the Tukey's multiple range tests^[17]. Mean differences were considered significant at $P < 0.05$.

RESULTS

The effect of variety on the chemical composition of pistachio hulls is presented in Table 1. The variety has significant effect on the chemical composition of pistachio hulls. The DM contents of pistachio hulls ranged from 26.45 to 29.25%. The CP contents of pistachio hulls ranged 7.27 to 14.99%. The hulls from Kırmızı, Ohadi and KG had significantly higher CP contents than the others. The NDF and ADF contents ranged from 18.25 to 22.49% and 14.32

to 18.29% respectively. The hulls from Ohadi and Uzun had significantly higher NDF and ADF contents than the others. The ash contents of pistachio hulls were ranged 8.50 to 19.86%. The hulls from Uzun had significantly higher ash content than that of Sultani. The CT contents of hulls ranged from 2.03 to 2.63%. The hulls from BB had significantly higher CT content than that of KG.

The effect of variety on gas production of pistachio hulls at different time intervals is presented in Fig. 1. At 3, 6 and 12 hours incubation the gas production for Kırmızı and Uzun were significantly lower than the others. At 24, 48 and 72 h incubation times, the gas production for Kırmızı, Uzun and KG were significantly higher than the others. At 72 h incubation times the gas productions for BB and Ohadi were significantly higher than the others.

The effect of variety on gas production kinetics, ME, OMD of pistachio hulls is presented in Table 2. The variety had a significant effect on gas production kinetics, ME, OMD of pistachio hulls. The gas production rate ranged from 0.071 to 0.107%. The gas production rate of pistachio hulls from Sultani and KG were significantly higher than the others. The potential gas production of pistachio hulls ranged from 65.92 to 73.46 ml. The potential gas production of pistachio hulls from BB and Ohadi were significantly higher than the others.

The ME and OMD contents of pistachio hulls ranged from 9.76 to 11.05 MJ/kg DM and 69.00 to 74.50% respectively. The ME contents of pistachio hulls from BB and Ohadi were significantly higher than those of Kırmızı, Uzun and KG whereas the OMD contents of pistachio

Table 1. The effect of variety on the chemical composition of pistachio hulls (on a dry matter basis)

Tablo 1. Çeşidin Antep fıstığı dış kabuğunun kimyasal kompozisyonuna etkisi (kuru madde bazında)

Nutrient Composition	Variety						SEM	Sig.
	BB	Kırmızı	Ohadi	Sultani	Uzun	KG		
DM	26.45 ^c	26.73 ^c	29.25 ^a	28.54 ^{ab}	28.98 ^{ab}	26.85 ^b	0.549	**
Ash	16.66 ^{ab}	18.62 ^{ab}	11.38 ^{ab}	8.50 ^b	19.86 ^a	15.44 ^{ab}	3.070	*
CP	11.06 ^c	14.99 ^a	13.30 ^b	7.27 ^d	10.77 ^c	14.49 ^b	0.357	***
NDF	18.25 ^b	19.03 ^b	22.13 ^a	18.51 ^b	22.49 ^a	18.84 ^b	0.225	***
ADF	15.15 ^b	14.32 ^b	18.29 ^a	15.38 ^b	18.02 ^a	14.47 ^b	0.479	***
CT	2.63 ^a	2.34 ^{ab}	2.37 ^{ab}	2.07 ^{ab}	2.55 ^{ab}	2.03 ^b	0.183	**

^{a,b,c} Row means with common superscripts do not differ ($P < 0.05$), SEM - standard error mean, Sig. - significance level, DM - Dry matter %, CP - Crude protein, NDF - Neutral detergent fiber, ADF - Acid detergent fiber, CT - Condensed tannin, BB - Beyaz Ben, KG - Keten Gömleği, * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$

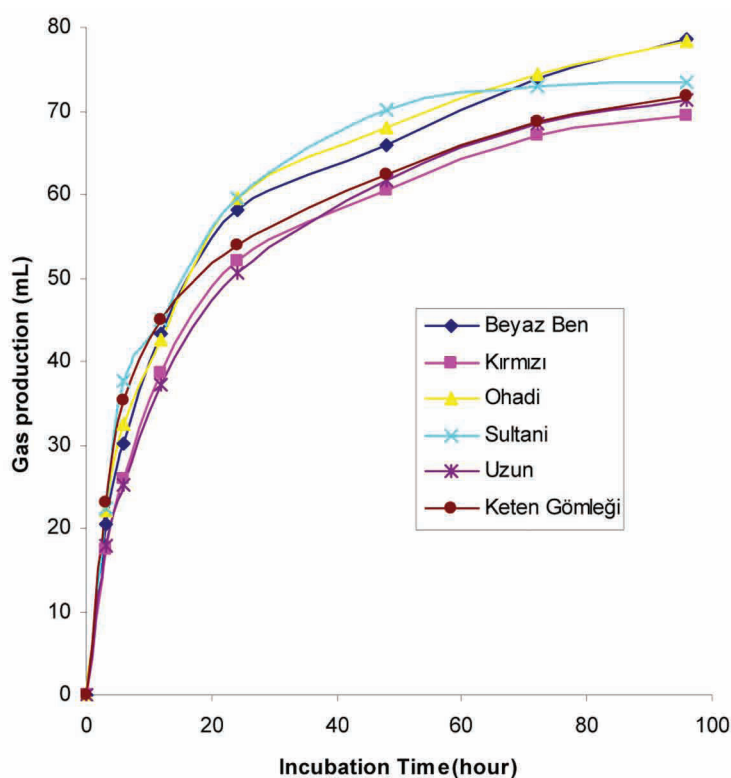


Fig 1. The effect of variety on gas production of pistachio hulls

Şekil 1. Çeşidin Antep fıstığı dış kabuğunun gaz üretimine etkisi

Table 2. The effect of variety on the gas production kinetics, metabolisable energy and organic matter digestibility of pistachio hulls**Tablo 2.** Çeşidin Antep fıstığı dış kabuğunun gaz üretim par/ametrelere, metobolik enerji ve sindirim derecesine etkisi

Parameters	Variety						SEM	Sig.
	BB	Kırmızı	Ohadi	Sultani	Uzun	KG		
c	0.077 ^b	0.074 ^b	0.081 ^b	0.098 ^a	0.071 ^b	0.107 ^a	0.005	***
A	73.11 ^a	66.01 ^c	73.46 ^a	70.96 ^b	67.39 ^c	65.92 ^c	0.538	***
ME	10.74 ^a	10.11 ^{cd}	11.05 ^a	10.70 ^{ab}	9.76 ^d	10.34 ^{bc}	0.108	***
OMD	72.65 ^{ab}	69.00 ^c	74.50 ^a	71.60 ^b	71.60 ^b	70.26 ^{bc}	0.711	***

^{a,b,c} Row means with common superscripts do not differ ($P < 0.05$), **SEM** - standard error mean, **Sig.** - significance level, **c** - gas production rate (%), **A** - potential gas production (mL), **ME** - Metabolisable energy (MJ/kg DM), **OMD** - Organic matter digestibility %, *** $P < 0.001$

hulls from Ohadi were significantly higher than those of pistachio hulls for Kırmızı, Sultani, Uzun and KG.

DISCUSSION

There are significant variations among varieties in terms of chemical compositions of pistachio hulls. The NDF, ADF and CP contents obtained in the current study were considerably lower than those reported by Gholizadeh *et al.*^[6] who reported that NDF and ADF, CP contents were 25, 20 and 16.6% respectively. On the other hand, except for Uzun and Sultani, ash contents of pistachio hulls obtained in the current study were considerably higher than those reported by Gholizadeh *et al.*^[6] who reported that ash content was 12.7%. The differences between two studies are possibly associated differences in variety, growing conditions, kernel maturity and de-hulling process applied.

Except for Sultani, the crude protein contents of pistachio hulls are comparable with those proposed as the minimum requirements for lactation (12% of DM) and growth (11.3% of DM) in ruminants^[18]. Therefore pistachio hulls investigated in the current experiment have the potential for ruminant animals to meet the protein requirements during the critical periods when there is a shortage of high quality forages. However, Kumar and Singh^[19] suggested that CP in diet may be restricted due to high levels of condensed tannin (50 g/kg DM) owing to excessive formation of tannin-protein complexes, *e.g.* with endogenous protein, which pass through the animal largely undigested. On the other hand Barry^[20] suggested that CT which is lower than 2-3% may have beneficial effects to ruminants due to reduction in protein degradation as a result of the formation of protein-tannin complexes in the rumen. However, in the current experiment, the condensed tannin levels of pistachio hulls were lower than those considered detrimental to ruminant animals.

There are significant variation among pistachio varieties in terms of in vitro gas production and estimated parameters such as OMD and ME contents of hulls. The variation in gas production and estimated parameters among pistachio

varieties can be attributed to compositional differences of pistachio hulls, especially cell wall, CP contents and some other anti-nutritive factors such as CT. Babayemi *et al.*^[21] indicated that there are many factors that may determine the amount of gas to be produced during the fermentation, depending on the nature and level of cell content, the presence of secondary metabolites in feedstuffs. Blummel and Orskov^[22] suggested that in vitro gas production is associated with volatile fatty acid (VFA) production following fermentation of substrate so the more fermentation of a substrate the greater the gas production, although the fermentation end products do correlate more closely with gas production.

Organic matter digestibility of pistachio hulls obtained in the current study were considerably higher than those reported by Noghabi and Rauzbehan^[23] who reported that OMD of pistachio hulls ranged from 52.6 to 65.6% and polyethylene glycol (PEG) supplementation significantly increased in vitro OMD of pistachio hulls.

Although there is considerable amount pistachio hull production in Turkey, there is no information about the biomass production of pistachio hulls as a feed source for ruminant animals. It would be more useful if further studies should be focused on the biomass production potential of pistachio hulls in Turkey.

In conclusion, chemical characterization with CP, OMD and ME suggests that the pistachio hulls moderately high CP concentration and was quite digestible therefore pistachio hulls provide feedstuffs of an acceptable quality for ruminant animals.

REFERENCES

1. Valizadeh R, Norouzi MA, Salemi M, Ghiasi E, Yari M: Effects of feeding pistachio by-products on hematology and performance of Balouchi lambs. *J Anim Vet Adv*, 9 (7): 1115-1119, 2010.
2. TÜİK: Türkiye Antep Fıstığı Üretimi İstatistikleri. Türkiye İstatistik Kurumu, 2002-2010.
3. Chahed T, Dhifi W, Hamrouni I, Msaada K, Bellila A, Kchouk ME, Marzouk B: Comparison of pistachio hull essential oils from different Tunisian localities. *Ital J Biochem*, 56 (1): 35-39, 2007.
4. Mahdavi A, Zaghari M, Zahedifar M, Nikkhan A, Aghashani AR: Determination of nutritive value and effects of different levels

replacement of ensiled pistachio epicarp with corn silage on fattening parameters and lamb performance. *Anim Sci J*, 22 (1): 45-54, 2009.

5. Vahmani P, Naserian AA: Pistachio hulls as a feed ingredients for lactating dairy goats. In Proceedings of the British Society of Animal Science. p.145. The Boston Security Analysts Society, UK, 2006.

6. Gholizadeh H, Naserian AA, Valizadeh R, Tahmasebi A: Effects of feeding pistachio hull and interaruminal infusion of urea on feed intake, ruminal and abomasum N-NH₃ and blood metabolites in Iranian Balochi sheep. In Proceedings of British Society of Animal Science. p.163. The Boston Security Analysts Society, UK, 2009.

7. Kamalak A, Canbolat O, Gurbuz Y, Erol A, Ozay O: Effect of maturity stage on chemical composition, *in vitro* and *in situ* dry matter degradation of tumbleweed hay (*Gundelia tournetortii* L.). *Small Ruminant Res*, 58 (2): 149-156, 2005.

8. Kamalak A, Canbolat O, Gurbuz Y, Ozkan CO, Kizilsimsek M: Determination of nutritive value of wild mustard, *Sinapsis arvensis* harvested at different maturity stages using *in situ* and *in vitro* measurements. *Asian Austral J Anim Sci*, 18 (9): 1249-1254, 2005.

9. Kamalak A: Determination of potential nutritive value of *Polygonum aviculare* hay harvested at three maturity stages. *J Appl Anim Res*, 38 (1): 69-71, 2010.

10. Kamalak A, Canbolat O: Determination of nutritive value of wild narrow-leaved clover (*Trifolium angustifolium*) hay harvested at three maturity stages using chemical composition and *in vitro* gas production. *Trop Grassland*, 44 (2): 128-133, 2010.

11. AOAC: Official Method of Analysis. 15th ed., pp.66-88, Association of Official Analytical Chemists, Washington, DC, USA, 1990.

12. Van Soest PJ, Wine RH: The use of detergents in the analysis of fibrous feeds. IV. Determination of plant cell wall constituents. *J Assoc Off Anal Chem*, 50, 50-55, 1967.

13. Van Soest PJ: The use of detergents in the analysis of fibrous feeds. II. A rapid method for the determination of fiber and lignin. *J Assoc Off Anal*

Chem, 46, 829-835, 1963.

14. Makkar HPS, Blummel M, Becker K: Formation of complexes between polyvinyl pyrrolidones or polyethylene glycols and their implication in gas production and true digestibility *in vitro* techniques. *Brit J Nutr*, 73 (6): 897-913, 1995.

15. Menke KH, Raab L, Salewski A, Steingass H, Fritz D, Schneider W: The estimation of the digestibility and metabolizable energy content of ruminant feedingstuffs from the gas production when they are incubated with rumen liquor *in vitro*. *J Agric Sci (Camb)*, 93 (1): 217-222, 1979.

16. Orskov ER, McDonald I: The estimation of protein degradability in the rumen from incubation measurements weighted according to rate of passage. *J Agric Sci Camb*, 92 (2): 499-503, 1979.

17. Pearse ES, Hartley HO: Biometrika Tables for Statisticians. Cambridge University Press, 1966.

18. ARC: The nutrient requirements of ruminant livestock. Technical review by an agricultural research council working party. Commonwealth Agricultural Research Bureau, Farnham Royal, UK. 1984.

19. Kumar R, Singh M: Tannins: their adverse role in ruminant nutrition. *J Agr Food Chem*, 32 (3): 447-453, 1984.

20. Barry TN: Secondary compounds of forages. In, Hacker JB, Ternouth JH (Eds): Nutrition of Herbivores. 1st ed., pp.91-120, Academic Press, Sydney, 1987.

21. Babayemi OJ, Demeyer D, Fievez V: Nutritive value and qualitative assessment of secondary compounds in seeds of eight tropical browse, shrub and pulse legumes. *Commun Agric Appl Biol Sci*, 69 (1): 103-110, 2004.

22. Blummel M, Orskov ER: Comparison of an *in vitro* gas production and nylon bag degradability of roughages in predicting feed intake in cattle. *Anim Feed Sci Tech*, 40 (2-3): 109-119, 1993.

23. Noghabi RL, Rouzbehan Y: The *in vitro* organic matter digestibility of pistachio hull using rumen fluid in Taleshi sheep. *Iran J Anim Sci*, 42 (3): 231-237, 2011.

Geleneksel Yöntemle Üretilen Sucuklarda *Listeria monocytogenes*, *Staphylococcus aureus* ve Koliform Varlığının Araştırılması

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Özet

Uygun olmayan koşullarda üretilen sucuklarda gıda kaynaklı patojenlerden *Listeria monocytogenes*, koagülaz (+) *Staphylococcus aureus* ve koliformlar gelişebilmekte ve bu ürünler insan sağlığı açısından risk arz edebilmektedir. Bu çalışmada, Mersin ve Adana illerine bağlı yaylalarda geleneksel yöntemlerle üretilen (kırsalda satışı ve servisi yapılan) 60 sucuk örneği *L. monocytogenes*, koagülaz (+) *S. aureus* ve koliformlar açısından incelenmiştir. Türk Gıda Kodeksine göre sucukta hiç bulunmaması gereken *L. monocytogenes* 7 örnekte tespit edilmiştir. Ayrıca 10 örnekte koagülaz (+) *S. aureus* ve 18 örnekte koliform grubu mikroorganizmaya rastlanmıştır. Kırsal alanda sucuk satış ve servis noktalarındaki denetimlerin artırılması ve üretim yapan kişilerin hijyen konusunda eğitimi sorunun çözümüne katkı sağlayacaktır.

Anahtar sözcükler: Et ürünleri, Sucuk, Mikrobiyel kalite, Gıda kaynaklı patojenler

An Investigation on the Microbiological Quality of Sucuk Produced by Traditional Methods

Summary

Foodborne pathogens like *Listeria monocytogenes*, coagulase(+) *Staphylococcus aureus* and coliforms may grow in sucuk produced under unsuitable conditions and possess risks for human health. In this study, 60 sucuk samples produced by traditional methods in plateaus of Mersin and Adana provinces were investigated in terms of *L. monocytogenes*, coagulase (+) *S. aureus* and coliforms. *L. monocytogenes*, which must not to be present in sausages according to Turkish Food Codex, was detected in 7 samples. In addition, 10 of samples were found to be contaminated with coagulase (+) *S. aureus* and 18 of samples with coliforms. Increasing controls of sausage sale and service points in rural areas, and training of producers on hygiene will contribute to the solution of the problem.

Keywords: Meat products, Turkish dry fermented sausage (sucuk), Microbial quality, Foodborne pathogens

GİRİŞ

Sucuğun bileşimi (yağ, baharat ve diğer katkı maddelerinin miktar ve çeşitleri) pazar isteklerine göre değişmektedir. Geleneksel sucuk üretimi iklim koşullarına bağlı olarak özellikle sıcaklığın, hava akımının ve rutubetin uygun olduğu sonbaharda yapılır. Sucuk hamuru kılıflara doldurulduktan sonra askılara alınarak olgunlaşmaya bırakılır. Bu yöntemde sucuklar 20-30 günde olgunlaşır ^[1]. Fermente sucuklarda kalite niteliklerinin geliştirilip artırılmasında bazı mikroorganizmaların enzimatik ve teknolojik etkileri büyük önem taşır. Bu mikroorganizmaların başlıcaları laktobasil, mikrokok ve stafilokok grubunda yer alan bakteri türleridir. Bunlardan laktobasiller doğrudan oluşturdukları laktik asit ile fermente sucuklarda

pH'nın düşmesi, aroma oluşumu ve dolaylı olarak renk oluşumu; mikrokok/stafilokoklar ise nitrati indirgeyerek renk ve aroma oluşumunda rol oynarlar ^[2]. Geleneksel yöntemlerle yapılan fermente Türk sucuğu, olgunlaşmada önemli rol oynayan faydalı bakterilerin yanı sıra istenmeyen bazı mikroorganizmaları da barındırabilir. Bu çalışmada incelenen bakterilerden biri olan *S. aureus*'un, dünyada bakteriyel gıda zehirlenmelerinden sorumlu tutulan bakteriyel ajanlar arasında ilk sırada yer aldığı belirtilmektedir ^[3].

Koliform grubu bakteriler 37°C'de 48 saat içinde laktozdan asit ve gaz oluşturabilen bakterilerdir. Koliform



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grubu bakteriler gıdalarda hijyen indikatörü olarak değerlendirilir. *Listeria* cinsinin 8 türü vardır. Bu türler *Listeria monocytogenes*, *L. innocua*, *L. seeligeri*, *L. welshimeri*, *L. ivanovii*, *L. marthii*, *L. rocourtiae* ve *L. grayi*'dir. *Listeria* türleri içerisinde insan listeriozisi ile genel olarak *Listeria monocytogenes* seyrek olarak da *L. seeligeri* ve *L. ivanovii* ilişkilendirilmektedir ^[4]. *Listeria* ubiquiter bir bakteri olduğundan insanlara geçişinde gıdalar başta olmak üzere birçok muhtemel yol vardır. İnsan listeriozunun asıl kaynağını *L. monocytogenes* ile kontamine hayvansal gıdalar oluşturmaktadır ^[3]. Sucuk hamurunun başlangıç pH'sının yüksek olması *L. monocytogenes* gelişmesi için uygun ortam sağlamaktadır. Olgunlaşmanın ileri günlerinde değişik faktörlerin (örn. bakteriyosin) kombine etkisi ile sayı azalmaktadır.

Gıdalarda *L. monocytogenes* belirlenmesinde sıfır tolerans politikası geçerlidir. Bu yüzden *L. monocytogenes* analizlerinde 25 gr'da var/yok testi uygulanmaktadır. Türk Gıda Kodeksi Mikrobiyolojik Kriterler Yönetmeliğine göre ^[5] fermente sucukta *L. monocytogenes* bulunmamalıdır. Aynı yönetmenlikte *S. aureus* ve koliform için herhangi bir kriter mevcut değildir. Türk Standartları Enstitüsü'nün 2002 yılında yayımladığı TS 1070 Türk sucuğu standardına göre; incelenen her beş örnekte *S. aureus* için bir örnekte 5×10^3 kob/g dört örnekte 10^2 kob/g'ı ve koliform grubu bakterilerin 10 kob/g'ı geçmemesi gerektiği öngörülmektedir. Kök ve ark.^[6] Aydın ilinde satışa sunulan fermente sucukların %14'ünde *Listeria* spp. ve %4'ünde *L. monocytogenes* tespit etmişlerdir. Ayrıca değişik oranlarda koagülaz (+) *S. aureus* ve koliform grubu bakteri belirlediklerini bildirmişlerdir. Öksüztepe ve ark. Elazığ'da yaptıkları çalışmada sucuk örneklerinin %9'unda *Listeria* spp. ve %10'unda koagülaz (+) *S. aureus* tespit etmişlerdir ^[7]. Koluman ve ark.^[8] inceledikleri 50 kıyma örneğinde %70 oranında *S. aureus*, Güven ve ark.^[9] ise 80 kıyma örneğinin %46.25'inde bu bakteriyi tespit ettiklerini bildirmişlerdir.

Son yıllarda fast food kültürünün ve işlenmiş et ürünlerindeki rekabetin artması ile geleneksel Türk sucuğu daha çok şehirden uzak noktalarda ve kırsalda yemek hizmeti veren işletmelerde bulunur hale gelmiştir. Ülkemizde halkın damak tadına hitabeden ve sevilerek tüketilen bir ürün olan fermente sucukta halk sağlığı açısından muhtemel risklerinin belirlenmesi, bilimsel ve yasal bir gerekliliktir. Ham maddesinin et olması dolayısıyla sucuğun, hammadde ve üretim aşamasından tüketiciye ulaşıncaya kadar geçen her dönemde oluşabilecek mikrobiyolojik riskler halk sağlığı açısından önem taşımaktadır. Bu çalışma, kırsal piyasadan temin edilen fermente sucukların *L. monocytogenes*,

koagülaz (+) *S. aureus* ve koliformlar açısından mikrobiyolojik kalitesini ortaya koymak amacıyla yapılmıştır.

MATERYAL ve METOT

Çalışmamızda 60 adet fermente sucuk kullanıldı. Örnekler Adana, Tarsus, Mersin yaylalarından Eylül-Ekim-Kasım 2011 zaman diliminde toplandı. Aseptik şartlarda steril numune poşetlerine alınarak soğuk zincirle laboratuara getirilen örnekler hemen analize alındı.

L. monocytogenes aranması ^[10] ve *S. aureus* sayımı için ^[11] FDA/BAM yöntemleri kullanıldı. Koliform grubu bakterilerin sayımı için Halkman'ın belirttiği metot kullanıldı ^[12].

BULGULAR

Çalışmamızda kullanılan 60 adet fermente sucuk örneğinin 7'sinde *L. monocytogenes*, 10'unda koagülaz (+) *S. aureus* ve 18'inde koliform grubu mikroorganizma tespit edilmiştir. Katı vasatta tipik *L. monocytogenes* kolonisi tespit edilen örneklerdeki sonuçlar, aynı örneklerin vidas sonuçları ile uyumlu bulundu. Fermente sucukların mikrobiyolojik analizleri sonucu elde edilen pozitif örnekler Tablo 1'de, fermente sucuk örneklerinde mikroorganizma sayılarının yüzde dağılımı ise Tablo 2'de verilmiştir.

TARTIŞMA ve SONUÇ

Özellikle et ve et ürünleri ile insanlara bulaşan *Listeria*'lar insanlarda ciddi infeksiyonlara yol açabilmektedir. Bu çalışmada analize alınan örneklerin %11.6'sında *L. monocytogenes* tespit edilmiştir. Sancak ve ark. ^[13], fermente Türk sucuklarında yaptıkları çalışmada, örneklerin %15'inde *L. monocytogenes* tespit ettiklerini bildirmişlerdir. Aynı çalışmada salam örneklerinin %10'unun, sosis örneklerinin ise %25'inin *L. monocytogenes* ile konta-

Tablo 1. Fermente sucuk örneklerinin mikrobiyolojik sonuçları (n=60)

Table 1. The results of microbiological of samples fermented sausage (n=60)

Mikroorganizma	(+) Örnek Sayısı	%	Toplam Örnek Sayısı
<i>L. monocytogenes</i>	7	11.6	60
Koagülaz(+) <i>Staphylococcus aureus</i>	10	16.6	60
Koliform	18	30.0	60

Tablo 2. Fermente sucuk örneklerindeki mikroorganizma sayılarının yüzdesi (kob/g, n=60)

Table 2. The fermented sausages samples of microorganism count percent (cfu/g, n=60)

Mikroorganizma	<0.1x10		0.1x10 ¹ - 9.9x10 ¹		1.0x10 ² - 9.9x10 ²		1.0x10 ³ - 9.9x10 ³		>1.0x10 ⁴	
	n	%	n	%	n	%	n	%	n	%
Koagülaz(+) <i>Staphylococcus aureus</i>	50	83.4	1	1.66	5	8	4	6.4	-	-
Koliform	42	70	1	1.66	9	15	8	13.33	-	-

mine olduğu bildirilmiştir. Çalışmamızda tespit ettiğimiz %11.6 *L. monocytogenes* oranı Sancak ve ark.^[13] tarafından tespit edilen %15'lik ve Bozkurt^[14] tarafından 25 sucuk örneğinde bildirilen %31.58'lik oranlardan düşüktür. Kök ve ark.^[6] Aydın'da, Öksüztepe ve ark.^[7] Elazığ'da yaptıkları çalışmalarda fermente sucukların %4'ünde *L. monocytogenes* bulunduğunu bildirmişlerdir. Araştırmacıların tespit ettikleri oran, bu çalışmada belirlenen orandan oldukça düşüktür. Bu farklılıklar örneklerde kullanılan sorbat, nitrat ve starter gibi katkı maddelerinin varlığından ve farklı oranlarda bulunmasının yanı sıra ambalaj farklılıklarından da kaynaklanabilir. Sucuk üretiminde kullanılan tuz, sodyum nitrit gibi maddelerin antilisterial etkilerinin yanı sıra, olgunlaşma sırasında floraya hâkim olan laktobasillerin pH'yı düşürmesi ve yine bu bakterilerin ürettiği bakteriosinlerin antagonistik etkisinden dolayı sucukta *L. monocytogenes* sayısı azalabilir. Çolak ve ark.^[15] İstanbul piyasasından elde ettikleri 300 sucuk örneğinin 35'inde (%11.6) *L. monocytogenes* tespit etmişlerdir. Bu veriler çalışmamızda elde ettiğimiz verilerle oransal bakımdan tam uyum içerisindedir.

pH 4.2'nin üzerindeki fermente et ürünlerinde bulunma ihtimali yüksek olan *S. aureus* incelediğimiz örneklerden 10'unda (%16.6) tespit edilmiştir. Kök ve ark.^[6] inceledikleri örneklerin %12'sinde, Öksüztepe ve ark.^[7] %10'unda Sancak ve ark.^[16] %16'sında (koagülaz (+) *S. aureus* tespit ettiklerinin bildirmişlerdir. Bulgularımız Sancak ve ark.^[16] bildirdiği bulgular ile uyum içerisindedir. Erdoğan ve ark.^[17] fermente sucuklarda bildirdiği %6.6 *S. aureus* oranından ise oldukça fazladır. Tespit oranlarının farklı olmasında incelenen örnek sayısı değişkenliğinin etkili olduğu düşünülmektedir. Osmotolerant bir bakteri olan *S. aureus* ile sucuk yapımı sırasında az sayıda kontaminasyon olsa dahi fermentasyon sürecindeki düşük a_w düzeylerinde gelişip toksik etki gösterebilir. Uygun koşullarda olgunlaştırılmayan ve starter kültür içermeyen sucuklarda stafilkokların gelişerek enterotoksin oluşturabilecekleri belirtilmiştir^[3]. Stafilkokal gıda zehirlenmelerinin önüne geçmek için üretim, personel ve alet ekipman hijyeni önem arz etmektedir. Kıralsda üretilen sucuklarda koruyucu ve starter kültür kullanılmadığı düşünülmektedir.

Bu çalışmada kullanılan örneklerde hijyen indikatörü olarak bilinen koliform bakterilerine %30 oranında rastlanmıştır (Tablo 1). Bu durum ham maddenin kontamine olmasından, üretim hijyenine dikkat edilmemesinden, alet ekipmanın kontamine olmasından ve yetersiz olgunlaştırmadan kaynaklanabilir. Koliform grubu bakteriler ürünün raf ömrünü azaltan önemli bir faktördür. Kök ve ark.^[6] sucuk örneklerinde ortalama koliform bakteri sayısı 1.62 log kob/g olarak bümüştür. Erdoğan ve ark.^[17] inceledikleri sucuk örneklerinde ortalama koliform bakteri sayısını 244/g, olarak bildirmişlerdir. Bu çalışmada koliform sayısı üreme olan örneklerde 9.0×10^1 - 6.0×10^3 kob/g arasında bulunmuştur. Çalışmalar arasındaki farklılıklar üretim yerindeki hijyen, ham maddenin bakteriyel yükü, personel

ve alet-ekipman hijyeninden kaynaklanabilir. Özellikle koagülaz (+) *S. aureus* ve koliform sayısında tespit edilen miktarlar bu parametrelerin sucukla ilgili mikrobiyolojik kriterler yönetmeliğine girmesi gerektiğini ortaya koymaktadır. Toplam 42 örnekte koliform grubu bakteriye rastlanmadı. Bunun sebebi olgunlaşma sırasında su aktivitesinin azalması, pH'nın düşmesi, rekabetçi flora ve bakteriyosinlerin varlığı olabilir.

Sonuç olarak elde edilen veriler sucuk örneklerinde *L. monocytogenes*, koagülaz (+) *S. aureus* ve koliform grubu bakterilerin varlığını ortaya koymuştur. Bu durum ürünün raf ömrünün kılmasına neden olmasının yanında halk sağlığı açısından da büyük önem arz etmektedir. Sucuğun uygun olmayan koşullarda üretilmesi ve değişik özelliklerdeki ham maddeler kullanılması kaynaklanan mikrobiyel riskleri bertaraf etmek için kırsal alandaki satış ve servis noktalarında denetimlerin artırılması gerekir. Gelecekte sucuk üretimi yapan kişilerin ham madde ve üretim hijyeni yanı sıra muhafaza ve satış koşulları konularında eğitilmesi ürünlerin halk sağlığı riski oluşturma ihtimalini düşürecektir.

KAYNAKLAR

1. Arslan A: Et Muayenesi ve Et Ürünleri Teknolojisi. s.330-344, Medipres Matbaacılık, Malatya, 2002.
2. Dinçer B, Özdemir H, Mutluer B, Yaglı Ö, Erol İ, Akgün S: Türk fermente sucuğuna özgü starter kültür bakterilerinin izolasyon, identifikasyon ve üretimleri. *Ankara Üniv Vet Fak Derg*, 42, 285-293, 1995.
3. Erol İ: Gıda Hijyeni ve Mikrobiyolojisi. s.126-144, Pozitif Matbaacılık, Ankara, 2007.
4. den Bakker HC, Cummings CA, Ferreira V, Vatta P, Orsi RH, Degoricija L, Barker M, Petruskeno O, Furtado MR, Wiedmann M: Comparative genomics of the bacterial genus *Listeria*: Genome evolution is characterized by limited gene acquisition and limited gene loss. *BMC Genomics*, 11, 688-708, 2010.
5. Anonim: Türk Gıda Kodeksi Mikrobiyolojik Kriterler Yönetmeliği. Resmi Gazete, 29.12.2011-28157, 2011.
6. Kök F, Özbey G, Muz A: Aydın ilinde satışa sunulan fermente sucukların mikrobiyolojik kalitelerinin incelenmesi. *Fırat Üniv Sağ Bil Derg*, 21 (6): 249-252, 2007.
7. Öksüztepe G, Güran HŞ, İncili GK, Gül SB: Elazığ'da tüketime sunulan fermente sucukların mikrobiyolojik ve kimyasal kalitesi. *Fırat Üniv Sağ Bil Derg*, 25 (3): 107-114, 2011.
8. Koluman A, Ünlü T, Dikici A, Tezel A, Akçelik EN, Burkan ZT: Presence of *Staphylococcus aureus* and staphylococcal enterotoxins in different foods. *Kafkas Üniv Vet Fak Derg*, 17 (Suppl-A): S55-S60, 2011.
9. Güven A, Gülmez M, Kamber U: Kars ilinde tüketime sunulan kıymalarda bazı patojen mikroorganizmaların araştırılması ve kıymaların mikrobiyolojik kalitesinin belirlenmesi. *Kafkas Üniv Vet Fak Derg*, 3 (1): 57-65, 1997.
10. FDA/BAM: Bacteriological Analytical Manual Chapter 10 (January 2003) Detection and Enumeration of *Listeria monocytogenes* in Foods, 2003.
11. FDA/BAM: *Staphylococcus aureus*. January Bacteriological Analytical Manual, Chapter 12 *Staphylococcus aureus*. 2001.
12. Halkman AK: Gıda Mikrobiyolojisi Uygulamaları. s. 193, 141, 262, Başak Matbaacılık, Ankara, 2005.
13. Sancak YC, İşleyici Ö, SAĞUN E: Van'da tüketime sunulan bazı et ürünlerinde *Listeria monocytogenes* varlığı. *YYU Vet Fak Derg*, 18 (1): 93-99, 2007.

14. Bozkurt ENN: Et ve et ürünlerinden *Listeria monocytogenes*'in izolasyonu. *Doktora Tezi* Yüzüncü Yıl Üniv. Sağlık Bil. Enst., 2003.

15. Çolak H, Hampikyan H, Ulusoy B, Bingo EB: Presence of *Listeria monocytogenes* in Turkish style fermented sausage (sucuk). *Food Control*, 18, 30-32, 2007.

16. Sancak YC, Kayaardı S, Sagun E, İşleyici Ö, Sancak H: Van'da

tüketime sunulan fermente türk sucuklarının fiziksel, kimyasal, mikrobiyolojik ve organoleptik niteliklerinin incelenmesi, *YYU Vet Fak Derg*, 7, 67-73, 1996.

17. Erdoğan Ö, Ergün Ö: Kahramanmaraş piyasasında tüketilen sucukların bazı fiziksel, kimyasal, duyuşal ve mikrobiyolojik özellikleri. *İstanbul Üniv Vet Fak Derg*, 31 (1): 55-65, 2005.

Comparison of Three Methods for Routine Detection of *Staphylococcus aureus* Isolated from Bovine Mastitis

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Summary

The present study aimed to compare three identification methods that are routinely used for the detection of *Staphylococcus aureus* as bovine mastitis agent. The evaluated methods were as followed: conventional biochemical method, commercial identification system BioLog (Gen III MicroPlate) and amplification of species-specific gene (*nuc*) by polymerase chain reaction (PCR). A total of 73 staphylococcal isolates were collected from 453 individual milk samples from dairy cows with subclinical and clinical mastitis from different farms in Bulgaria. This isolates were determined as 60 coagulase-positive, 3 catalase-negative and 10 coagulase-negative by conventional methods. BioLog system identified 72 isolates as *S. aureus* subsp. *aureus* and one coagulase-positive isolate as *S. schleiferi* subsp. *coagulans*. PCR amplification of *nuc* gene further confirmed *S. aureus* subsp. *aureus* isolates identified by the BioLog system. The primary identification of *S. aureus* on the basis of coagulase level led to erroneous determination of 14 (19.2%) of the isolates. Based on the findings, BioLog system and PCR appear to be more reliable detection systems for *S. aureus* from milk. In conclusion, the present study showed that a routine approach using a combination of phenotypic and molecular detection systems could improve *S. aureus* detection in milk.

Keywords: *Staphylococcus aureus*, Bovine mastitis, Identification, BioLog, *nuc* gene, PCR

Bovin Mastitislerde *Staphylococcus aureus*'un Rutin Tespitinde Üç Metodun Karşılaştırılması

Özet

Bu çalışma Bovin mastitis etkeni olarak *Staphylococcus aureus*'un belirlenmesinde rutin olarak kullanılan üç tespit yöntemini karşılaştırmayı amaçlamaktadır. Denenen yöntemler şunlardır; konvensiyonel biyokimyasal metot, ticari tespit sistem BioLog (Gen III MicroPlate) ve polimeraz zincir reaksiyonu (PZR) ile amplifiye edilmiş tür spesifik gen (*nuc*). Bulgaristan'daki farklı çiftliklerde subklinik ve klinik mastitisli sütçü ineklerden alınan 453 adet örnekten toplam 73 staphylococcal izolat elde edildi. Konvensiyonel metotlarla bu izolatların 60'ı koagülaz pozitif, 3'ü katalaz negatif ve 10'u koagülaz negatif olarak belirlendi. BioLog sistemi ile 72 izolat *S. aureus* subsp. *aureus* olarak tespit edilirken bir adet koagülaz pozitif izolat *S. schleiferi* subsp. *coagulans* olarak belirlendi. PZR ile *nuc* geninin amplifikasyonu kullanılarak BioLog sistemi ile *S. aureus* subsp. *aureus* olarak tespit edilen izolatlar teyit edildi. Koagülaz seviyesine dayanarak *S. aureus*'un primer identifikasyonu 14 (%19,2) hatalı tespite neden oldu. Bu bulgulara dayanarak BioLog sistem ve PZR, *S. aureus*'un süttten tespitinde daha güvenilir yöntemler olarak belirlendi. Sonuç olarak, bu çalışma ile rutin olarak fenotipik ve moleküler tespit yöntemlerinin beraber kullanılmasının süttten *S. aureus*'un tespitinde başarıyı artıracığı ortaya konuldu.

Anahtar sözcükler: *Staphylococcus aureus*, Bovin mastitis, Tespit,, BioLog, *nuc* geni, PZR

INTRODUCTION

Bovine mastitis is among the leading issues that cause serious economic losses in dairy industry throughout the

world, resulting in reduced milk production, reduction of the quality of milk through contamination and due to



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treatment-associated hazards^[1]. Of all mastitis causative agents, *S. aureus* is most commonly isolated^[2], while the coagulase-negative staphylococci (CoNS) are also common but not considered as significant mastitis pathogens^[3]. Therefore, the identification of the staphylococci by their ability to produce coagulase is deemed the main criterion for differentiation of those with pathogenic potential from others. On the other hand, some coagulase-positive staphylococci (CoPS) other than *S. aureus* such as *S. intermedius* and *S. hyicus* could cause intramammary infections in cows^[4]. In that respect, the differentiation of the staphylococcal mastitis causative agents is important not only for the therapeutic approach and the remedial schemes, but also for the ability of some of them to produce a wide range of virulent factors^[5].

Several conventional phenotypic and molecular methods have been used as routine approaches for the identification of staphylococci. The conventional culture methods require isolation of the bacteria on suitable media and subsequent identification with numerous biochemical tests. Commercial identification systems recognizing *S. aureus* are available such as BD Phoenix^[6], BBL Crystal and Api Staph system^[7] and Vitek2^[8]. Over the last few years, molecular techniques have become more popular and are currently used as detecting systems of mastitis pathogens in many diagnostic laboratories^[9].

The present study aimed to compare three methods for routine identification of *S. aureus*, isolated from dairy cows, the first based upon the conventional identification and coagulase activity; the second using an identification system (BioLog) and the third - a molecular approach using PCR amplifying the species-specific gene, *nuc*, as a reference method.

MATERIAL and METHODS

Isolation and Primary Identification of *S. aureus*

A total of 453 individual milk samples were collected from cows with subclinical and clinical mastitis from different farms in Bulgaria. The indication for sampling from the subclinical mastitis animals was the positive result for increased somatic cell count after screening with Mastitis test NK (Biovet, Czech Republic). The milk sampling was done according to the guidelines of bacteriological examination. The samples (10 µL) were inoculated on trypticase soy agar (TSA, Fluka, India) supplemented with 5% defibrinated sheep blood (TSBA) and incubated at 37°C for 24-48 h under aerobic conditions. The primary identification of staphylococcal isolates was made by Gram staining, catalase and oxidase activity, colony morphology and pigmentation, hemolysis, growth and mannitol fermentation on Chapman agar (NCIPD, Bulgaria). Further, the pathogenic potential of the isolates was tested by tube coagulase test with rabbit plasma (NCIPD, Bulgaria).

A weak coagulase activity was recorded as positive reaction.

Identification of the Isolates with BioLog System

The suspicious for *S. aureus* isolates were checked with identification system BioLog Gen III microplates following precisely the manufacturer's instructions (Biolog, Hayward, USA). In brief, protocol A was used for identifying of *S. aureus*. The isolates were cultured on TSBA and the inocula were prepared in a special type of liquid medium, provided by the company until achieving the desired cell density determined by turbidimeter. The plates were filled with 100 mL of the readily prepared inocula and incubated at 33°C for 24 h under aerobic conditions. After incubation, the plates were read by the computer system software OmniLog.

Extraction of DNA and PCR Analysis

Bacterial colonies were suspended in 100 µL bidistilled water and boiled for 10 min to extract DNA. After centrifugation at 12.000 g for 5 min at room temperature, the supernatant that contains the nucleic acids was used as DNA template for subsequent PCR analysis. DNA was also isolated by universal commercially available kit - prepGem (ZyGem, USA), according to the company's protocol. The concentration and purity of DNA extract was determined by DNA/RNA spectrophotometer Gene Quant 1300 at A260 and A280. The DNA extracts were then stored at -20°C until the beginning of the experiments.

The PCR methodology used in the study was described previously by Brakstad et al.¹⁰ with an expected amplicon size of 270 bp. Positive and negative control strains were included for each PCR experiment.

RESULTS

From collected 453 milk samples, 73 staphylococcal strains were isolated. Gram-stained smears of the pure cultures exhibited clusters of Gram-positive cocci. Catalase test revealed three isolates with catalase-negative reaction. The hemolytic activity of isolates was different, variations were observed in the pigmentation of the colonies as well. Ten of the beta-hemolytic isolates gave negative results in coagulase tests in two independent trials. With exception of one coagulase-positive isolate, all others fermented mannitol of Chapman agar under aerobic conditions (Table 1).

BioLog system determined catalase-negative and coagulase-negative isolates as *S. aureus* subsp. *aureus* with probability of 0.919 - 0.999. The typical coagulase-positive isolates were confirmed as *S. aureus* subsp. *aureus*. The coagulase-positive non-hemolytic isolate with white colonies and non-fermenting mannitol was identified as *S. schleiferi* subsp. *coagulans* with probability of 0.970. Three

Table 1. Comparison of the methods for detection of *S. aureus* isolated from bovine mastitis**Tablo 1.** Bovin mastitislerden *S. aureus*'un tespiti amacıyla kullanılan metotların karşılaştırılması

Isolates n/%	Criteria for Primary Identification							BioLog ID	nuc gene
	Catalase /oxidase	Haemolysis			Pigment	Coagulase	Mannitol		
		Double	Beta	No					
3/4.1	-/-	+			creamy-white	+	+	<i>S. aureus</i>	+
10/13.7	+/-		+		greyish	-	+	<i>S. aureus</i>	+
32/43.8	+/-	+			yellow	+	+	<i>S. aureus</i>	+
6/8.2	+/-	+			greyish	+	+	<i>S. aureus</i>	+
14/19.2	+/-		+		greyish	+	+	<i>S. aureus</i>	+
7/9.6	+/-		+		yellow	+	+	<i>S. aureus</i>	+
1/1.4	+/-			-	white	+	-	<i>S. schleiferi coagulans</i>	-
Key: + = positive reaction; - = negative reaction									

Key: + = positive reaction; - = negative reaction

independent trials were performed to confirm the species affiliation of the isolate.

The PCR protocol based on the *nuc* gene further confirmed *S. aureus* subsp. *aureus* isolates identified by BioLog system (Table 1). The two DNA isolation techniques were of similar quality as template for PCR.

DISCUSSION

The primary conventional identification of CoPS is very important and suggests about *S. aureus*, however, prior to testing with rabbit plasma some tests as Gram staining, catalase and oxidase activity that are particularly valuable for the discrimination of the agent from streptococci and micrococci have to be done as routine tests [11]. An interesting and unique finding in this study was the three isolates with catalase-negative reaction which was preserved throughout subsequent subcultivations. The isolates showed double hemolysis, which gave us the clue and motivated us further to continue with the identification process. The review of the literature revealed reports for catalase-negative *S. aureus*, but they referred to isolates of human origin [7,12-14]. Piau et al. [8] determined a catalase-negative *S. aureus* strain with point mutations in the *kata* gene as a possible mechanism for the loss of catalase activity. To the best of our knowledge, this is the first report of catalase-negative *S. aureus* from bovine origin in particular from cases of subclinical mastitis, determined by the primary routine identification.

According to Bannerman [15] the detection of free coagulase in the plasma tube coagulase test is the gold standard for *S. aureus* identification. In this regard, another interesting finding in this study was the isolation of 10 coagulase-negative strains from subclinical mastitis with a wide range of beta hemolysis. Worldwide reports of coagulase-negative variants of *S. aureus* in bovine mastitis are still rare [16]. Akineden et al. [4] determined two *S. aureus*

coagulase-negative strains, that would be interpreted as CoNS naturally. The very weak beta hemolysin activity of the isolates motivated the authors to further identify the strains on a molecular level, that confirmed the species affiliation of the isolates as *S. aureus*.

An important criterion for the presumptive primary identification of *S. aureus* is also the hemolytic activity of isolates in the initial cultures. In our study 41 (56.2%) of isolates showed double hemolysis (alpha + beta hemolysins), 31 (42.5%) beta and 1 (1.4%) was non-hemolytic. The isolates with double hemolysis coagulated rabbit plasma in the test tube by the 4th h, a quick (less than 4 h) coagulation was also observed in some beta hemolytic isolates (8/31) as well as in the non-hemolytic and the others required overnight incubation. In our study we did not find *S. aureus* isolates without hemolytic activity.

The staphylococcal isolates were subjected to further analysis and identification by the BioLog system. The atypical catalase-negative (n=3) and coagulase-negative (n=10) isolates were determined as *S. aureus* subsp. *aureus* with a very high probability (91.9-99.9%). Vitek 2 (bioMérieux, France) Gram-positive identification card identified the catalase-negative isolate as *S. aureus* with probability of 93 % [8]. To et al. [17] compared two identification systems for the identification of a catalase-negative *S. aureus* strain from a patient with endocarditis and pericarditis of the mitral valve. While Becton Dickinson Phoenix PID panel identified the strain as *S. aureus*, Vitek system Gram-positive identification test could not identify the bacterium. Our coagulase-positive isolate without hemolytic activity and mannitol fermenting ability was identified as *S. schleiferi* subsp. *coagulans*. *S. schleiferi* subsp. *coagulans* is considered pathogenic mainly in dogs as causative agent of otitis externa [11]. In our case the bacterium was isolated from clinical mastitis in pure culture. Working with BioLog Gen III does not require any preliminary tests, even Gram staining, which is important in selection of

other identification systems. BioLog system recognizes the biochemical profile of approximately 2500 bacterial species, including many important veterinary pathogens and those responsible for mastitis. In that respect, we do not agree with Ahmadi et al.^[9] who affirmed that most commercial identification systems were not designed to determine the important veterinary pathogens.

S. aureus isolates identified with BioLog were confirmed by PCR amplification of the fragment of agent's species-specific *nuc* gene. The results of our research showed an excellent correlation between the BioLog system and PCR detection of *S. aureus* in bacterial colonies from bovine mastitis. A protocol for the identification of *S. aureus* in milk by PCR was proposed by Ahmadi et al.^[9]. Real-time PCR-based commercial reagent kit is now available for investigation of bovine mastitis pathogens without conventional culturing^[18]. Although the identification system BioLog and the PCR protocol for detection of *S. aureus* require a culture step, they proved to be very reliable methods, especially in the developing countries, where advanced molecular techniques are not yet applicable.

The comparison of the methods for routine detection of *S. aureus* showed that the conventional identification on coagulase activity level alone would result in false determination of 14/73 (19.2%) *S. aureus* isolates - 3 catalase-negative and 10 coagulase-negative, and also of one coagulase-positive identified as *S. schleiferi* subsp. *coagulans*. Special attention is required when working with atypical *S. aureus* strains in udder health laboratories, where the identification systems and PCR based methods are not currently used as diagnostic approaches. In conclusion, this study has shown that a routine approach using a combination of phenotypic and molecular detection systems could improve *S. aureus* detection in milk.

REFERENCES

1. Abeer AA Abdel All, Bashandy MM, Yasin MH, Ibrahim AK: Assessment of conventional and molecular features of *Staphylococcus aureus* isolated from bovine milk samples and contact dairy workers. *Global Veterinaria*, 4, 168-175, 2010.
2. Piccinini R, Tassi R, Daprà V, Pilla R, Fenner J, Carter B, Anjum MF: Study of *Staphylococcus aureus* collected at slaughter from dairy cows with chronic mastitis. *J Dairy Res*, 79, 249-255, 2012.
3. Pate M, Zdovc I, Avberšek J, Ocepek M, Pengov A, Podpečan O: Coagulase-negative staphylococci from non-mastitic bovine mammary gland. *J Dairy Res*, 79, 129-134, 2012.
4. Akineden Ö, Hassan A A, Schneider E, Usleber E: A coagulase-negative variant of *Staphylococcus aureus* from bovine mastitis milk. *J Dairy Res*, 78, 38-42, 2011.
5. Rahimi E, Mommtaz H, Shakerian A, Kavyani HR: The detection of classical enterotoxins of *Staphylococcus aureus* in raw cow milk using the ELISA method. *Turk J Vet Anim Sci*, 36, 319-322, 2012.
6. Layer F, Ghebremedhin B, Moder KA, König W, König B: Comparative study using various methods for identification of *Staphylococcus* species in clinical specimens. *J Clin Microbiol*, 44, 2824-2830, 2006.
7. Grüner BM, Han SR, Meyer HG, Wulf U, Bhakdi S, Siegel EK: Characterization of a catalase-negative methicillin resistant *Staphylococcus aureus* strain. *J Clin Microbiol*, 45, 2684-2685, 2007.
8. Piau C, Jehan J, Leclercq R, Daurel C: Catalase-negative *Staphylococcus aureus* strain with point mutations in the *katA* gene. *J Clin Microbiol*, 46, 2060-2061, 2008.
9. Ahmadi M, Rohani SMR, Ayremlo, N: Detection of *Staphylococcus aureus* in milk by PCR. *Comp Clin Pathol*, 19, 91-94, 2010.
10. Brakstad OG, Aasbakk K, Maeland JA: Detection of *Staphylococcus aureus* by polymerase chain reaction amplification of the *nuc* gene. *J Clin Microbiol*, 30, 1654-1660, 1992.
11. Quinn PJ, Markey BK, Leonard FC, FitzPatrick ES, Fanning S, Hartigan PJ: Veterinary Microbiology and Microbial Disease. 2nd ed., Wiley-Blackwell, J Wiley and Sons Ltd Publication, UK, 2011.
12. Lee N, Chang LC, Chiu CP: A case of carbuncle caused by a catalase-negative strain of *Staphylococcus aureus*. *Diagn Microbiol Infect Dis*, 24, 221-223, 1996.
13. Carvalho ALI, Zanella RC, Yoshikawa LP, Bokermann S, Guerra MLLS, Atobe JH, Lovgren M: Catalase-negative, methicillin-resistant *Staphylococcus aureus* as a cause of septicemia. *Rio de Janeiro*, 39, 45-48, 2003.
14. Dezfulian A, Salehia MT, Amini V, Dabiri H, Azimirad M, Aslani MM, Zali MR, Fazel I: Catalase-negative *Staphylococcus aureus* isolated from a diabetic foot ulcer. *Iran J Microbiol*, 2, 165-167, 2010.
15. Bannerman TL: *Staphylococcus*, *micrococcus*, and other catalase-positive cocci that grow aerobically. In: Manual of Clinical Microbiology. pp.384-404, Washington DC, American Society for Microbiology, 2003.
16. Malinowski E, Lassa H, Klossowska A, Smulski S, Kaczmarowski M: Atypical *Staphylococcus aureus* as an aetiological agent of mastitis in cows. *B Vet J Pulawy*, 53, 383-387, 2009.
17. To KKW, Cheng VCC, Chan JFW, Wong ACY, Chau S, Tsang FHF, Curream SOT, Lau SKP, Yuen KY, Woo PCY: Molecular characterization of a catalase-negative *Staphylococcus aureus* subsp. *aureus* strain collected from a patient with mitral valve endocarditis and pericarditis in the *katA* gene. *J Clin Microbiol*, 49, 3398-3402, 2011.
18. Taponen S, Salmikivi L, Simojoki H, Koskinen MT, Pyörälä S: Real-time polymerase chain reaction-based identification of bacteria in milk samples from bovine clinical mastitis with no growth in conventional culturing. *J Dairy Sci*, 92, 2610-2617, 2009.

Bir Ceylanda (*Gazella gazella*) Generalize Reaktif Amiloidoz Olgusu

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Özet

Bu raporda 1.5 yaşında, erkek bir dağ ceylanında saptanan generalize reaktif amiloidozis tanımlandı. Makroskopik incelemede karın boşluğundaki serozalarda hiperemi ve rumende ise kenarları kanamalı 7 cm çapında ülser belirlendi. Her iki böbreğin yüzeyinde 1-2 mm büyüklüğünde boz-beyaz renkte odaklar gözlemlendi. Mikroskopik incelemelerde böbreklerde glomerulus, tubulus ve akıtıcı kanal bazal membranları ve damar duvarlarında, dalakta foliküllerde, karaciğerde Disse aralığında ve ince barsaklarda lamina propriada pembe renkte, homojen ve amorf yapıda birikimler gözlemlendi. Bu birikimlerin Kongo kırmızısı ile tuğla kırmızısı renkte boyandıkları, potasyum permanganat ile muamele edildikten sonra Kongo kırmızısı ile boyanmadıkları görülerek AA-amiloidoz (sekonder amiloidozis) olduğu kanısına varıldı. %10'luk formaldehitte bulunan stok doku parçalarının Lugol ve sülfirik asitle muamele edilmeleri sonucunda amiloid birikimlerinin koyu mavi renkte oldukları gözlemlendi. Rumende ülserasyonla birlikte generalize amiloidoz saptanan bu olguda, amiloid birikimlerinin makroskopik olarak sadece taze dokularda değil formaldehit solüsyonunda tespit edildikten sonra da gösterilebileceğine dikkat çekilmiştir.

Anahtar sözcükler: Generalize amiloidozis, Ceylan, Patoloji

Generalized Reactive Amyloidosis in a Gazelle (*Gazella gazella*)

Summary

This report was described generalized amyloidosis in a male gazelle, 1.5 years old. In necropsy, hyperaemia on all the serosal surfaces in abdomen, a hemorrhagic ulcer, 7 cm in diameter, on mucosa of rumen, and whitish colored foci, 1-2 mm diameter, on the surfaces of both kidneys were present. Microscopically, there were homogenous pink depositions in the glomeruli, intertubular interstitium, tubular basement membranes and walls of blood vessel in kidney, lymphoid follicles in spleen, space of Disse in liver and lamina propria of the small intestine. These deposits were seen brick-red color stained with Congo red on tissue sections, but these were not stained with Congo red after treated with potassium permanganate. For this reason, AA (secondary) amyloidosis was evaluated. When stock tissue samples stored in 10% formalin were treated with Lugol and acid sulphuric, amyloid depositions were seen as dark blue. Generalized reactive amyloidosis with ulceration in the rumen was diagnosed. Amyloid deposits in the tissues are not only fresh tissue but also tissue stored formalin can be determined as a macroscopic.

Keywords: Generalized amyloidosis, Gazelle, Pathology

GİRİŞ

Amiloid hücre dışında toplanan, çözünmeyen ve proteolitik enzimlerle eritemeyen anormal fibriller bir proteindir. Amiloid protein yapısına göre AL (Amyloid Light Chain) protein ve AA (Amyloid Associated) protein olarak ikiye ayrılır ^[1]. Dağılımına göre sistemik ve lokal, patogeneze göre primer ve sekonder olarak sınıflandırılmaktadır ^[2].

Amiloidoz yabani hayvanlardan Sibiry kaplanlarında, vizonlarda (mink), siyah bacaklı kediler ve siyah bacaklı gelinciklerde ^[2], çitalarda ^[3], Dorcas ceylanında ^[4], Arap ceylanında ^[5], dağ ceylanında ^[6], Rocky dağı büyük boynuzlu koyunlarında ^[7], tavşanlarda ^[8], aslanlarda ^[9], kuşlarda ve


kıvalı kekliklerde ^[10,11], deniz aslanı ^[12], yunus ^[13] ve balinalarda ^[14] bildirilmiştir.

Bu makalede, hayvanat bahçesinde ölen bir dağ ceylanında, organlardaki amiloid birikiminin patolojik yöntemlerle tanımlanması ve amiloidozun tiplendirilmesi amaçlandı.

OLGUNUN TANIMI

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Veteriner Fakültesi Patoloji Anabilim Dalı'na getirilen 1.5 yaşlı erkek bir ceylan oluştu.

Ceylanın nekropsisinde; karın boşluğundaki tüm serozalarda hiperemi, rumende 7 cm çapında ve kenarları kanamalı ülser ile böbreklerde kapsula altında toplu iğne başı büyüklüğünde boz-beyaz renkte odaklar gözlemlendi.

Nekropside alınan parçalar %10'luk formolde tespit edildikten sonra bilinen yöntemlerle takibi yapılarak parafin bloklar elde edildi. Bu bloklardan 5 mikron kalınlığında kesitler alınarak Hematoksilen-Eozin (HE) ve Kongo Kırmızı (KK) ile boyandı [15]. Amiloidin tipini belirlemek amacıyla kesitler önce potasyum permanganat ile muamele edildikten sonra KK ile boyanarak [16] ışık mikroskopunda incelendi.

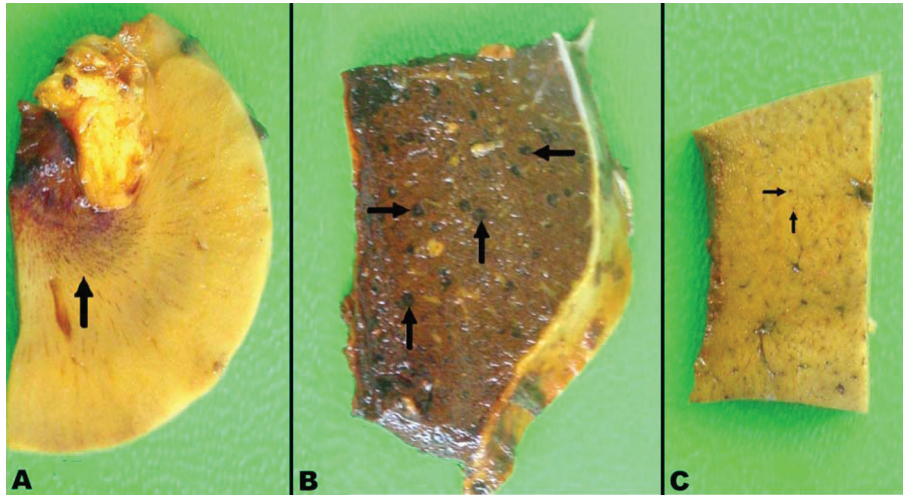
Mikroskopik incelemelerde rumende, çevresinde geniş nekrozlar içeren, ülserasyonlu yangısal granülasyon dokusu tespit edildi (Şekil 4 C-D). Böbreklerde glomerular kapillar damar duvarında belirgin, medullada ise tubulus ve akıtıcı kanal bazal membranları ve interstisyumda damar duvarlarında pembe renkte, homojen ve amorf yapıda birikimler gözlemlendi (Şekil 2 A-C). Benzer birikimlere dalakta sentral arter duvarı ve foliküllerin tamamında (Şekil 3 A-B), karaciğerde Disse aralığında fokal olarak (Şekil 3 D-E), akciğerlerde damar

duvarlarında (Şekil 4 A), ince barsaklarda ise lamina propriyada rastlandı (Şekil 4 B). Bu organların KK ile boyanan kesitlerinde amiloid birikimlerinin tuğla kırmızısı renginde boyandıkları dikkati çekti (Şekil 2 D-F, Şekil 3 B,E). Diğer organ kesitlerinde ise KK ile boyamalarda herhangi bir boyanma tespit edilmedi. Alınan kesitlerin potasyum permanganat ile muamele edildikten sonra KK ile boyanmadıkları (Şekil 3 C,F) ve bu birikimlerinin reaktif (sekonder) amiloidoziste tanımlanan AA-protein yapısında olduğu kanısına varıldı.

Histopatolojik incelemeler sonucunda amiloid belirlenen dokuların %10'luk formolde saklanan parçalarına Lugol ve sülfirik asit damlatıldığında amiloidli kısımların koyu mavi renk aldıkları dikkati çekti (Şekil 1 A-C).

TARTIŞMA ve SONUÇ

Yaban hayvanlarında genellikle sekonder amiloidozla ilişkin vaka raporları bildirilmektedir [2]. Dorcas ceylanlarında amiloidozun genellikle kronik enfeksiyonlar sonucunda geliştiği [4], ancak genetik predispozisyonun da önemli olabileceği bildirilmiştir [2]. Bir dağ ceylanı [6] ve bir Arap ceylanındaki [5] amiloidozun sebebinin ise idiyopatik olduğu öne sürülmüştür. Çeribaşı ve ark.[17] bir İran ceylanında fibrino-



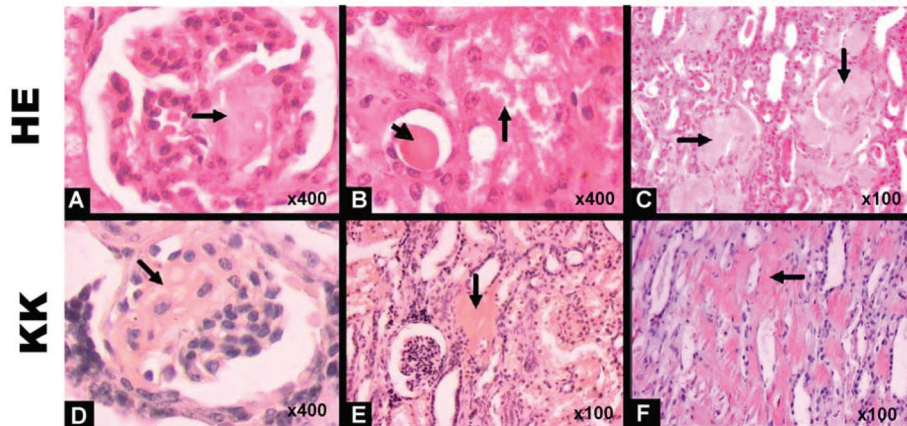
Şekil 1. Amiloid birikimleri, Lugol- sülfirik asit boyama. A- Böbrek (ok), B- Dalak (oklar), C- Karaciğer (oklar)

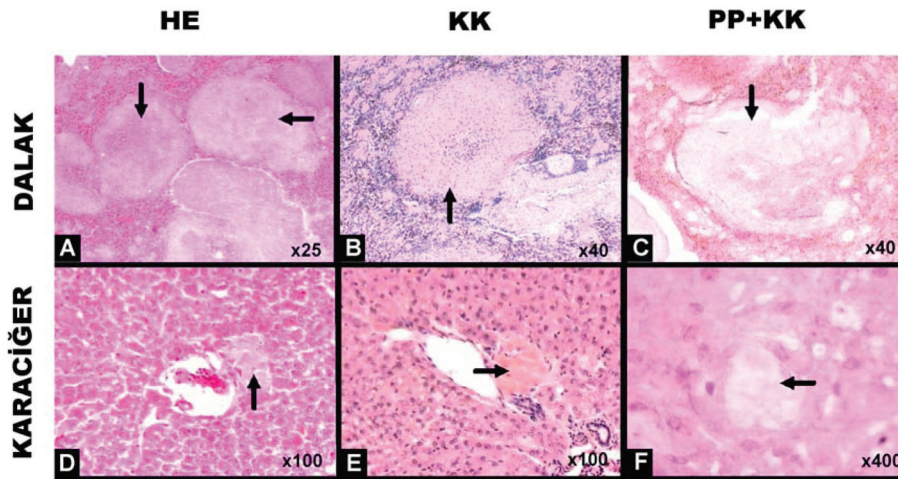
Fig 1. Amyloid deposits, Lugol-sulfuric acid staining. A- Kidney (arrow), B- Spleen (arrows), C- Liver (arrows)

BÖBREK

Şekil 2. Böbrekte amiloid birikimleri. A,D- Glomerulus (ok), B- Tubulus lümenlerinde hiyalin silindiri (okbaşı) ve epitellerde hiyalin damlacıkları (ok), C,E,F- İntertubuler interstisyumda amiloid birikimleri (oklar). Hematoksilen-Eozin (HE), Kongo Kırmızısı (KK)

Fig 2. Amyloid deposits in kidney. A,D- glomeruli (arrow), B- Hyaline cylinder in tubule lumens (arrowhead) and hyaline droplets in tubule epithelium (arrow), C,E,F- Amyloid deposits in intertubular interstitium. Hematoxylin-eosin (HE), Congo Red (KK)



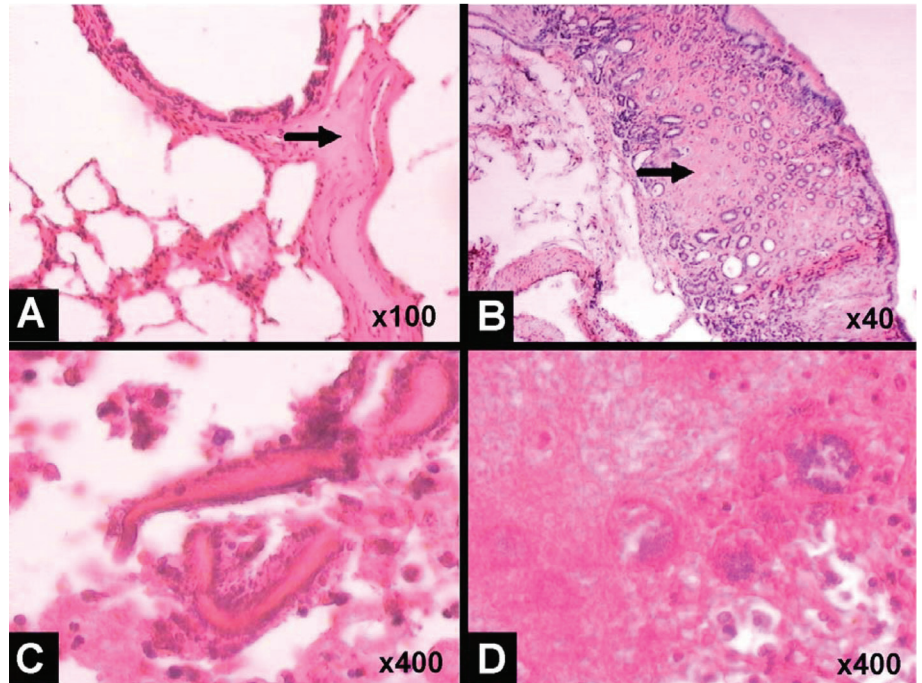


Şekil 3. Dalakta (A,B) ve karaciğerde (D,E) amiloid birikimleri. Sekonder amiloidozda dalak (C) ve karaciğerdeki (F) birikimlerin boya almaması. Potasyum Permanganat metodu [potasyum permanganat (PP) + Kongo Kırmızısı (KK)]

Fig 3. Amyloid deposits in spleen (A,B) and liver (D,E). Deposits in spleen (C) and liver (F) were not stained with potassium permanganate methode (PP + KK) on secondary amyloidosis

Şekil 4. A- Akciğerde interstisyumda amiloid birikimi (ok), HE, B- Bağırsaklarda lamina propriyada amiloid birikimi (ok), KK, C,D- Rumende epitelde nekroz, HE

Fig 4. A- Amyloid deposition in the lung interstitium (arrow), HE, B- Amyloid deposits in the lamina propria in intestine (arrow), Congo Red, C,D- Epithelial necrosis in rumen, HE



hemoajik pankreatitisle komplike generalize amiloidoz olgusunu tanımlamışlardır. Sunulan çalışmada, rumende geniş ülserle karakterize kronik yangının organlardaki amiloid birikiminin nedeni olabileceği düşünülmüştür.

Amiloidoz şüpheli dokuların nekropsisi sonrası Lugol ve sülfirik asit ile muamele edildiğinde amiloid birikimlerinin mavi renkte görüldüğü bildirilmektedir ^[1]. Sunulan bu olguda amiloid birikimiyle ilgili organlarda makroskopik bir bulgu gözlenmemesi nedeniyle nekropsiden sonra bu uygulamaya gerek duyulmamıştır. Histopatolojik olarak amiloid birikimlerinin saptanması üzerine formaldehit solüsyonu içerisinde stok olarak bekletilen karaciğer, dalak ve böbrek dokularının kesit yüzlerine Lugol ve sülfirik asit damlatılması sonucu benzer şekilde mavi renkte amiloid birikimleri gözlenmiştir. Amiloidozlu dokuların uzun süre formolde bırakılması sonucu amiloidin boyanma özelliğinin azalabileceği, ancak kısa süreli tespitlerde iodinle yeniden

boyanmanın mümkün olabileceği bildirilmiştir ^[18,19]. Bu sonuçla, formaldehit tespitinin amiloid birikiminin Lugol ve sülfirik asit muamelesiyle makroskopik teşhisini etkilemediği, nekropsisi sonrası sadece taze dokularda değil formaldehitte tespit edilen dokularda da makroskopik olarak amiloidozun teşhis edilebileceğini göstermektedir.

Amiloidoza bağlı ölümlerin genellikle böbrek yetmezliği sonucu olduğu öne sürülmekte olup; Papendick ve ark.^[3] amiloidozlu çitaların %74'ünde, Colegrove ve ark.^[12] ise Kaliforniya denizaslanlarının çoğunda amiloidoza bağlı renal yetmezlik oluştuğunu bildirmişlerdir. Bu çalışmada, her iki böbrekte korteks ve medulladaki amiloid birikimlerinin renal yetmezliğe neden olabileceğini düşündürmektedir.

Bu çalışmada bir dağ ceylanında generalize reaktif amiloidoz (AA-amiloidoz) olgusu makroskopik ve mikroskopik bulgularla tanımlanmış, amiloid birikimlerinin rumende

ülserli kronik yangıyla ilişkili olabileceği değerlendirilmiş ve amiloid birikimlerinin makroskopik olarak formol tespiti-
tinden sonra da gösterilebileceğine dikkat çekilmiştir.

KAYNAKLAR

1. Erer H, Kıran MM, Çiftçi MK: Veteriner Genel Patoloji. 3. Baskı, Bahçivanlar Basım Sanayi, Konya, 2009.
2. Woldesmeskel M: A concise review of amyloidosis in animals. *Vet Med Int*, Article ID 427296, 11 pages, doi:10.1155/2012/427296, 2012.
3. Papendick RE, Munson L, O'Brien TD, Johnson KH: Systemic AA amyloidosis in captive cheetahs (*Acinonyx jubatus*). *Vet Pathol*, 34 (6): 549-556, 1997.
4. Rideout BA, Montali RJ, Wallace RS, Bush M, Phillips LG, Antonovych TT, Sabnis SG: Renal medullary amyloidosis in dorcas gazelles, *Vet Pathol*, 26 (2): 129-135, 1989.
5. Fox RI, Payne-Johnson CE, Sanderson S: Identification of generalised (AA) amyloidosis in a Arabian gazelle (*Gazella gazella* sp). *Eur J Vet Pathol*, 7 (3): 123-125, 2001.
6. Linke RP, Hol PR, Geisel O: Immunohistochemical identification of generalized AA-amyloidosis in a mountain gazelle (*Gazelle gazelle*). *Vet Pathol*, 23, 63-67, 1986.
7. Hadlow WJ and Jellison WL: Amyloidosis in Rocky Mountain bighorn sheep. *J Am Vet Med Assoc*, 141 (2): 243-247, 1962.
8. Giesel O, Linke RP: Generalized AA-amyloidosis in two Hares (*Lepus europaeus*) immunohistochemically identified using poly-and monoclonal antibodies. *Vet Pathol*, 25, 391-393, 2012.
9. Williams JH, Van Wilpe E, Momberg M: Renal medullary AA amyloidosis, hepatocyte dissociation and multinucleated hepatocytes in a 14-year-old free-ranging lioness (*Panthera leo*). *J S Afr Vet Assoc*, 76 (2): 90-98, 2005.
10. Sato A, Koga T, Inoue M, Goto N: Pathological observations of amyloidosis in swans and other anatidae. *Jpn J Vet Sci*, 43, 509-519, 1981.
11. Terim Kapakin KA, Gürsan N, Kutsal O: Generalised amyloidosis in partridge (*Alectoris chukar*). *Kafkas Univ Vet Fak Derg*, 16 (1): 143-146, 2010.
12. Colegrove KM, Gulland FMD, Naydan KHDK, Lowenstine LJ: Pathological features of amyloidosis in stranded California sea lions (*Zalophus californianus*). *J Comp Path*, 140, 105-112, 2009.
13. Cowan DF: Amyloidosis in the Bottlenose dolphin (*Tursiops truncatus*). *Vet Pathol*, 32, 311-314, 1995.
14. Tajima Y, Shimada A, Yamada TK, Cowan DF: Amyloidosis in two Stejneger's beaked whales (*Mesoplodon stejnegeri*) stranded at the sea of Japan. *J Zoo Wild Med*, 38, 108-113, 2007.
15. Luna LG: Manual of Histologic Staining Methods of The Armed Forces Institute of Pathology. McGraw-Hill Book Company, New York, 1968.
16. Wright JR, Calkins E and Humphrey RL: Potassium permanganate reaction in amyloidosis. *Lab Invest*, 36 (3): 274-281, 1977.
17. Ceribasi AO, Eroksuz Y, Ceribasi S, Ozer H: Generalized AA amyloidosis and fibrino-hemorrhagic pancreatitis in a *Gazelle subgutturosa*: A case report. *Vet Med-Czech*, 52 (7): 340-344, 2009.
18. Drury RAB, Wallington EA: Carleton's Histological Techniques. 4th ed., pp.194-195, Oxford University Press, London, UK, 1976.
19. Brundeleit PJ: Long lasting amyloid staining techniques for museum specimens. *Acta Pathol Microbiol Scand*, 59 (2): 156-160, 1963.

Türkiye’de Köpeklerde *Babesia canis canis*’in Klinik ve Parazitolojik Olarak İlk Tespiti

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Özet

Bu makalede, Giemsa boyama ve Polimeraz Zincir Reaksiyonu (PZR) yöntemleri ile parazitolojik olarak *Babesia canis canis* tanısı konulan 3 köpekte klinik, biyokimyasal ve hematolojik bulgularının sunulması amaçlanmıştır. Olgu I ve II’de şiddetli ikterus, anemi, hemoglobinem, hemoglobinüri, trombositopeni, taşipne, taşikardi ve miyozitis ile karaciğerde hasar geliştiği tespit edildi. Olgu I’de bu bulgulara ek olarak kolesistitis, Olgu II’de ise renal hasarın olduğu saptandı. Olgu III’te hafif anemi ve ikterus tablosuyla birlikte karaciğerde hasar ve miyozitis şekillendiği belirlendi. Tedavi sonucunda Olgu II ve III’de iyileşme sağlanırken Olgu I öldü. Sonuç olarak, köpeklerde *B. canis canis* klinik ve parazitolojik olarak Türkiye’de ilk kez bildirilmektedir.

Anahtar sözcükler: *Babesia canis canis*, *Babesiosis*, Polimeraz Zincir Reaksiyonu, PZR, Köpek, Kars, Türkiye

Clinical and Parasitological Detection of *Babesia canis canis* in Dogs: First Report from Turkey

Summary

This study was aimed to describe clinical, biochemical and haemathological findings in three dogs with *Babesia canis canis* diagnosed by Giemsa staining and Polymerase Chain Reaction (PCR) methods. In Case I and II severe icterus, anemia, haemoglobinemia, haemoglobinuria, thrombocytopenia, tachypnea, tachycardia, liver damage and myositis were determined. In addition in Case I cholecystitis and in Case II renal damage were developed. In Case III mild anemia and icterus along with liver damage and myositis were developed and clinical signs were mild. Following treatment Case II and III completely recovered while Case I died. In conclusion, *B. canis canis* has been determined clinically and parasitologically for first time in Turkey.

Keywords: *Babesia canis canis*, *Babesiosis*, Polymerase Chain Reaction, PCR, Dog, Kars, Turkey

GİRİŞ

Kanin babesiosis kenelerin naklettiği protozoal parazit olan *Babesia* türleri tarafından oluşturulan bir enfeksiyondur. Köpek *Babesia* türleri piroprasmik formlarının morfolojilerine göre, büyük (5 x 2.5 µm) ve küçük (2 x 1.5 µm) olarak iki grupta sınıflandırılmaktadır. Büyük türler *B. canis canis*, *B. canis vogeli*, *B. canis rossi* ve *Babesia* sp. (Coco), küçük türler ise *B. gibsoni*, *B. conradae* ve *B. microti-like* (*Theileria annae*)’dan oluşmaktadır ^[1-3]. Son yıllarda *B. c. canis*, *B. c. rossi* ve *B. c. vogeli*’nin, her ne kadar morfolojik olarak çok benzeseler de, klinik belirtilerinin değişkenlik göstermesi, ayrı yayılış alanlarına ve farklı vektörlere

sahip olmaları nedeniyle, tek başlarına birer tür (*B. canis*, *B. vogeli*, *B. rossi*) olarak kabul edilmeleri eğilimi ağırlık kazanmıştır ^[2,4-6]. Perakut, akut, kronik ve subklinik seyir gösteren kanin babesiosis’de klinik belirtiler tür, vektör, ırk, yaş ve immun duruma göre değişebilmektedir ^[1-3,6-11]. *B. c. canis* orta derecede patojenik özellik gösterirken, *B. canis vogeli* daha hafif hastalık tablosuna neden olur. *B. c. canis* ve *B. gibsoni* ilerleyici hemolitik anemi yaparken, *B. canis rossi* disemine intravasküler koagulopati (DIC) ile birlikte şok, sistemik yangı ve organ bozukluğuna yol açmaktadır ^[2,8,11]. Bilgilerimize göre Türkiye’de *B. c. canis*’in



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neden olduğu babesiosis vakası bildirilmemiştir^[1,3,12,13]. Bu raporda, *B. c. canis*'in neden olduğu babesiosis'in teşhis, klinik, biyokimyasal ve hematolojik bulgularının sunulması amaçlanmıştır.

OLGULARIN TANIMI

Olgu materyalini Kafkas Üniversitesi Veteriner Fakültesi İç Hastalıkları Anabilim Dalı kliniklerine iştahsızlık, durgunluk ve sarılık şikayetleriyle getirilen farklı yaş, cinsiyet ve ırkta 3 köpek oluşturdu. Köpeklerden tedavi öncesi ve tedaviden 15 gün sonra kan alınarak hemogram değerleri ve biyokimyasal parametrelere bakıldı. Hemogram değerleri MS4e marka hemogram cihazında, serumda biyokimyasal parametreler ise otoanalizörde (Kodak Vitros DT60II) ölçüldü. İdrar analizinde Cybow 10 M idrar stripi kullanıldı.

OLGU I

İki yaşında, dişi sokak köpeğinin incelendiği birinci olguda hastada 4 gün önce başlayan ve giderek şiddetini arttıran bir iştahsızlık olduğu öğrenildi. Hastada ayrıca ayakta durmakta zorlanma, çevreye karşı ilgisinin olmaması ile birlikte idrarının kırmızı renkte olduğu tespit edildi. Hayvanın üzerinde çok sayıda kene ile birlikte, klinik olarak apati, şiddetli ikterus, yüksek ateş (41.3°C), hemoglobinuri, solunum ve nabız sayısının artış belirlendi. Hemogramda ise eritrosit (RBC), hemoglobin (Hb), hematokrit (HCT) değerlerinde ve trombosit sayısında şiddetli düşüş saptandı. Biyokimyasal analizlerde alkalen fosfataz (ALP), gamma gamma glutamilttransferaz (GGT), alanin aminotransferaz (ALT), aspartat aminotransferaz (AST), kreatin kinaz (CK),

laktat dehidrogenaz (LDH), total bilirubin ve direkt bilirubin değerlerinin yükseldiği tespit edildi (Tablo 1). İdrar muayenesinde bilirubinüriye rastlandı. Bu bulgulara göre olguda şiddetli anemi, trombositopeni ve myozitis şekillendiğinin belirlenmesinin yanında hastada kolangio-hepatitis geliştiği belirlendi.

OLGU II

İkinci Olguyu 4 gündür halsizlik, iştahsızlık, durgunluk şikayetleri olan 3 yaşında erkek husky oluşturdu. Hastada ayrıca depresyon, sallantılı yürüyüş, çevreye karşı ilgisizlik, ikterus, solunum ve nabız sayısında artış, yüksek ateş (40.5°C), hemoglobinuri belirlendi. Hayvanın derisinde farklı bölgelerde 8 adet keneye rastlandı. Hastanın RBC, Hb, HCT ve trombosit değerlerinde belirgin düşüş olduğu saptandı. Serum biyokimyasında ALP, CK, üre, kreatinin, total bilirubin ve direkt bilirubin değerlerinin yüksek olduğu belirlendi (Tablo 1). Elde edilen verilere göre hayvanda şiddetli anemi, trombositopeni ve miyozitis şekillendiği belirlendi. İdrar analizinde ise bilirubinuri ve proteinüri tespit edildi. Ayrıca mevcut verilere göre hastada renal ve hepatik hasarın gelişmiş olabileceği düşünüldü.

OLGU III

İştahsızlık, durgunluk şikayetleriyle getirilen 5 yaşında erkek kangal ırkı köpeğin incelendiği III. Olguda inguinal bölgede 3 adet keneye rastlanırken, hastanın çevreye karşı ilgisinin azaldığı, mukozalarında hafif ikterusla birlikte solgunluk ve yüksek ateşi (40.6°C) olduğu belirlendi. Hematolojik muayenede RBC, Hb, HCT değerlerinde hafif

Tablo 1. Olgularda tedavi öncesi ve sonrası bazı hematolojik ve biyokimyasal parametrelerindeki değişiklikler

Table 1. Changes in some haematological and biochemical parameters in cases before and after treatment

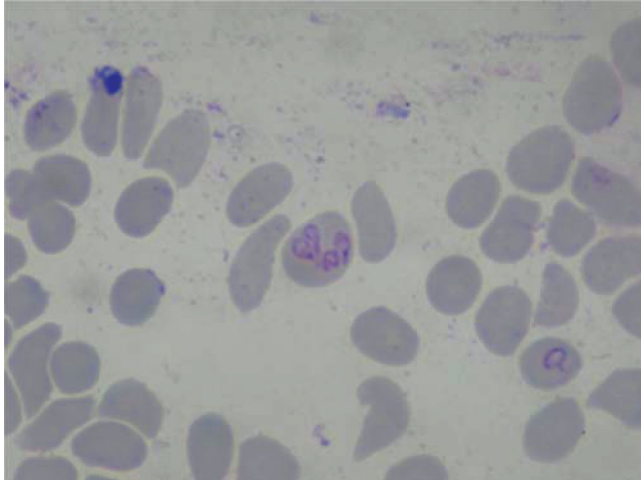
Parametreler	Referans Değerler ^[14]	Olgu I* TÖ	Olgu II TÖ	Olgu II TS	Olgu III TÖ	Olgu III TS
Hematolojik	WBC (x10 ³ /µL)	5.5-16.9	6.01	5.33	10.53	13.15
	RBC (x10 ⁶ /µL)	5.5-8.5	1.79	3.37	7.73	4.18
	HGB (g/dL)	12-18	4.4	8.7	11.9	10.9
	HCT (%)	37-55	12.3	23.1	36.4	31.6
	PLT (x10 ³ /µL)	175-500	11	6	248	341
Biyokimyasal	ALP (U/L)	20-150	840	424	142	214
	GGT (U/L)	1-10	20	5	3	3
	ALT (U/L)	10-88	250	56	53	47
	AST (U/L)	10-88	188	61	52	63
	CK (U/L)	20-200	236	277	152	684
	LDH (U/L)	50-495	977	750	103	3779
	Üre (mg/dL)	25-55	52.5	107.1	50.2	41.9
	Kreatinin (mg/dL)	0.5-1.5	1.2	2.8	0.9	0.86
	Total Protein (g/dL)	5.4-7.7	5.6	4.3	7.4	5.2
	Albumin (g/dL)	2.3-3.8	2.2	1.8	3.5	2.7
	Total Bilirubin (mg/dL)	0.1-0.6	16.1	12.4	0.12	0.19
	Direkt Bilirubin (mg/dL)	0-0.3	>7	6.5	0.1	0.05

TÖ: Tedavi öncesi, TS: Tedavi sonrası *Olgu I eks olduğu için TS değerler bulunmamaktadır

düşüş saptandı. Biyokimyasal analizlerde ise ALP, CK ve LDH değerlerinin yüksek olduğu belirlendi (Tablo 1). Hematolojik ve biyokimyasal analizler sonucunda köpekte hepatik hasar gelişmiş olabileceği düşünüldü. Bunun dışında miyozitis ve hafif anemi şekillendiği anlaşıldı.

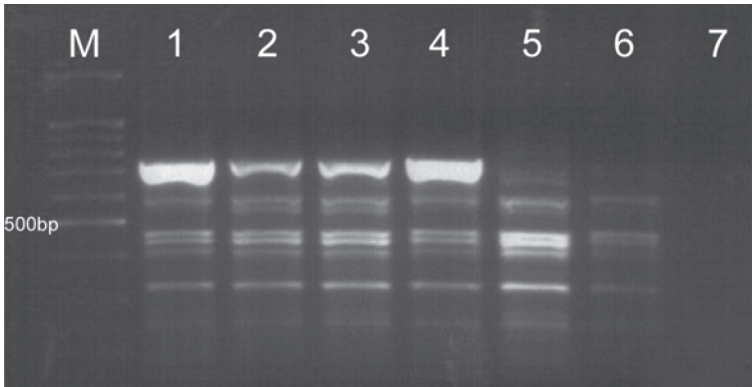
Her üç hayvanın üstünden toplanan kenelerin *Dermacentor reticulatus* olduğu anlaşılmış, yapılan sürme perifer kan frotilerinin Giemsa boyama sonuçlarında, mikroskopta eritrositler içinde büyük yapıda, çift veya çoklu piroplazmik formdaki *Babesia* etkenleri görülmüştür (Şekil 1).

Klinik belirtiler ve parazitolojik laboratuvar muayenesi (Giemsa boyama) sonucunda babesiosis teşhisi konulan olgular tedaviye alındı. Tedavide her üç köpeğe 6 mg/kg İM yolla tek doz imidokarb (İmicarp®, TeknoVet) uygulandı. Destekleyici tedavi amacıyla B kompleksi ve C vitaminleri ve intravenöz yolla laktatlı ringer + dekstroz verildi. Olgu 1 tedavi başladıktan bir gün sonra öldü. Hasta sahibinin istememesi nedeniyle nekropsi yapılamadı. İki ve üçüncü olgularda ise tedavi başladıktan 15 gün sonra hastaların iyileştiği, hematolojik ve biyokimyasal değerlerin normale döndüğü belirlendi (Tablo 1).



Şekil 1. Giemsa ile boyanmış sürme kan preparatında görülen tipik büyük *Babesia* etkenleri

Fig 1. Typical large *Babesia* agents detected in blood smear with Giemsa stain



İncelenen olgularda etkenlerin tür düzeyinde doğrulaması, *B. canis canis*'e özgü primerler kullanılarak PZR testi ile yapıldı. Hayvanlardan EDTA'lı tüplere alınan kan örneklerinden ticari kit (High Pure Nucleic Acid Isolation Kit, Roche) kullanılarak üreticinin bildirdiği şekilde DNA ekstraksiyonu yapıldı. Polimeraz Zincir Reaksiyonu testinde Duarte ve ark.'nın^[15] belirlediği ve 18S rRNA gen bölgesinden *B. c. canis*'e özgün 746 bp'lik bölgeyi çoğaltan Bab1 5'-GTG AAC CTT ATC ACT TAA AGG-3' ve Bab3 (5'-CTA CAC AGA GCA CAC AGC C-3') primerleri kullanıldı. Reaksiyon miktarı 25 µl olarak belirlendi ve her reaksiyon için 18.25 µl steril su, 2.5 µl 10x PCR buffer (Invitrogen), 1 µl 50 mM MgCl₂, 0.5 µl dNTP miks (Invitrogen), 20 pmol/µl konsantrasyondaki her primerden 0.5 µl, 0.25 µl Taq DNA polymerase 5U/µl (Invitrogen) ve 1.5 µl DNA örneği kullanıldı. Reaksiyon Biometra TGradient PCR cihazında toplam 35 döngü (94°C'de 30 sn, 56°C'de 30 sn, 72°C'de 45 sn) üzerinden yapıldı. İlk döngüden önce 94°C'de 2 dk denaturasyon ve en son döngüden sonra da 72°C'de 10 dk ekstensiyon basamakları uygulandı. Reaksiyon sonuçları %1.5 Agaroz jelde yürütülerek UV ile görüntüldü. Türe özgü PZR sonucunda, her 3 olguda da özgün 746 bp'lik bantlar oluşurken, kontrol olarak kullanılan *B. c. rossii* ve *B. c. vogeli* DNA'larından benzer bant elde edilmemiştir (Şekil 2).

TARTIŞMA ve SONUÇ

Köpeklerde *B. c. canis*'in neden olduğu babesiosis özellikle Orta Avrupa ve Rusya'da yaygın olarak görülmekte olup, son yıllarda kuzeye doğru bir yayılım göstererek Norveç ve Hollanda gibi ülkelerde de rapor edilmeye başlanmıştır^[1,2,16,17]. Etken *D. reticulatus* tarafından nakledilmekte olup, hastalık bu kenenin yayılım alanları ile ilişkilidir^[1,2]. Ülkemizde, günümüze kadar, genellikle *Rhipicephalus sanguineus* tarafından nakledilen *B. c. vogeli* ve *B. gibsoni* varlığı bildirilmiştir^[3,12,13]. Öte yandan, çalışmamızda babesiosisli köpeklerde *B. c. canis*'in tespiti Türkiye için ilk rapor olup, gerek etkenin gerekse de *D. reticulatus*'un ülkemizde belli bölgelerde kalıcı popülasyon oluşturduğuna işaret eder niteliktedir.

Etkene ve komplikasyonlara göre değişmekle birlikte babesiosisli köpeklerde genellikle anoreksi, depresyon, akut

Şekil 2. PZR sonuçları; M: 100bp DNA marker, 1- Olgu-I, 2- Olgu-II, 3- Olgu-III, 4- *B. canis canis* kontrol DNA, 5- *B. canis rossii* kontrol DNA, 6- *B. canis vogeli* kontrol DNA, 7- Negatif kontrol (su)

Fig 2. PCR results; M: 100bp DNA marker, 1- Case-I, 2- Case-II, 3- Case-III, 4- *B. canis canis* control DNA, 5- *B. canis rossii* control DNA, 6- *B. canis vogeli* control DNA, 7- Negative control (water)

hemoliz, hemoglobinüri, ikterus, taşipne, taşikardi, splenomegali değişen derecelerde hemolitik anemi ve trombositopeni görülmektedir ^[1,7,11,13]. Bu raporda klinik tablonun ağır seyrettiği olgularda (Olgu I ve II) anoreksi, apati, yüksek ateş, ikterus, hemoglobinüri, şiddetli anemi, trombositopeni, solunum ve nabız sayısında artış görüldü. Daha hafif klinik tabloya sahip olan köpekte ise (Olgu III) iştahsızlık, mukozalarda hafif ikterusla birlikte solgunluk ve ateşin yüksek olduğu belirlendi.

Kanin babesiosis'te en sık karşılaşılan hematolojik bulguları HCT değerinin düşmesi, eritrositopeni ve hemoglobinemidir ^[1,2,9,10]. Sunulan raporda da II. Olguda hematokrit değerinde belirgin düşüş, şiddetli eritrositopeni ve hemoglobinemide belirlenirken I. Olguda bu değerlerdeki düşüşün daha hafif olduğu görüldü. Kanin babesiosis'de en sık karşılaşılan bozukluklardan biri olan trombositopeninin ^[9-11] Olgu I ve II'de şiddetli olduğu belirlenirken Olgu III'te trombosit değerinin normal sınırlarda olduğu saptandı. Son yıllarda, *B. c. canis*'in kendi içinde farklı virulansa sahip iki genetik grup içerdiği ve bunların farklı hematolojik bulgulara, özellikle de farklı şiddette trombositopeniye neden olduğu bildirilmiştir ^[18]. Bizim olgularımızdaki farklılığın kesin olarak bundan kaynaklanıp kaynaklanmadığının ortaya konması için daha ayrıntılı çalışmaya ihtiyaç vardır. Yapılan bazı çalışmalarda *B. c. canis* enfeksiyonlarında lökopeni şekillendiği ^[7,11] bildirilmiş olsa da, bu çalışmada her üç olguda da lökosit sayısının referans değerler arasında olduğu görüldü.

Babesiosis'te böbrek yetmezliği, hepatopati ve miyozitis gibi çoklu organ bozuklukları gelişebilmektedir ^[9,11,19]. Olgu I'de ALT ve AST enzim değerlerinin yüksek olması, olgu II ve III'te ALP enzim değerinin yüksek bulunması üç olguda da karaciğerde olası bir hasar geliştiğine işaret etmektedir. Olgu I de ALP ve GGT enzimlerinin yüksek bulunması köpekte kolestazis'in hepatik hasara eşlik ettiği şeklinde yorumlanabilir. *B. c. canis* enfeksiyonlarında kolestazis şekillenmesi hakkında bir bilgiye rastlanmamıştır. Bu nedenle bu bulgunun önemli olduğu kanısındayız. Her üç olguda da CK ve LDH enzimlerinin yüksek bulunması miyozitis oluştuğunu göstermiştir. Yine literatürlerle uyumlu olarak ^[8,11,19] Olgu II'de üre ve kreatinin değerlerinin yüksek olması ve idrarda proteine rastlanması renal hasar gelişmiş olabileceği gösteren veriler olarak değerlendirilebilir.

Sunulan raporda *B. c. canis* enfeksiyonunun önemli biyokimyasal değişikliklerinden olan hiperbilirubinemi ve hemoglobinin ^[7,20], Olgu I ve II'de şiddetli geliştiği tespit edildi. Bu olgularda hemoglobininüri ve bilirubinüri gelişmesinin nedeni eritrositlerdeki aşırı hemolize bağlı olarak dolaşımda fazla miktarda Hb ve bilürübin bulunması, bu maddelerin renal eşiği aşmasından kaynaklanabilir.

Bu çalışma ile *B. c. canis* Türkiye'den ilk kez rapor edilmiştir. Olguların yerli hayvanlar olması ve bölgede de *D. reticulatus* popülasyonunun varlığı (yayınlanmamış bulgular), etkenin Kars gibi alpin iklimde sahip yerlerde

yerleşik hal aldığına işaret etmektedir. Bu çalışmada ayrıca, hastaların klinik ve biyokimyasal bulguları da izlenerek, hastalık konusunda ayrıntılı veriler elde edilmiştir.

KAYNAKLAR

1. Matijatko V, Torti M, Schetters TP: Canine babesiosis in Europe: How many diseases? *Trends Parasitol*, 28, 99-105, 2012.
2. Solano-Gallego L, Baneth G: Babesiosis in dogs and cats-expanding parasitological and clinical spectra. *Vet Parasitol*, 181, 48-60, 2011.
3. Aysul N, Ural K, Ulutaş B, Eren H, Karagenc T: First detection and molecular identification of *Babesia gibsoni* in two dogs from the Aydın province of Turkey. *Turk J Vet Anim Sci*, 37, 226-229, 2013.
4. Uilenberg G, Franssen FF, Perie NM, Spanjer AA: Three groups of *Babesia canis* distinguished and a proposal for nomenclature. *Vet Q*, 11, 33-40, 1989.
5. Schetters TP, Moubri K, Precigout E, Kleuskens J, Scholtes NC, Gorenflot A: Different *Babesia canis* isolates, different diseases. *Parasitology*, 115, 485-93, 1997.
6. Zahler M, Schein E, Rinder H, Gothe R: Characteristic genotypes discriminate between *Babesia canis* isolates of differing vector specificity and pathogenicity to dogs. *Parasitol Res*, 84, 544-548, 1998.
7. Solano-Gallego L, Trotta M, Carli E, Carcy B, Caldin M, Furlanello T: *Babesia canis canis* and *Babesia canis vogeli* clinicopathological findings and DNA detection by means of PCR-RFLP in blood from Italian dogs suspected of tick-borne disease. *Vet Parasitol*, 157, 211-212, 2008.
8. Ruiz de Gopegui R, Penalba B, Goicoa A, Espada Y, Fidalgo LE, Espino L: Clinicopathological findings and coagulation disorders in 45 cases of canine babesiosis in Spain. *Vet J*, 174, 129-132, 2007.
9. Matijatko V, Kis I, Torti M, Brkljacic M, Kucer N, Rafaj RB, Grden D, Zivcicjak T, Mrljak V: Septic shock in canine babesiosis. *Vet Parasitol*, 162, 263-270, 2009.
10. Schetters TP, Kleuskens JA, Van DCJ, De LPW, Finizio AL, Gorenflot A: Systemic inflammatory responses in dogs experimentally infected with *Babesia canis*: A haematological study. *Vet Parasitol*, 162, 7-15, 2009.
11. Matijatko V, Kiş I, Torti M, Brkljačić M, Rafaj RB, Žvorc Z, Mrljak V: Systemic inflammatory response syndrome and multiple organ dysfunction syndrome in canine babesiosis. *Veterinarski Arhiv*, 80, 611-626, 2010.
12. Gulamber A, Gorenflot A, Schetters TP, Carcy B: First molecular diagnosis of *Babesia vogeli* in domestic dogs from Turkey. *Vet Parasitol*, 139 (1-3): 224-230, 2006.
13. Ulutaş B, Bayramli G, Ulutaş PA, Karagenc T: Serum concentration of some acute phase proteins in naturally occurring canine babesiosis: A preliminary study. *Vet Clin Pathol*, 34, 144-147, 2005.
14. Turgut K: Veteriner Klinik Laboratuvar Teşhis. II. Baskı, Bahçıvanlar Basım Sanayi AŞ, Konya, 2000.
15. Duarte SC, Linhares GF, Romanowsky TN, da SNOJ, Borges LM: Assessment of primers designed for the subspecies-specific discrimination among *Babesia canis canis*, *Babesia canis vogeli* and *Babesia canis rossi* by PCR assay. *Vet Parasitol*, 152, 16-20, 2008.
16. Matjila TP, Nijhof AM, Taoufik A, Houwers D, Teske E, Penzhorn BL, de Lange T, Jongejan F: Autochthonous canine babesiosis in The Netherlands. *Vet Parasitol*, 131, 23-29, 2005.
17. Oines O, Storli K, Brun-Hansen H: First case of babesiosis caused by *Babesia canis canis* in a dog from Norway. *Vet Parasitol*, 171, 350-353, 2010.
18. Adaszek L, Winiarczyk S, Skrzypczak M: The clinical course of babesiosis in 76 dogs infected with protozoan parasites *Babesia canis canis*. *Pol J Vet Sci*, 12, 81-87, 2009.
19. Matijatko V, Mrljak V, Kis I, Kucer N, Forseck J, Zivcicjak T, Romić Z, Simec Z, Ceron JJ: Evidence of an acute phase response in dogs naturally infected with *Babesia canis*. *Vet Parasitol*, 144, 242-250, 2007.
20. Furlanello T, Fiorio F, Caldin M, Lubas G, Solano-Gallego L: Clinicopathological findings in naturally occurring cases of babesiosis caused by large form *Babesia* from dogs of northeastern Italy. *Vet Parasitol*, 134, 77-85, 2005.

A Case Report: Isolation of *Alysiella filiformis* from Pig's Lungs ^[1]

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Summary

Alysiella filiformis is considered a common resident in the oral cavities of many animals. All reports of *Alysiella* indicate that it is restricted to the oral cavity of warm-blooded vertebrates, where it apparently is nonpathogenic. However, increased losses of young pigs occurred in one farm in Serbia. Spumous content in bronchia and partly clotted blood in blood vessels of the lungs were present. Characteristic signs of oedema disease were present and *E. coli* serogroup 0139 was isolated. Furthermore, *Alysiella filiformis* was the single agent isolated from the lungs of diseased pigs. This is the first isolation of *Alysiella filiformis* from pig lungs.

Keywords: *Alysiella filiformis*, Pig, Lungs

Olgu Sunumu: Domuz Akciğerlerinden *Alysiella filiformis* İzolasyonu

Özet

Alysiella filiformis birçok hayvanın ağız boşlukları ortak bir sakini olarak kabul edilir. Literatüre göre etkenin sıcakkanlı hayvanların ağız boşluğu ile sınırlı ve patojenik olmadığı bildirilmiştir. Sırbistan bir çiftlikte genç domuz kayıpları artışı gözlenmiştir. Postmortem incelemede akciğer kan damarları ve bronşlarda kısmen pıhtılaşmış kan ve köpüklü içerik gözlenmiştir. Ödem hastalığının karakteristik işaretler mevcut olduğunu ve *E. coli* serogrup 0139 izole edildi. Ayrıca, *Alysiella filiformis* hastalıklı domuz akciğerlerinden izole edilmiş tek bir ajan idi. Bu çalışmada hastalıklı domuz akciğerlerden *Alysiella filiformis* izolasyonu ilk kez bildirilmiştir.

Anahtar sözcükler: *Alysiella filiformis*, Domuz, Akciğerler

INTRODUCTION

Alysiella filiformis (*A. filiformis*) is considered a common resident in the oral cavities of many animals. All reports of *Alysiella* indicate that it is restricted to the oral cavity of warm-blooded vertebrates, where it apparently is nonpathogenic. *A. filiformis* is the solitary member of genus *Alysiella*, family *Neisseriaceae* ^[1,2]. Genus *Alysiella* comprises organisms that exist in characteristic flat, ribbon-like multicellular filaments. The cells within the filament are paired, and in axenic culture the filament often breaks up into groups of two or four cells. The width of an individual cell is about 2.0–3.0mm, and the length of a cell is about 0.6 mm ^[1,2]. *A. filiformis* has been isolated from the oral cavity of different animals including guinea pigs ^[3], sheep ^[4], and cows ^[5], and in Turkey it was isolated from hot spring

water indicating its presence in the environment ^[6]. This paper presents a case of the isolation of *Alysiella filiformis* from the lungs of pig with oedema disease.

CASE HISTORY

During May 2009, one swine farm with 1.000 pigs situated in northwest of Serbia, reported increased losses of young pigs. According to anamnestic data, live affected pig showed a staggering gate, puffy eyelids giving a sleepy appearance and an abnormal high pitched squeak. During nine days, 14 pigs (weight around 15 kg) in very good condition died, and dispnea was the only clinical symptom present in diseased pigs. Post-mortem examinations showed



İletişim (Correspondence)



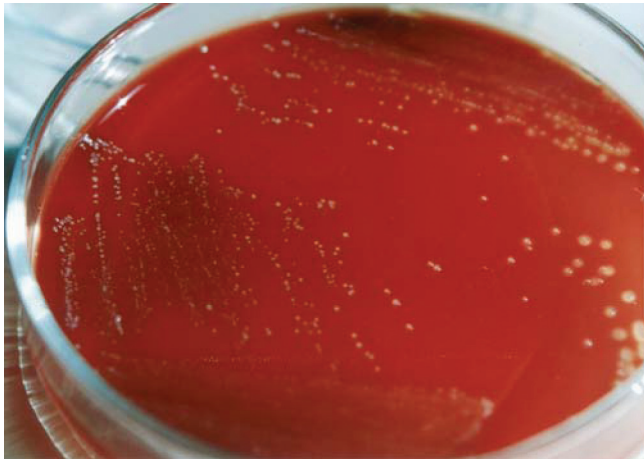
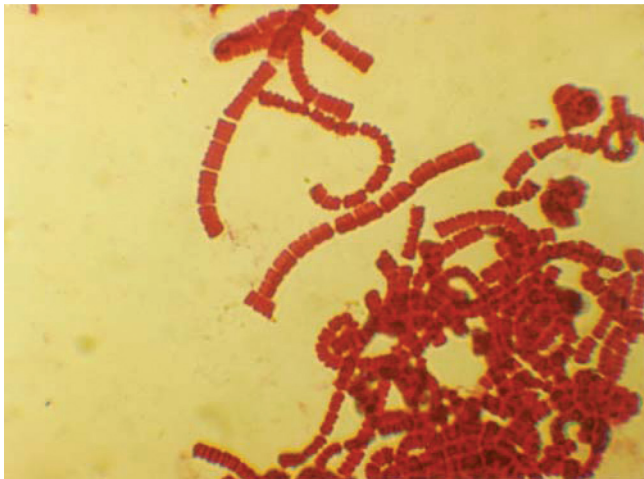
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Table 1. Biochemical characteristics of *Alysiella filiformis* isolate**Tablo 1.** *Alysiella filiformis* biyokimyasal özellikleri izole

Medium	Reaction	Medium	Reaction
Catalase	positive	Inositol	acid not produced
Oxidase	positive	Xylose	acid not produced
Glucose	acid produced	Arabinose	acid not produced
Maltose	acid produced	Mannitol	acid not produced
Trehalose	acid produced	Gelatin	negative
Sucrose	acid produced	H ₂ S	not produced
Fructose	acid produced	MR	negative
Lactose	acid not produced	VP	negative
Cellobiose	acid not produced	Indole	negative
Raffinose	acid not produced	Urease	negative
Rhamnose	acid not produced	Aesculin	not hydrolyzed
Mannose	acid not produced	Nitrate	not reduced
Dulcitol	acid not produced	Nitrite	not reduced
Salicin	acid not produced	Simmon's citrate	no growth

**Fig 1.** Colonies of *A. filiformis* on blood agar**Şekil 1.** Kan agar *A. filiformis* kolonileri**Fig 2.** Ribbon-shaped filaments of *A. filiformis* (Gram stain, x1.000)**Şekil 2.** *A. filiformis* şerit şeklindeki filamentler (Gram boyama, x1.000)

oedema of the greater curvature of the stomach wall, coiled colon and eyelids. Spumous content in bronchia and partly clotted blood in blood vessels of the lungs were present.

Samples of small intestines were inoculated on nutritive agar (HiMedia) containing 5% sheep blood and MacConkey agar (HiMedia), and incubated aerobically at 37°C for 24 h. Lung samples were also inoculated on 5% sheep blood agar and incubated aerobically at 37°C for 24 h. The cultural, biochemical and morphological characteristics of grown cultures were examined. Bacterial isolates were identified using standard methods for phenotypic characterization as previously described [1,2,7,8]. After 24 h of incubation, *E. coli* was grown on sheep blood agar and MacConkey agar from the small intestine samples. Using antisera (*E. coli* O antisera, HiMedia), isolate was identified to belong to serogroup O139. From lung samples, no growth was seen on MacConkey agar, but large numbers of smooth, convex, β -haemolytic, white-grayish colonies, about 1.5mm diameter, were present in pure culture on sheep blood agar.

After subcultivation on MacConkey agar at 37°C and 25°C, as well as on sheep blood agar at 25°C, culture was sterile, but bacterial growth occurred after subcultivation on sheep blood agar at 37°C. Gram staining of isolate revealed gram-negative cells, 0.6 x 2-3 μ m, arranged side by side in pairs to form flat, ribbon-shaped filaments, with variable length.

Isolate was further tested by biochemical tests (bioMérieux, HiMedia).

On the basis of the cell morphology, cultural and biochemical characteristics, isolate was identified as *Alysiella filiformis*.

DISCUSSION

In difference from the other representatives of this family, cells of genera *Simonsiella* and *Alysiella* may exhibit a characteristic multicellular micromorphology. Moreover, the unusual morphology of the *Alysiella* filaments serves to differentiate the genus from all other procaryotic organisms. *Alysiella* filaments differ from *Simonsiella* in consisting of continuous pairs of cells instead of being segmented into units of eight cells, and the terminal cells are not rounded, but square [2]. In this case, *A. filiformis* was isolated from lung tissue in pure culture, with ribbon-shaped filaments of with variable length, similar to other authors [2]. The biochemical properties of bacteria isolated from lung tissue in this study are very similar to those described in other studies [2,7,8].

The origin and significance of the presence of *A. filiformis* in the lung tissue is not clear. *A. filiformis* has been described as a comensal in oral cavity of animals [3-5], but

there is no report of its presence in other tissues or organs. Also, there is no known pathology as a consequence of presence of this bacterium. Because of this fact, it is hard to determine the role bacterium in described macroscopic lesions present in lung tissue of diseased pig. In the present case, suspicion of possible role of *A. filiformis* in lung pathology was based on fact that it was isolated from lung tissue in pure culture, with abundant growth and presence of clearly visible zone of beta-haemolysis around colonies.

The reasons for the colonization of lung tissue with *A. filiformis*, and its potential role in the etiology of pathoanatomical changes in lung tissue remain unclear. Further studies are needed to determine the pathogen potential of this bacterium.

REFERENCES

- Holt JG, Krieg NR, Sneath PHA, Staley JT, Williams ST:** Group 15: Nonphotosynthetic, nonfruiting gliding bacteria. **In**, William R Hensyl (Ed): Bergey's Manual of Determinative Bacteriology. 9th ed., pp.489-491, Williams & Wilkins, Baltimore, Maryland, 1994.
- Tønnum T, Family I Neisseriaceae:** The Proteobacteria. Part C: The Alpha-, Beta-, Delta-, and Epsilonproteobacteria. **In**, Brenner DJ, Krieg NR, Staley JT, Garrity GM (Eds): Bergey's Manual of Systematic Bacteriology. Vol. 2, 2nd ed., pp. 775-801, Springer, New York, USA, 2005.
- Berger U:** Reinzüchtung von *Simonsiella* spp. *Z Hyg*, 149, 336-340, 1963.
- Steed PDM:** *Simonsiellaceae* fam. nov. with characterization of *Simonsiella crassa* and *Alysiella filiformis*. *J Gen Microbiol*, 29, 615-624, 1962.
- McCowan RP, Cheng KJ, Costerton JW:** Colonization of a portion of the bovine tongue by unusual filamentous bacteria. *Appl Environ Microbiol*, 37, 1224-1229, 1979.
- Sariözlü NY, Demirel R, Kivanc M:** Microbial population of hot spring waters in Eskişehir/Turkey. *J Sci Technol*, 2, 31-39, 2012.
- Heiske A, Muters R:** Differentiation of selected members of the family Neisseriaceae (*Alysiella*, *Eikenella*, *Kingella*, *Simonsiella* and CDC groups EF-4 and M-5) by carbohydrate fingerprints and selected phenotypic features. *Zentralbl Bakteriell*, 281, 67-79, 1994.
- Hedlund BP, Kuhn DA:** The genera *Simonsiella* and *Alysiella*. The Proteobacteria. **In**, Dworkin M, Falkow S, Rosenberg E, Schleifer KH, Stackebrandt E (Eds): The Prokaryotes A Handbook on the Biology of Bacteria. pp.828-839, Springer, New York, USA, 2006.

Possible Natural Toxins in Organic Livestock Farming

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Summary

Public concern about food quality has intensified in recent years. In the last decades, the amount of farmland managed under certified organic practices has expanded dramatically and it is expected to increase in the future. There is a growing demand for organic foods driven primarily by consumers' perceptions of the quality and safety of these foods and to the positive environmental impact of organic agriculture practices. This growth in demand is expected to continue in the future. Obviously many factors in production systems have an impact on the health and welfare of the animals involved. Animals feed or forage may be the source of disorders for farm animals also could lead to human illness. The types of feeds administered do not differ significantly in organic farming and in this respect there are mostly quantitative differences observed. Pesticides, agricultural and industrial chemicals, heavy metals, radionuclides as well as natural pollutants as mycotoxins may pollute animal feed. The aim of this review was to find out the possible natural toxins of animals associated with organic farming.

Keywords: Organic farming, Feeding, Natural toxins, Pollutants

Organik Hayvancılıkta Olası Doğal Toksinler

Özet

Son yıllarda halkın gıda kalitesi ile ilgili endişeleri artmıştır. Son dönemlerde sertifikalı organik üretim uygulamaları yapılan alanlar önemli ölçüde genişlemiş ve gelecekte daha da artacağı tahmin edilmektedir. Organik ürün eldesiyle ilgili uygulamaların çevreye olan olumlu etkileri, elde edilen ürünlerin gıda kalitesi ve güvenliği ile ilgili tüketicilerin algıları bu ürünlere olan talebi giderek artırmaktadır. Organik ürünlere olan sözkonusu talebin gelecekte de artacağı tahmin edilmektedir. Organik üretim sistemindeki birçok uygulamanın hayvan sağlığı ve refahı üzerine olan etkileri açıktır. Hayvan yemlerinin çiftlik hayvanlarında bazı metabolik bozuklukların kaynağı olabilir, hatta insanlarda da hastalıklara yol açabilir. Organik hayvansal üretimde hayvanlara verilen yem çeşidinin niteliğinde belirgin bir fark yokken, genellikle miktarda nicelik farklılıkları gözlenmektedir. Pestisidler, tarımsal ve endüstriyel kirleticiler, ağır metaller, radyoaktif bileşikler ile doğal kirleticilerden mikotoksinler de dahil olmak üzere birçok etken hayvan yemini kirlitebilirler. Bu derlemenin amacı, organik hayvancılıkta olası doğal toksinler hakkında bilgi sunmaktır.

Anahtar sözcükler: Organik tarım, Beslenme, Doğal toksinler, Kirleticiler

INTRODUCTION

The industrial agriculture system consumes fossil fuel, water, and topsoil at unsustainable rates ^[1]. The expansion of factory style animal agriculture creates environmental and public health concerns, including pollution from the high concentration of animal wastes and the extensive use of antibiotics, which may compromise their effectiveness in medical use. The type of agriculture that has become conventional throughout the industrialized world is, in historical terms, a new phenomenon ^[2-5]. Humans have practiced agriculture for more than 10000 years, but only in the past 50 years or so have farmers become heavily

dependent on synthetic chemical fertilizers and pesticides and fossil fuel-powered farm machinery. In the last decades, however, industrial agriculture has increasingly separated animals from the land ^[6].

Organic farming is the result of theory and practice since the early years of the 20th century, involving various alternative methods of agricultural production, mainly in northern Europe ^[7-9]. Organic livestock farming is based on the principle of a close link between the animals and the soil. The need for a link with the soil requires animals



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to have free access to outside areas for exercise, and also implies that their feed should be not only organic, but preferably produced on the farm. This sector of organic farming is, also, strictly regulated by rules of animal welfare and veterinary care ^[10-13]. Organic farming was developed and driven by people searching for a sustainable way of farming. They often sold their products directly to consumers, bypassing the many marketing steps between the farm and the consumer. Today the situation is very different ^[14-18]. Organic farming is likely to receive a major boost in the European Union (EU) and most probably also worldwide since consumers have lost some trust in food derived from conventional production. However, this growth in organic farming is also expected to continue in the presumable future in Turkey ^[19]. This is as a result of recent crises and due to concerns apropos the use of pesticides in farming and antibiotics in livestock feed ^[11,20-23].

European Council Regulation (EEC) No 2092/91 on organic production of agricultural products and suggestions referring on agricultural products and foodstuffs was adopted on 24 June 1991. In 1999, the Council adopted Regulation (EC) No 1804/1999 of 19 July 1999, which lays down Community rules for producing organic livestock products; this completes the framework of Community legislation, which now covers both crop and animal products. Part B of Annex I to Regulation (EEC) No 2092/91, as amended on 19 July 1999 by Regulation No 1804/1999, lays down minimum rules for organic livestock production. The Member States may adopt stricter rules, under Article 12 of Regulation (EEC) No 2092/91, concerning the animals and animal products produced on their territory ^[13,24].

As the nutritional quality of organic foods for humans there seems to be no difference in nutritional quality between an organic diet and conventional diet, though the perception of consumers is that organically-produced crops and animal products are of higher nutritional quality and safety ^[25-28]. The small number of papers published is not surprising since the development of organic farming and its philosophy ^[29]. Therefore, this article is limited to review the possible natural toxins that represent significant risks to organic livestock.

1. ORGANIC FARMING AND ANIMAL FEEDING

To get insight into the different types of feeds used in organic and conventional production systems information was obtained from different sources on the internet. This because the information in scientific articles was too limited and fragmented ^[11,27,29]. From the information assessed it is clear that a major difference between organic and conventional production of feeds is that in organic production only limited crop production and other products are permitted. A remark should be made the

natural source of the products is no guarantee for non-toxicity for humans ^[30-33].

International food safety standards and food hygiene requirements are equally valid for conventionally and organically produced food. The issue of other food quality characteristics is less clear-cut. There is a widely-held view that the food industry is best placed to make decisions about the quality of their products based on their understanding of market demands ^[9,18,34]. However, compulsory and optional quality standards do exist to ensure that essential product requirements are met and to protect consumers against fraudulent practices ^[35,36]. Most problems that occur in conventional agriculture may also be present in organic farming, such as erosion, nitrogen leaching, ammonia volatilisation from animal wastes, high levels of native soil cadmium, accumulation of trace metals in soil, and subsoil compaction caused by farm machinery ^[37-41]. Organic farming methods do not offer solutions to many of these problems. In contrast to conventional agriculture, organic farming without purchase of feed may result in a nutrient depletion of soils. Through the import of feeding stuff to farms, which means a net input of nutrients, depletion is normally avoided. As the feeding stuff may be produced elsewhere with inorganic fertilizers, organic farming indirectly depends on the soil fertility of conventional farming ^[42,43]. However, regulations about the conventionally grown feeding stuff to be used in organic farming differ between countries ^[24,34,44,45].

For cattle the types of feeding of the animals do not differ between organic and conventional systems. Feeding of organic cattle should however be derived from organic production systems ^[46-48]. Also the percentage of mixed feeds and raw feeds differ in organic cattle production. For example 60% of the feeds should be raw feeds for organic cows. For organic cows also no synthetic additives, antibiotics etc. are allowed. Rules for pigs and chickens are similar ^[15,20,49]. In conclusion the major differences in the quality of feeds between organic and conventional products systems seems to originate from differences in production of plants (such as limited use of crop production products) and the lack of certain additives in the feeds. The types of feeds administered do not differ significantly and there are mostly quantitative differences observed (more raw feeds) ^[13,14,27,50,51].

2. ENVIRONMENTAL CONTAMINANTS

One reason for the increase in organic agriculture in many countries in Europe today is we need to solve environmental problems. In such situations, we often tend to accept appealing solutions. Furthermore, the intensive propaganda by representatives of organic farming movements has had a strong influence on public opinion,

politicians, and scientists^[38,44,52,53]. A wide range of organic and inorganic compounds may occur in feedstuffs, including pesticides, industrial pollutants, radionuclides and heavy metals. Pesticides that may contaminate feeds originate from most of the major groups, including organochlorine, organophosphate and pyrethroid compounds. Although pesticides are potentially toxic to farm livestock, the primary focus of concern centers on residues in animal products intended for human consumption^[31,54-59]. It should be noted that organic producers are not prohibited from using all pesticides -certain pesticides from natural sources can be used. Natural pesticides like the chemically synthesised pesticides must be subject to safety evaluation. Natural pesticides used in organic management are usually restricted under certification schemes. International guidelines for organically produced foods include lists of substances that can be used for plant pest and disease control if the need for such is recognised by the certification body. In organic management, biological control is the preferred method of pest management^[13,35,50].

Contaminants in animal feeds, such as pesticide residues, agricultural and industrial chemicals, heavy metals and radioactive nuclides, can result in safety hazards in foods of animal origin. As EC regulations (EC No 1804/1999) require that livestock, claimed to be produced organically, is fed on organically produced feed stuffs, the potential for contamination with pesticide residues and other agricultural chemicals is greatly reduced compared to conventional farming methods^[3,39,60]. However, organic agriculture does not reduce the levels of persistent environmental pollutants in organically grown products. These may therefore be present in organic feedstuffs and hence in organic food of animal origin^[13,32,35,61]. Conversely, excluding pesticides may result in increased concentrations of secondary plant metabolites and of mycotoxins of field fungi. Thus, to exclude pesticides does not necessarily mean that crop products do not contain unwanted substances^[31,44].

3. POSSIBLE NATURAL TOXINS OF ANIMAL FEEDS

Animal products are a primary source of proteins, amino acids and fats, and when they are a major constituent of diets they contribute a significant part of total calories. They are, however, perishable products which require special attention to prevent their deterioration and contamination by various agents, biological as well as chemical. Some of these contaminants can be transferred by the feed^[22,62-64]. Animal feeds are routinely subject to contamination from diverse sources, including environmental pollution and activities of insects and microbes. Animal feeds may also contain endogen toxins arising principally from specific primary and secondary substances produced by fodder plants^[65-67]. Thus, feed toxins include compounds of both plant and microbial origin. Although these toxins are often

considered separately, because of their different origins, they share several common underlying features. Therefore, particular compounds within both plant and microbial toxins may exert antinutritional effects or lessen reproductive performance in farm animals^[23]. Also, the combined effects may be the result of additive or synergistic interactions between the two groups of compounds. The extent and impact of these interactions in practical livestock feeding remain to be quantified. Feed contaminants and toxins occur on a global scale but there are distinct geographical differences in the relative impact of individual compounds. The term "feed" is generally used in its widest context to include compound blends of straight ingredients as well as forages. With such a broad perspective, it is necessary and more instructive to introduce some focus^[54,59,68].

3.1. Contamination from Natural Fertilisers

Variation in stocking rates of grazing ruminants can change the structure and composition of pastures with potential impacts on biodiversity and produce methane, a greenhouse gas^[69,70]. The search for reducing agricultural surpluses led the EU to promote livestock production extensification and a decrease in stocking rates on grasslands. The value of species rich grasslands are also reinforced by the increased interest of consumers in site-specific and origin-labelled products, and the growing scientific evidence of the role of local grassland flora on various sensory characteristics of both cheese and meat products, such as colour and flavour of cheese and meat, respectively^[34,71,72]. Animal manure and other organic waste are the main fertilisers used in organic farming. These natural fertilisers are also widely used in conventional agriculture with chemically synthesised fertilisers. Microbiological contamination arising from the use of natural fertilisers and measures needed to address it must focus on both organic and conventional agriculture. Untreated or improperly treated manure or biosolids used as fertilisers or soil nutrient agents, whether in organic or nonorganic agriculture, can lead to contamination of products water sources^[42,53,73,74]. Animal and human faecal matters are known to contain human pathogens. Properly treated manure or biosolids are effective and safe fertilisers. Growers need to follow good agricultural practices for handling these natural fertilisers to minimise microbial hazards. Researches indicates that pathogenic organisms can survive up to 60 days under compost conditions. The Codex General Principles of Food Hygiene provide the basic rules for ensuring food safety for all foods. Organic production, as with all other types of food production, must follow the rules outlined in this international code of practice. However, for some, drift and runoff from near fields may result in microbial hazards. Growers may consider scheduling application of manure on near fields to maximise the time between manure application to those fields and harvest of fresh market products^[5,35,50,51,53].

3.2. Bacterial Contamination

There is considerable interest in the occurrence of *Escherichia coli* (*E. coli*) in animal feeds following the association of the O157:H7 type of these bacteria with human illness [2]. The US Centre for Disease Control (CDC) identifies the main source of human infection with *E. coli* as meat contaminated during slaughter. Virulent strains of *E. coli*, such as *E. coli* O157:H7 develop in the digestive tract of cattle, which is mainly fed with starchy grain. Replication of faecal *E. coli*, including the O157:H7 type was demonstrated in various feeds under conditions likely to occur on cattle farms in the summer months [2,8,49]. Since faecal contamination of feeds is widespread on farms, it is an important route for exposure of cattle to *E. coli* and other organisms. The potential for exposure to bacteria also exists when poultry litters are fed to cattle. Cows mainly fed with hay produce less than 1% of the *E. coli* found in the faeces of grain-fed animals. It is one of the most important goals of organic farming to keep the nutrient cycles closed. Therefore, ruminants like cattle and sheep are fed diets with a high proportion of grass, silage and hay. It can be concluded that organic farming potentially reduces the risk of *E. coli* infection [55,56,75-78]. *Listeria monocytogenes* occur in poor-quality silages and big-bale silage. When grass is ensiled under anaerobic conditions, the low pH regime ensures that *Listeria spp.* is excluded from the resulting silage. However, in big-bale silage a degree of aerobic fermentation may occur, raising pH levels and allowing the growth of *Listeria spp.* These bacteria also survive at low temperatures and in silages with high levels of dry matter. Contamination of silage with *Listeria spp.* is important as it causes abortion, meningitis, encephalitis and septicaemia in animals and humans. The incidence of various forms of listeriosis has been increasing in recent years [79-81].

3.3. Mycotoxins

There are consistent reports of worldwide contamination of feeds with fungi and their spores. In the tropics,

Aspergillus is the predominant genus in dairy and other feeds [82,83]. Other species include *Penicillium*, *Fusarium* and *Alternaria*, which are also important contaminants of cereal grains. Fungal contamination is undesirable because of the potential for mycotoxin production. However, spores from mouldy hay, silage, brewers' grain and sugar beet pulp may be inhaled or consumed by animals with destructive effects termed "mycosis" [31,59,84-86]. Most important mycotoxins were resumed in Table 1 [87].

Mycotoxins are metabolites produced by fungi of various genera while they grow on agricultural products before or after harvest or during transport or storage. Mycotoxin contamination of forages and cereals often occurs in the field following infection of plants with particular pathogenic fungi or with symbiotic endophytes. Contamination may also occur during processing and storage of harvested products and feed whenever environmental conditions are appropriate for spoilage fungi [88,89]. Mycotoxins are regularly found in animal feed ingredients such as maize, sorghum grain, rice meal, cottonseed meal, groundnuts and legumes, wheat, barley and others. Most are relatively stable compounds and are not destroyed by the processing of feed and may even be concentrated in screenings. Some fungi such as *Aspergillus spp.* and *Penicillium spp.* can invade grain after harvest and produce mycotoxins, while others, such as *Fusarium spp.*, typically infest grains and produce mycotoxins before harvest [85,90-92]. In some circumstances *Aspergillus spp.* can grow and produce mycotoxins before the crop is harvested. Both intrinsic and extrinsic factors influence fungal growth and mycotoxin production. The intrinsic factors include moisture content and acidity, whereas extrinsic factors also include appropriate substrates (rice, corn, nuts, wheat, and food and feeds originated from them) and production period for 3-6 days [35,57,93-95]. There is only limited information available on the occurrence of mycotoxin residues in animal products intended for human consumption. It is known that milk cows can convert Aflatoxin B₁ into

Table 1. Common mycotoxins, commodity affected, and negative effects on health

Tablo 1. Sıklıkla karşılaşılan mikotoksinler, etkilenen ürünler ve sağlığa olan olumsuz etkileri

Mycotoxin	Commodities	Fungal Source(s)	Effects of Ingestion
Aflatoxin B ₁ , B ₂ , G ₁ , G ₂	Corn, peanuts, and many other commodities	<i>Aspergillus flavus</i> <i>Aspergillus parasiticus</i>	Aflatoxin B ₁ identified as potent carcinogen by IARC ^a Adverse effects in various animals especially chickens
Deoxynivalenol Nivalenol (Vomitoxin)	Wheat, corn and barley	<i>Fusarium graminearum</i> <i>Fusarium crookwellense</i> <i>Fusarium culmorum</i>	Human toxicoses in India, China, Japan, and Korea Toxic to animals, especially pigs
Zearalenone	Corn, wheat	<i>Fusarium graminearum</i> <i>Fusarium culmorum</i> <i>Fusarium crookwellense</i>	Identified as possible carcinogen by IARC. Affects reproductive system in laboratory animals and pigs
Ochratoxin A	Barley, wheat and many other commodities	<i>Aspergillus ochraceus</i> <i>Penicillium verrucosum</i>	Suspected by IARC as human carcinogen Carcinogenic in laboratory animals and pigs
Fumonisin B ₁	Corn	<i>Fusarium moniliforme</i> and several less common species	Suspected by IARC as human carcinogen. Toxic to pigs and poultry. Cause of equine leukoencephalomalacia (ELEM) a fatal disease of horses

^a: International Agency for Research on Cancer

Aflatoxin M₁, which is found in milk. This has caused considerable trade problems when Aflatoxin M₁ is found in milk, and concern about the safety of such milk [35,57,96].

Mycotoxicoses are diseases caused by exposure to foods or feeds contaminated with mycotoxins. Mycotoxins show various biological effects in animals, such as liver and kidney toxicity, central nervous system effects or estrogenic effects. There are differences between animals on the susceptibility towards different mycotoxins [68,85,97]. Poultry secrete mycotoxins relatively fast because of a particular digesting system. The ingredients used for animal feeding should be checked to ensure that satisfactory quality standards are maintained and that mycotoxins are not present at higher than acceptable levels [96,98]. Further research is needed to study the metabolism of mycotoxins by animals and the residues of mycotoxins and their metabolites in animal tissues. However, in many instances the problems of mycotoxins in feeds have more direct effects that they can create illness in animals and prevent efficient growth or feed use [49,84,89].

Since fungicides are not allowed in organic production and given that mycotoxins constitute a major health hazard, their relative presence in foods produced organically or conventionally has been the subject of many studies. From these studies it cannot be concluded that organic farming leads to an increased risk of mycotoxin contamination. It is important to emphasise that good agricultural, handling and storage practices are required in organic as in conventional agriculture to minimise the risk of mould growth and mycotoxin contamination [35,91,98,99]. Several research teams in EU have carried out comparative surveys of the frequency and levels of mycotoxins in conventional

and organic foods. The results are surprisingly consistent. Averaged across 24 direct comparisons of mycotoxins in conventional and organic foods in published studies, mycotoxins were detected in conventional food about 50 per cent more often than in corresponding organic food [11,31]. Mycotoxin levels in conventional food averaged a little over twice as high as in the corresponding organic foods. The probable explanations for the higher levels of mycotoxins in the conventional wheat crops grown in EU is the routine use of high levels of nitrogen fertilizer, and fungicide applications to prevent diseases and losses [31].

It has been suggested that organically produced food has higher levels of mycotoxin contamination because organic farming bans the use of fungicides. There is no evidence to support this claim [98,99]. In fact organic farmers would contend that their crops are less prone to fungal diseases because high doses of nitrogen increase the growth rate of crops leading to a thinning of the plant cell walls making the crop more vulnerable to fungal attack [49,99,100]. Good animal feeding practices also require that feed is stored in such a way to avoid contamination. As organically raised livestock are fed greater proportions of hay, grass and silage, there is reduced opportunity for mycotoxin contaminated feed to lead to mycotoxin contaminated milk [35,89,98].

3.4. Potential Plant Toxins

Many plant components have the potential to precipitate adverse effects on the productivity of farm livestock. These compounds are present in the foliage seeds of almost every plant that is used in practical feeding [101]. Typical concentrations of selected toxins were presented in Table 2 [59].

Table 2. Plant toxins: sources and concentrations

Tablo 2. Bitkisel toksinler: kaynakları ve konsantrasyonları

Toxin	Principal Source(s)	Typical Concentrations
Lectins	Jackbean	73 units/mg protein
	Winged bean	40-320 units/mg
	Lima beans	59 units/mg protein
Trypsin inhibitors	Soybean	88 units/mg
Antigenic proteins	Soybean	-
Cyanogens	Cassava root	186 mg HCN/kg
Condensed tannins	Acacia spp.	65 g/kg
	Lotus spp.	30-40 g/kg
Quinolizidine alkaloids	Lupin	10-20 g/kg
Glucosinolates	Rapeseed	100 mmol/kg
Gossypol	Cottonseed	0.6-12 g/kg (free)
Saponins (steroidal)	<i>Brachiaria decumbens</i> , <i>Panicum spp.</i>	-
S-methyl cysteine sulfoxide	Kale	40-60 g/kg
Mimosine	<i>Leucaena leucocephala</i>	145 g/kg (seed)
		25 g/kg (leaf)
Phyto-oestrogens	Clover, lucerne, soybean	-

Plant toxins may be divided into heat-labile and heat-stable groups. Heat-labile group comprising lectins, proteinase inhibitors and cyanogens, which are sensitive to standard processing temperatures. Heat-stable group includes many compounds such as antigenic proteins, condensed tannins, quinolizidine alkaloids, glucosinolates, gossypol, saponins, nonprotein amino acids (such as S-methyl cysteine sulfoxide and mimosine), and phyto-oestrogens^[102-104]. The role of these substances as anti-nutritional factors has been considered in detail by D'Mello^[59]. Contamination of animal feeds with weed seeds is also a major problem worldwide. The impact of weed seeds arises from the toxins they contain and from their diluent effects on nutrient density of feeds. Examples of weed seeds that are controlled by legislation in various countries include those of *Datura spp.*, common vetch, castor-oil plant and *Crotalaria spp.*^[35,54,59].

4. CONTROL OF FEEDBORNE HAZARDS

Over many years feeding animals and preparing feed ingredients has not received adequate quality and safety attention. FAO organized in 1997 an Expert Consultation on Animal Feeding and Food Safety to determine how to better address the problems of feed ingredients and contaminated feed^[54]. The primary purpose of this Expert Consultation was to discuss current animal feed problems, and to develop a draft Code of Practice for Good Animal Feeding for consideration by the Codex Alimentarius Commission, as advice to FAO member countries. This draft Code covers good animal feeding practices, and adherence to Good Manufacturing Practices (GMPs) in the procurement, handling, manufacturing, storage and distribution of commercially-produced feeds for food-producing animals. Feed and feed ingredients should be obtained and preserved in stable conditions to prevent hazardous effects due to contamination or deterioration. When received, feeds should be in good condition and meet generally accepted quality standards. GMPs should be followed always^[54,84].

Preventive management of mycotoxin contamination of food and animal feed should be checked regularly in organic farming. Strategies to prevent mycotoxin contamination must be applied strictly. It is important for producers to realise that good agricultural practices (GAPs) represent the primary line of defence against contamination of cereals with mycotoxins, followed by the implementation of GMPs during the handling, storage, processing, and distribution of cereals for human food and animal feed^[105]. Many strategies to prevent mycotoxin contamination of food and animal feed have been developed. Various crop genotypes resistant to fungus infection^[106-108], pre-harvest control strategies^[89,109], field and harvest management as well as post-harvest applications including improving drying and storage conditions^[110-113],

and the use of biological agents^[114-116] have been shown important in the prevention of mycotoxigenic mould growth and mycotoxin formation. Preservation can be facilitated by low temperature storage, ensiling, dehydration or the addition of appropriate chemicals^[54]. However, several natural plant extract and spice oils of eugenol, cinnamon, oregano, onions, lemongrass, turmeric and mint are known to prevent both mould growth and mycotoxin formation during post-harvest season^[112,117-119]. Although activated charcoal, hydrated sodium calcium aluminosilicate, aluminosilicate, zeolite, and bentonite have shown good potential for use in the animal feed to help overcome aflatoxicosis. Recently, there has been an increasing interest in the use of bacteria, yeast, and fungi to help reduce the toxic effect of mycotoxins^[114,120,121]. Another approach to the problem has been the use of mycotoxin-binding agents in the diet that sequester the mycotoxin in the gastrointestinal tract thus reducing their bio-availability^[94,95,119,122]. The suitability of these applications may be questionable regarding the organic farming regulations. At the same time it should be noted that chemical treatment is not allowed within the EU for commodities destined for human consumption.

More information may be found in the first edition of the "Prevention and Reduction of Food and Feed Contamination" publication^[105] that contains all the codes of practice related to the prevention and reduction of contaminants in foods and feeds adopted by the Codex Alimentarius Commission until 2011.

CONCLUSION

There is a growing demand for organic foods driven primarily by a consumer's perceptions of the quality and safety of these foods and to the positive environmental impact of organic agriculture practices. This growth in demand is expected to continue in the foreseeable future. Animal feed, including herbage, may be contaminated with organic and inorganic compounds. Main contamination sources are available in all farming systems and must be taken seriously. Mycotoxins holds many problems among the natural toxins. Naturally occurring toxicant contamination of feeds and foods with mycotoxins is inevitable and unpredictable and poses a unique challenge to organic farming as well as conventional farming. The best way to reduce the mycotoxin content in food and feed is the prevention of mycotoxin formation in the field, but this is often not sufficient, consequently other effective methods are required. A well maintained quality assurance system has to be set up based on the occurrence, detection and prevention. Good agricultural, handling, manufacturing and storage practices are required in both organic and conventional agriculture to minimize the risk of natural toxins. Residues transferred to edible animal products presents consumer health risks. Food safety is the shared responsibility of governments, academia, the food and feed

industry farmers and the consumer. In view of consumer expectations, it is important that governments, industry and consumer groups carefully examine issues related to organic food quality and safety and make whatever interventions may be necessary to ensure an appropriate level of consumer protection.

REFERENCES

- Benbrook C, McCullum-Gomez C:** Organic vs conventional farming. *J Am Diet Assoc*, 109, 809-811, 2009.
- Holdrege C:** Blame factory farming, not organic food. *Nat Biotechnol*, 25, 165, 2007.
- Toma L, Mathijs E:** Environmental risk perception, environmental concern and propensity to participate in organic farming programmes. *J Environ Manage*, 83, 145-157, 2007.
- Bavec M, Turinek M, Grobelnik-Mlakar S, Slatnar A, Bavec F:** Influence of industrial and alternative farming systems on contents of sugars, organic acids, total phenolic content, and the antioxidant activity of red beet (*Beta vulgaris* L. ssp. *vulgaris* Rote Kugel). *J Agr Food Chem*, 58, 11825-11831, 2010.
- Suarez-Serrano A, Ibanez C, Lacorte S, Barata C:** Ecotoxicological effects of rice field waters on selected planktonic species: Comparison between conventional and organic farming. *Ecotoxicology*, 19, 1523-1535, 2010.
- Horrigan L, Lawrence RS, Walker P:** How sustainable agriculture can address the environmental and human health harms of industrial agriculture. *Environ Health Persp*, 110, 445-456, 2002.
- Lockeretz W, Shearer G, Kohl DH:** Organic farming in the corn belt. *Science*, 211, 540-547, 1981.
- Macilwain C:** Organic: is it the future of farming? *Nature*, 428, 792-793, 2004.
- Dawson JC, Serpolay E, Giuliano S, Schermann N, Galic N, Chable V, Goldringer I:** Multi-trait evolution of farmer varieties of bread wheat after cultivation in contrasting organic farming systems in Europe. *Genetica*, 140, 1-17, 2012.
- Blakeway S:** Welfare and organic farming. *Vet Rec*, 134, 71, 1994.
- Sekkin S, Kum C:** Possible natural toxins in organic farming. *III. Symposium of Livestock Production with International Participation*, September 12-14, Ohrid, pp. 597-604, 2007.
- Greenough PR:** Animal welfare in dairy farming: Lameness and the organic movement. *Vet J*, 180, 3-4, 2009.
- EC (European Communities):** Organic Farming: Guide to Community Rules. EC, Belgium. 2012. Available on-line (November, 2012) at http://ec.europa.eu/agriculture/organic/eu-policy/legislation_en.
- Vaarst M, Roderick S, Lund V, Lockeretz W, Hovi M:** Organic principles and values: The framework for organic animal husbandry. In: Vaarst M, Roderick S, Lund V, Lockeretz W (Eds): *Animal Health and Welfare in Organic Agriculture*. pp. 1-12, Cabi Publishing, Cambridge, 2004.
- Velimirov A, Huber M, Lauridsen C, Rembialkowska E, Seidel K, Bugel S:** Feeding trials in organic food quality and health research. *J Sci Food Agr*, 90, 175-182, 2010.
- Kahl J, Baars T, Bugel S, Busscher N, Huber M, Kusche D, Rembialkowska E, Schmid O, Seidel K, Taupier-Letage B, Velimirov A, Zalecka A:** Organic food quality: A framework for concept, definition and evaluation from the European perspective. *J Sci Food Agr*, 92, 2760-2765, 2012.
- Migliori C, Di Cesare LF, Lo Scalzo R, Campanelli G, Ferrari V:** Effects of organic farming and genotype on alimentary and nutraceutical parameters in tomato fruits. *J Sci Food Agr*, 92, 2833-2839, 2012.
- Oates L, Cohen M, Braun L:** Characteristics and consumption patterns of Australian organic consumers. *J Sci Food Agr*, 92, 2782-2787, 2012.
- Özen A, Doğan Ö, Gül RTB, Özkul T, Yüksel E:** Türkiye'de veteriner hekimliği üzerine araştırmalar: III. İş fırsatları ve sektörel yönelimlere ilişkin görüş ve beklentiler. *Kafkas Univ Vet Fak Derg*, 18, 907-911, 2012.
- Benoit M, Laignel G:** Constraints under organic farming on French sheep meat production: A legal and economic point of view with an emphasis on farming systems and veterinary aspects. *Vet Res*, 33, 613-624, 2002.
- Sekkin S:** Organik tarım ve gıda güvenliği. *I. Uluslararası Organik Hayvansal Üretim ve Gıda Güvenliği Kongresi*, 28 Nisan - 1 Mayıs, Kuşadası, s. 92-108, 2004.
- Siderer Y, Maquet A, Anklam E:** Need for research to support consumer confidence in the growing organic food market. *Trends Food Sci Tech*, 16, 332-343, 2005.
- Garmo RT, Waage S, Sviland S, Henriksen BI, Osteras O, Reksen O:** Reproductive performance, udder health, and antibiotic resistance in mastitis bacteria isolated from Norwegian Red cows in conventional and organic farming. *Acta Vet Scand*, 52, 11, 2010.
- Hammarberg KE:** Animal welfare in relation to standards in organic farming. *Acta Vet Scand*, Suppl 95, 17-25, 2001.
- Kouba M:** The product quality and health implications of organic products. In: Kyriazakis I, Zervas G (Eds): *Organic Meat and Milk from Ruminants*. EAAP (European Association for Animal Production) Publication no: 106, pp. 57-64, Wageningen Academic Publishers, Athens, 2002. Available on-line (November, 2012) at <http://www.eaap.org/docs/Publications/eaap106%20-%207249456W.pdf>.
- Trewavas A:** A critical assessment of organic farming and food assertions with particular respect to the UK and the potential environmental benefits of no-till agriculture. *Crop Prot*, 23, 757-781, 2004.
- Kijlstra A, Groot M, Roest J, Kasteel D, Eijk I:** Analysis of black holes in our knowledge concerning animal health in the organic food production chain. pp. 1-55, 2003. Available on-line (November, 2012) at <http://orgprints.org/00001034> or <http://orgprints.org/1034/1/IDhiaten.pdf>.
- Van-Loo EJ, Alali W, Ricke SC:** Food safety and organic meats. *Annu Rev Food Sci Technol*, 3, 203-225, 2012.
- Lund V, Algers B:** Research on animal health and welfare in organic farming - A literature review. *Livest Prod Sci*, 80, 55-68, 2003.
- Jensen KD, Larsen HN, Mølgaard JP, Andersen JO, Tingstad A, Marckmann P, Astrup A:** Organic foods and human health. Organic food and farming. *Towards Partnership and Action in Europe Proceedings*, 10-11 May 2001, pp. 171-177, Nørhaven A/S, Denmark (EMAS), Copenhagen, 2001.
- Benbrook CM:** Breaking the mold-impacts of organic and conventional farming systems on mycotoxins in food and livestock feed. An Organic Center State of Science Review. 1-58, 2005. Available on-line (November, 2012) at http://www.organic-center.org/reportfiles/Mycotoxin_SSR.pdf.
- Sekkin S:** Potential toxic agents of animals associated with feeding in organic farming. *10th Congress of the European Society of Veterinary and Comparative Nutrition*, October 5 - 7, National Veterinary School of Nantes, p. 183, 2006.
- Jansen JP, Defrance T, Warnier AM:** Effects of organic-farming-compatible insecticides on four aphid natural enemy species. *Pest Manag Sci*, 66, 650-656, 2010.
- Sirri F, Castellini C, Bianchi M, Petracci M, Meluzzi A, Franchini A:** Effect of fast-, medium- and slow-growing strains on meat quality of chickens reared under the organic farming method. *Animal*, 5, 312-319, 2011.
- FAO (Food and Agriculture Organization):** Food safety and quality as affected by organic farming. ERC/00/7, Agenda Item 10.1. *22nd FAO Regional Conference for Europe*. Porto, 24-28 July 2000. Available on-line (November, 2012) at <http://www.fao.org/docrep/meeting/X4983e.htm>.
- Browne AW, Harris PJC, Hofny-Collins AH, Pasiecznik N, Wallace RR:** Organic production and ethical trade: Definition, practice and links. *Food Policy*, 25, 69-89, 2000.
- McCann E, Sullivan S, Erickson D, De-Young R:** Environmental Awareness, Economic Orientation, and Farming Practices: A Comparison of Organic and Conventional Farmers. *Environ Manage*, 21, 747-758, 1997.
- Thamsborg SM:** Organic farming in the Nordic countries-animal

health and production. *Acta Vet Scand*, Suppl 95, 7-15, 2001.

39. Pandey J, Pandey U: Accumulation of heavy metals in dietary vegetables and cultivated soil horizon in organic farming system in relation to atmospheric deposition in a seasonally dry tropical region of India. *Environ Monit Assess*, 148, 61-74, 2009.

40. MacRae G: Rice farming in Bali: Organic production and marketing challenges. *Crit Asian Stud*, 43, 69-92, 2011.

41. Akbaba U, Sahin Y, Turkez H: Element content analysis by WDXRF in pistachios grown under organic and conventional farming regimes for human nutrition and health. *Toxicol Ind Health*, 28, 783-788, 2012.

42. Mader P, Fliessbach A, Dubois D, Gunst L, Fried P, Niggli U: Soil fertility and biodiversity in organic farming. *Science*, 296, 1694-1697, 2002.

43. Tuomisto HL, Hodge ID, Riordan P, MacDonald DW: Does organic farming reduce environmental impacts? A meta-analysis of European research. *J Environ Manage*, 112, 309-320, 2012.

44. Kirchmann H, Thorvaldson G: Challenging targets for future agriculture. *Eur J Agron*, 12, 145-161, 2000.

45. Yu W, Zhang L, Shen S, Lian H: A long-term field trial on fertilization and on use of recycled nutrients in farming systems III. Recycling rate of P and N through a composting cycle and recoveries of P and N in organic manure. *J Appl Ecol*, 13, 1407-1409, 2002.

46. Weller RF, Cooper A: Health status of dairy herds converting from conventional to organic dairy farming. *Vet Rec*, 139, 141-142, 1996.

47. Dalgaard T, Kjeldsen C, Hutchings NJ and Hansen JF: N-losses and energy use in a scenario for conversion to organic farming. *Sci World J Suppl* 2, 822-829, 2001.

48. Prandini A, Sigolo S, Piva G: Conjugated linoleic acid (CLA) and fatty acid composition of milk, curd and Grana Padano cheese in conventional and organic farming systems. *J Dairy Res*, 76, 278-282, 2009.

49. Schneewis I, Meyer K, Ritzmann M, Hoffmann P, Dempfle L, Bauer J: Influence of organically or conventionally produced wheat on health, performance and mycotoxin residues in tissues and bile of growing pigs. *Arch Anim Nutr*, 59, 155-163, 2005.

50. DEFRA (Department for Environment, Food and Rural Affairs): Compendium of UK organic standards, DEFRA, London. pp. 1-104, 2006. Available on-line (November, 2012). at <http://archive.defra.gov.uk/foodfarm/growing/organic/standards/pdf/compendium.pdf>.

51. SA (Soil Association): Soil Association organic standards farming and growing. Revision-16, pp. 1-241, 2012. Available on-line (November, 2012) at <http://www.soilassociation.org/LinkClick.aspx?fileticket=l-qUg6illo%3D&tabid=353>.

52. Reidsma P, Tekelenburg T, Berg M, Alkemade R: Impacts of land-use change on biodiversity: An assessment of agricultural biodiversity in the European Union. *Agr Ecosyst Environ*, 114, 86-102, 2006.

53. Rio M, Franco-Uria A, Abad E, Roca E: A risk-based decision tool for the management of organic waste in agriculture and farming activities (FARMERS). *J Hazard Mater*, 185, 792-800, 2011.

54. FAO (Food and Agriculture Organization): Animal feeding and food safety, FAO Food and nutrition paper-69. pp. 1-47, Report of an FAO Expert Consultation, Rome, Italy, 10-14 March 1997. FAO Food and Nutrition Division. 1998. Available on-line (November, 2012) at <ftp://ftp.fao.org/esn/food/W8901E.pdf>.

55. Baars T: Review of animal health and welfare. In, Isart J, Llerena JJ (Eds): Organic Farming Research in the EU, Towards 21st Century, ENOF White Book, pp. 65-74, CAB Direct, Barcelona, 1999.

56. Baars T: Review of Grassland and Fodder Production. In, Isart J, Llerena JJ (Eds): Organic Farming Research in the EU, Towards 21st Century, ENOF White Book, pp. 77-83, CAB Direct, Barcelona, 1999.

57. AFSSA (Agence Française de Sécurité Sanitaire des Aliments): Evaluation nutritionnelle et sanitaire des aliments issus de l'agriculture biologique. Chapitre 4: Aspect Sanitaires. pp. 61-113, 2003. Available on-line (November, 2012) at <http://www.anses.fr/Documents/NUT-Ra-AgriBio.pdf>.

58. Saba A, Messina F: Attitudes towards organic foods and risk/benefit

perception associated with pesticides. *Food Qual Prefer*, 14, 637-645, 2003.

59. D'Mello JPF: Contaminants and toxins in animal feeds. 2006. [http://www.fao.org/waicent/faoinfo/agricult/aga/agap/frg/Feed safety/ffsp6.htm](http://www.fao.org/waicent/faoinfo/agricult/aga/agap/frg/Feed%20safety/ffsp6.htm), Available on-line: November, 2012.

60. Vaarst M, Bennedsgaard TW: Reduced medication in organic farming with emphasis on organic dairy production. *Acta Vet Scand*, Suppl 95, 51-57, 2001.

61. Malmauret L, Massin DP, Hardy JL, Verger P: Contaminants in organic and conventional foodstuffs in France. *Food Addit Contam*, 19, 524-532, 2002.

62. Lo M, Matthews D: Results of routine testing of organic food for agro-chemical residues. In, Powell J (Ed): UK Organic Research 2002. *Proceedings of the COR Conference*, 26-28th March, Aberystwyth, 61-64, 2002. Available on-line (November, 2012) at http://orgprints.org/8268/1/Lo_matthews_testing_agrochemical_residues.pdf

63. Keck G: Contaminants et résidus chimiques dans les aliments d'origine animale. *Rev Fr Lab*, 348, 21-27, 2002.

64. Jensen AN, Dalsgaard A, Stockmarr A, Nielsen EM, Baggesen DL: Survival and transmission of *Salmonella enterica* serovar typhimurium in an outdoor organic pig farming environment. *Appl Environ Microb*, 72, 1833-1842, 2006.

65. Ammann K: Why farming with high tech methods should integrate elements of organic agriculture. *New Biotechnol*, 25, 378-388, 2009.

66. Schmid F, Moser G, Muller H, Berg G: Functional and structural microbial diversity in organic and conventional viticulture: Organic farming benefits natural biocontrol agents. *Appl Environ Microb*, 77, 2188-2191, 2011.

67. Fagnano M, Fiorentino N, D'Egidio MG, Quaranta F, Ritieni A, Ferracane R, Raimondi G: Durum wheat in conventional and organic farming: yield amount and pasta quality in Southern Italy. *Sci World J*, 2012 Article ID 973058, 1-9, 2012.

68. Duarte SC, Lino CM, Pena A: Ochratoxin A in feed of food-producing animals: an undesirable mycotoxin with health and performance effects. *Vet Microbiol*, 154, 1-13, 2011.

69. Milne JA: Societal expectations of livestock farming in relation to environmental effects in Europe. *Livest Prod Sci*, 96, 3-9, 2005.

70. Oleskowicz-Popiel P, Kadar Z, Heiske S, Klein-Marcuschamer D, Simmons BA, Blanch HW, Schmidt JE: Co-production of ethanol, biogas, protein fodder and natural fertilizer in organic farming-evaluation of a concept for a farm-scale biorefinery. *Bioresour Technol*, 104, 440-446, 2012.

71. Hovi M, Sundrum A, Thamsborg SM: Animal health and welfare in organic livestock production in Europe: Current state and future challenges. *Livest Prod Sci*, 80, 41-53, 2003.

72. Gibon A: Managing grassland for production, the environment and the landscape. Challenges at the farm and the landscape level. *Livest Prod Sci*, 96, 11-31, 2005.

73. Esperschütz J, Gattinger A, Mader P, Schlöter M, Fliessbach A: Response of soil microbial biomass and community structures to conventional and organic farming systems under identical crop rotations. *FEMS Microbiol Ecol*, 61, 26-37, 2007.

74. Rashad FM, Saleh WD, Moselhy MA: Bioconversion of rice straw and certain agro-industrial wastes to amendments for organic farming systems: 1. Composting, quality, stability and maturity indices. *Bioresour Technol*, 101, 5952-5960, 2010.

75. Suttmoller P: Contaminated food of animal origin: hazards and risk management. Contamination of Animal Products: Prevention and Risks for Public Health, *OIE Sci Tech Rev*, 16, 1-26, 1997. Available on-line (November, 2012) at <http://siteresources.worldbank.org/INTARD/843432-1111149860300/20434404/ContaminatedFood.pdf>.

76. Sanders TAB: Food production and food safety. *Brit Med J*, 318, 1689-1693, 1999.

77. Bourn D, Prescott J: A comparison of the nutritional value, sensory qualities, and food safety of organically and conventionally produced

foods. *Crit Rev Food Sci Nutr*, 42, 1-34, 2002.

78. Walk ST, Mladonicky JM, Middleton JA, Heidt AJ, Cunningham JR, Bartlett P, Sato K, Whittam TS: Influence of antibiotic selection on genetic composition of *Escherichia coli* populations from conventional and organic dairy farms. *Appl Environ Microb*, 73, 5982-5989, 2007.

79. Kersting AL, Medeiros LC, LeJeune JT: Differences in *Listeria monocytogenes* contamination of rural Ohio residences with and without livestock. *Foodborne Pathog Dis*, 7, 57-62, 2010.

80. Schwaiger K, Schmied EM, Bauer J: Comparative analysis on antibiotic resistance characteristics of *Listeria* spp. and *Enterococcus* spp. isolated from laying hens and eggs in conventional and organic keeping systems in Bavaria, Germany. *Zoonoses Public Health*, 57, 171-180, 2010.

81. Luo X, Cai X: A combined use of autolysin p60 and Listeriolysin-O antigens induces high protective immune responses against *Listeria monocytogenes* infection. *Curr Microbiol*, 65, 813-818, 2012.

82. Kryszka-Traczyk E: Microflora of the farming work environment as an occupational risk factor. *Med Pr*, 51, 351-355, 2000.

83. Rintala H, Pitkaranta M, Taubel M: Microbial communities associated with house dust. *Adv Appl Microbiol*, 78, 75-120, 2012.

84. Boutrif E: Risk of undesired substances in feeds and animal food products. Food safety management in developing countries. *Proceedings of the International Workshop, CIRAD-FAO*, 11-13 December, Montpellier, pp. 1-6, 2002. Available on-line (November, 2012) at <http://www.abef.com.br/download/workshop/trabalhocientifico/10>.

85. Kaya S: Mikotoksinler. In: Kaya S, Pirinççi İ, Bilgili A (Eds): Veteriner Hekimliğinde Toksikoloji. Baskı 2. s. 537-573, Medisan Yayınevi, Ankara, 2002.

86. Pattono D, Gallo PF, Civera T: Detection and quantification of Ochratoxin A in milk produced in organic farms. *Food Chem*, 127, 374-377, 2011.

87. Dohman E: Mycotoxin hazards and regulations. Impacts on food and animal feed crop trade. Economic Research Service/USDA, International Trade and Food Safety, AER-828, pp. 97-108, 2006. Available on-line (November, 2012) at <http://www.ers.usda.gov/publications/aer828/aer828h.pdf>.

88. Suproniene S, Justesen A, Nicolaisen M, Mankeviciene A, Dabkevicius Z, Semaskiene R, Leistrumaitė A: Distribution of trichothecene and zearalenone producing *Fusarium* species in grain of different cereal species and cultivars grown under organic farming conditions in Lithuania. *Ann Agr Env Med*, 17, 79-86, 2010.

89. Bernhoft A, Torp M, Clasen PE, Loes AK, Kristoffersen AB: Influence of agronomic and climatic factors on *Fusarium* infestation and mycotoxin contamination of cereals in Norway. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess*, 29, 1129-1140, 2012.

90. Akar F, Sarı M, Akbaş L, Sekkin S, Kum C: Aydın ili ve çevresinden sağlanan mısır örneklerinin Fumonisin B₁ ile kirlenme durumu üzerinde çalışmalar. *Bornova Vet Kont Araşt Enst Derg*, 24, 15-19, 1999.

91. Arino A, Juan T, Estopanan G, Gonzalez-Cabo JF: Natural occurrence of *Fusarium* species, fumonisin production by toxigenic strains, and concentrations of Fumonisin B₁ and B₂ in conventional and organic maize grown in Spain. *J Food Protect*, 70, 151-156, 2007.

92. Pozzo L, Cavallarin L, Nucera D, Antoniazzi S, Schiavone A: A survey of ochratoxin A contamination in feeds and sera from organic and standard swine farms in northwest Italy. *J Sci Food Agr*, 90, 1467-1472, 2010.

93. Demet Ö, Oğuz H, Çelik İ, Adıgüzel H: Buğday, mısır, pirinç ve yerfıstığında aflatoksin üretilmesi. *Vet Bil Derg*, 11:135-140, 1995.

94. Oğuz H, Kurtoglu V: Effect of clinoptilolite on performance of broiler chickens during experimental aflatoxicosis. *Br Poult Sci*, 41, 512-517, 2000.

95. Oğuz H: A review from experimental trials on detoxification of aflatoxin in poultry feed. *Eurasian J Vet Sci*, 27, 1-12, 2011.

96. D'Mello JPF, MacDonald AMC: Mycotoxins. *Anim Feed Sci Tech*, 69, 155-166, 1997.

97. Khoshpey B, Farhud D, Zaini F: Aflatoxins in Iran: Nature, hazards and carcinogenicity. *Iran J Public Health*, 40, 1-30, 2011.

98. Wyss GS: Assessing the risk from mycotoxins for the organic food

chain: results from Organic HACCP-project and other research. Systems development: quality and safety of organic livestock products. *Proceedings of the 4th SAFO Workshop*, Frick, Switzerland, pp. 133-136, 2005. <http://orgprints.org/5921/01/Wyss-2005-SAFO-mycotoxins.pdf>, Available on-line: November, 2012.

99. Heaton S: Organic farming, food quality and human health: A review of the evidence. Soil Association Bristol House, Registered charity no: 206862. pp. 1-88, 2001. http://www.soilassociation.org/LinkClick.aspx?fileticket=cY8kfP3Q%2BgA%3D&tnqh_x0026;tabid=388, Available on-line: November, 2012.

100. Bordeleau G, Myers-Smith I, Midak M, Szeremeta A: Food Quality: A comparison of organic and conventional fruits and vegetables. Ecological Agriculture Den Kongelige Veterinær-og Landbohøjskole. 1-81, 2002. <http://www.courseinfo.life.ku.dk/KurserLPLF10355/presentation/~media/Kurser/UJV/250069/foodqualityfinal.pdf.ashx>, Available on-line: November, 2012.

101. Durmic Z, Blache D: Bioactive plants and plant products: Effects on animal function, health and welfare. *Anim Feed Sci Tech*, 176, 150-162, 2012.

102. Villalba JJ, Provenza FD, Hall JO, Lisonbee LD: Selection of tannins by sheep in response to gastrointestinal nematode infection. *J Anim Sci*, 88, 2189-2198, 2010.

103. Villalba JJ, Provenza FD, Manteca X: Links between ruminants' food preference and their welfare. *Animal*, 4, 1240-1247, 2010.

104. Norman HC, Masters DG, Barrett-Lennard EG: Halophytes as forages in saline landscapes: Interactions between plant genotype and environment change their feeding value to ruminants. *Environ Exp Bot*, (in press), 2012.

105. Codex Alimentarius: Prevention and Reduction of Food and Feed Contamination. pp. 1-178, WHO-FAO Rome, 2012.

106. Weltmeier F, Maser A, Menze A, Hennig S, Schad M, Breuer F, Schulz B, Holtschulte B, Nehls R, Stahl DJ: Transcript profiles in sugar beet genotypes uncover timing and strength of defense reactions to *Cercospora beticola* infection. *Mol Plant Microbe Interact*, 24, 758-772, 2011.

107. Pechanova O, Pechan T, Williams WP, Luthe DS: Proteomic analysis of the maize rachis: potential roles of constitutive and induced proteins in resistance to *Aspergillus flavus* infection and aflatoxin accumulation. *Proteomics*, 11, 114-127, 2011.

108. Chen ZY, Brown RL, Damann KE, Cleveland TE: PR10 expression in maize and its effect on host resistance against *Aspergillus flavus* infection and aflatoxin production. *Mol Plant Pathol*, 11, 69-81, 2010.

109. Jard G, Liboz T, Mathieu F, Guyonvarc'h A, Lebrihi A: Review of mycotoxin reduction in food and feed: from prevention in the field to detoxification by adsorption or transformation. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess*, 28, 1590-1609, 2011.

110. Magan N, Aldred D, Mylona K, Lambert RJ: Limiting mycotoxins in stored wheat. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess* 27, 644-650, 2010.

111. Magan N, Aldred D: Post-harvest control strategies: minimizing mycotoxins in the food chain. *Int J Food Microbiol*, 119, 131-139, 2007.

112. Kabak B, Dobson AD, Var I: Strategies to prevent mycotoxin contamination of food and animal feed: A review. *Crit Rev Food Sci Nutr*, 46, 593-619, 2006.

113. Aldred D, Magan N: Prevention strategies for trichothecenes. *Toxicol Lett*, 153, 165-171, 2004.

114. Edlayne G, Simone A, Felicio JD: Chemical and biological approaches for mycotoxin control: A review. *Recent Pat Food Nutr Agric*, 1, 155-161, 2009.

115. Kabak B, Dobson AD: Biological strategies to counteract the effects of mycotoxins. *J Food Prot*, 72, 2006-2016, 2009.

116. Castoria R, Mannina L, Duran-Patron R, Maffei F, Sobolev AP, De Felice DV, Pinedo-Rivilla C, Ritieni A, Ferracane R, Wright SA: Conversion of the mycotoxin patulin to the less toxic desoxypatulinic acid by the biocontrol yeast *Rhodosporidium kratochvilovae* strain LS11. *J Agric Food Chem*, 59, 11571-11578, 2011.

-
- 117. Juglal S, Govinden R, Odhav B:** Spice oils for the control of co-occurring mycotoxin-producing fungi. *J Food Prot*, 65, 683-687, 2002.
- 118. Souza MF, Tome AR, Rao VS:** Inhibition by the bioflavonoid ternatin of aflatoxin B1-induced lipid peroxidation in rat liver. *J Pharm Pharmacol*, 51, 125-129, 1999.
- 119. Srinivasan S, Sarada DV:** Antifungal activity of phenyl derivative of pyranocoumarin from *Psoralea corylifolia* L. seeds by inhibition of acetylation activity of trichothecene 3-o-acetyltransferase (Tri101). *J Biomed Biotechnol*, doi: 10.1155/2012/310850, 2012.
- 120. Awad WA, Ghareeb K, Bohm J, Zentek J:** Decontamination and detoxification strategies for the *Fusarium* mycotoxin deoxynivalenol in animal feed and the effectiveness of microbial biodegradation. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess*, 27, 510-520, 2010.
- 121. He J, Zhou T:** Patented techniques for detoxification of mycotoxins in feeds and food matrices. *Recent Pat Food Nutr Agric*, 2, 96-104, 2010.
- 122. Jansen van Rensburg C, Van Rensburg CE, Van Ryssen JB, Casey NH, Rottinghaus GE:** *In vitro* and *in vivo* assessment of humic acid as an aflatoxin binder in broiler chickens. *Poult Sci*, 85, 1576-1583, 2006.

YAZIM KURALLARI

1- Yılda 6 (Altı) sayı olarak yayımlanan Kafkas Üniversitesi Veteriner Fakültesi Dergisi'nde (Kısaltılmış adı: Kafkas Univ Vet Fak Derg) Veteriner Hekimlik ve Hayvancılıkla ilgili (klinik ve paraklinik bilimler, hayvancılıkla ilgili biyolojik ve temel bilimler, zoonozlar ve halk sağlığı, hayvan besleme ve beslenme hastalıkları, hayvan yetiştiriciliği ve genetik, hayvansal orijinli gıda hijyeni ve teknolojisi, egzotik hayvan bilimi) orijinal araştırma, kısa bildiri, ön rapor, gözlem, editöre mektup, derleme ve çeviri türünde yazılar yayımlanır. Dergide yayımlanmak üzere gönderilen makaleler Türkçe, İngilizce veya Almanca dillerinden biri ile yazılmış olmalıdır.

2- Dergide yayımlanması istenen yazılar Times New Roman yazı tipi ve 12 punto ile A4 formatında, 1.5 satır aralıklı ve sayfa kenar boşlukları 2.5 cm olacak şekilde hazırlanmalı ve resim, tablo, grafik gibi şekillerin metin içindeki yerlerine Türkçe ve yabancı dilde adları ve gerekli açıklamaları mutlaka yazılmalıdır.

Dergiye gönderilecek makale ve ekleri (şekil vs) <http://vetdergi.kafkas.edu.tr> adresindeki online makale gönderme sistemi kullanılarak yapılmalıdır.

Başvuru sırasında yazarlar yazıda yer alacak şekilleri (13 X 18 cm boyutlarından büyük olmamalı) online makale gönderme sistemine yüklemelidirler. Yazının kabul edilmesi durumunda tüm yazarlarca imzalanmış *Telif Hakkı Devir Sözleşmesi* editörlüğe gönderilmelidir.

3- Yazarlar yayımlamak istedikleri makale ile ilgili olarak gerekli olan etik kurulu onayı aldıkları kurumu ve onay numarasını Materyal ve Metot bölümünde belirtmelidirler. Yayın kurulu gerekli gördüğünde etik kurul onay belgesini ayrıca isteyebilir.

4- Makale Türleri

Orijinal Araştırma Makaleleri, yeterli bilimsel inceleme, gözlem ve deneylere dayanarak bir sonuca ulaşan orijinal ve özgün çalışmalardır.

Türkçe yazılmış makaleler Türkçe başlık, Türkçe özet ve anahtar sözcükler, yabancı dilde başlık, yabancı dilde özet ve anahtar sözcükler, giriş, materyal ve metot, bulgular, tartışma ve sonuç ile kaynaklar bölümlerinden oluşmalı ve metin, tablo, şekil vs dahil) 10 sayfayı aşmamalıdır. Yabancı dilde yazılmış makaleler yabancı dilde başlık, yabancı dilde özet ve anahtar sözcükler, Türkçe başlık, Türkçe özet ve anahtar sözcükler ile birlikte Türkçe makale yazım kurallarında belirtilen diğer bölümlerden oluşur. Türkçe ve yabancı dilde özetlerin her biri yaklaşık 200±20 sözcükten oluşmalıdır.

Kısa Bildiri, konu ile ilgili yeni bilgi ve bulguların bildirildiği fakat orijinal araştırma olarak sunulamayacak kadar kısa olan yazılardır. Kısa bildiriler, orijinal araştırma makalesi formatında olmalı, fakat özetlerin her biri 100 sözcüğü aşmamalı, referans sayısı 15'in altında olmalı ve 4 sayfayı aşmamalıdır. Ayrıca, en fazla 4 şekil veya tablo içermelidir.

Ön Rapor, kısmen tamamlanmış, yorumlanabilecek aşamaya gelmiş orijinal bir araştırmanın kısa (en çok 6 sayfa) anlatımıdır. Bunlar orijinal araştırma makalesi formatında yazılmalıdır.

Gözlem, uygulama, klinik veya laboratuvar alanlarında ender olarak rastlanılan olguların sunulduğu makalelerdir. Bu yazıların başlık ve özetleri orijinal makale formatında yazılmalı, bundan sonraki bölümleri giriş, olgunun tanımı, tartışma ve sonuç ile kaynaklardan oluşmalı ve 4 sayfayı geçmemelidir.

Editöre Mektup, bilimsel veya pratik yararı olan bir konunun veya ilginç bir olgunun resimli ve kısa sunumudur ve 1 sayfayı geçmemelidir.

Derleme, güncel ve önemli bir konuyu, yazarın kendi görüş ve araştırmalarından elde ettiği bulguların da değerlendirildiği özgün yazılardır. Bu yazıların başlık ve özet bölümleri orijinal araştırma makalesi formatında yazılmalı, bundan sonraki bölümleri giriş, metin ve kaynaklardan oluşmalı ve 10 sayfayı geçmemelidir.

Çeviri, makalenin orijinal formatı dikkate alınarak hazırlanmalıdır.

Yazarla ilgili kişisel ve kuruma ait bilgiler ana metin dosyasına değil, on-line başvuru sırasında sistemdeki ilgili yerlere unvan belirtilmeksizin eklenmelidir.

5- Makale ile ilgili gerek görülen açıklayıcı bilgiler (tez, proje, destekleyen kuruluş vs) makale başlığının sonuna üst simge olarak işaret konularak makale başlığı altında italik yazıyla belirtilmelidir.

6- Kaynaklar, metin içinde ilk verileden başlanarak numara almalı ve metin içindeki kaynağın atfı yapıldığı yerde parantez içinde yazılmalıdır.

Kaynak dergi ise, yazarların soyadları ve ilk adlarının başharfleri, makale adı, dergi adı (orijinal kısa ad), cilt ve sayı numarası, sayfa numarası ve yıl sıralamasına göre olmalı ve aşağıdaki örnekte belirtilen karakterler dikkate alınarak yazılmalıdır.

Örnek: Gokce E, Erdogan HM: An epidemiological study on neonatal lamb health. *Kafkas Univ Vet Fak Derg*, 15 (2): 225-236, 2009.

Kaynak kitap ise yazarların soyadları ile adlarının ilk harfleri, eserin adı, baskı sayısı, sayfa numarası, basımevi, basım yeri ve basım yılı olarak yazılmalıdır.

Editörlü ve çok yazarlı olarak yayınlanan kitaptan bir bölüm kaynak olarak kullanılmışsa, bölüm yazarları, bölüm adı, editör(ler), kitap adı, baskı sayısı, sayfa numarası, basımevi, basım yeri ve basım yılı sırası dikkate alınarak aşağıdaki örneğe göre yazılmalıdır.

Örnek: McIlwraith CW: Disease of joints, tendons, ligaments, and related structures. **In**, Stashak TS (Ed): *Adam's Lameness in Horses*. 4th ed. 339-447, Lea and Febiger, Philadelphia, 1988.

Online olarak ulaşılan kaynaklarda web adresi ve erişim tarihi kaynak bilgilerinin sonuna eklenmelidir.

Diğer kaynakların yazımında bilimsel yayın ilkelerine uyulmalıdır.

Kaynak listesinde "et al." ve "ve ark." gibi kısaltmalar yapılmaz.

7- Bakteri, virus, parazit ve mantar tür isimleri ve anatomik terimler gibi latince ifadeler orijinal şekliyle ve italik karakterle yazılmalıdır.

8- Editörlük, dergiye gönderilen yazılar üzerinde gerekli görülen kısaltma ve düzeltmeleri yapabileceği gibi önerilerini yazarlara iletebilir. Yazarlar, düzeltilmek üzere yollanan yazıları online sistemde belirtilen sürede gerekli düzeltmeleri yaparak editörlüğe iade etmelidirler. Editörlükçe ön incelemesi yapılan ve değerlendirmeye alınması uygun görülen makaleler ilgili bilim dalından bir yayın danışmanı ve iki raportörün olumlu görüşü alındığı takdirde yayımlanır.

9- Yayımlanan yazılardan dolayı doğabilecek her türlü sorumluluk yazarlara aittir.

10- Yazarlara telif ücreti ödenmez.

11- Resim ve baskı masrafları için yazarlardan ücret alınır. Ücret bilgileri <http://vetdergi.kafkas.edu.tr/> adresinden öğrenilebilir.

12- Yazarlara 50 adet ayrı baskı ücretsiz olarak yollanır.

INSTRUCTIONS FOR AUTHORS

1- Kafkas Üniversitesi Veteriner Fakültesi Dergisi (Journal of the Faculty of Veterinary Medicine, Kafkas University) (abbreviated title: Kafkas Univ Vet Fak Derg), published bi-monthly, covers original papers, short communication and preliminary scientific reports, case reports, observation, letter to the editor, review and translation on all aspects of veterinary medicine and animal science (clinical and paraclinical sciences, biological and basic sciences, zoonoses and public health, animal feeding and nutritional diseases, animal breeding and genetics, hygiene and technology of food from animal sources, exotic animal science). Manuscripts submitted for publication should be written in Turkish, English or German.

2- The manuscripts submitted for publication should be prepared in the format of Times New Roman style, font size 12, A4 paper size, 1.5 line spacing and 2.5 cm margins of all edges. The legend or caption of all illustrations such as figure, table and graphic must clearly be written in both Turkish and foreign language and their appropriate position should be indicated in the text.

The manuscript and its supplementary (figure etc.) should be submitted by using online manuscript submission system at the address of <http://vetdergi.kafkas.edu.tr/>

During the submission, the authors should upload the figures of the manuscript (the dimensions must not to exceed 13 X 18 cm) to the online manuscript submission system. If the manuscript is accepted for publication, the copyright transfer agreement form signed by all the authors should be send to the editorial office.

3- Authors should indicate the name of institute approves the necessary ethical commission report and the serial number of the approval in the material and methods section. If necessary, editorial board may also request the official document of the ethical commission report.

4- Original (full-length) manuscripts are original and proper scientific papers based on sufficient scientific investigations, observations and experiments.

Manuscripts written in Turkish consist of the title in Turkish, summary and keywords in Turkish, introduction, material and methods, results, discussion and references and it should not exceed 10 pages including text, tables and illustrations. Manuscripts written in a foreign language should follow the title in foreign language, summary and keywords in foreign language, title in Turkish, summary and keywords in Turkish and the remaining sections described above for the manuscripts written in Turkish.

Summaries written in Turkish or foreign language should contain 200±20 words.

Short communication manuscripts contain recent information and findings in the related topics; however, they are written with insufficient length to be a full-length original article. They should be prepared in the format of full-length original article but each of the summaries should not exceed 100 words, the reference numbers should not exceed 15 and the length of the text should be no longer than 4 pages. Additionally, they should not contain more than 4 figures or tables.

Preliminary scientific reports are short description (maximum 6 pages) of partially completed original research findings at interpretable level. These should be prepared in the format of full-length original articles.

Case reports describe rare significant findings encountered in the application, clinic and laboratory of related fields. The title and summary of these articles should be written in the format of full-length original articles and the remaining sections should follow introduction, case history, discussion and references without exceeding the total of 4 pages.

Letters to Editor are short and picture-documented presentations of subjects with scientific or practical benefits or interesting cases without exceeding 1 page.

Reviews are original manuscripts gather the literature on current and significant subject along with the commentary and findings of the author on the particular subject. The title and summary of this manuscript should be prepared as described for the full-length original articles and the remaining sections should follow introduction, text and references without exceeding 10 page.

Translations should be prepared based on the format of original document being translated.

The information about author/s and institution/s should be added during the online submission and the main document should be free of these information.

5- The necessary descriptive information (thesis, projects, financial supports etc) scripted as an italic font style should be explained below the manuscript title after placing a superscript mark at the end of title.

6- References should be listed with numerical order as they appear in the text and the reference number should be indicated inside the parentheses at the cited text place. References should have the order of surnames and initial letters of the authors, title of the article, title of the journal (original abbreviated title), volume and issue numbers, page numbers and the year of publication and the text formatting should be performed as shown in the example below.

Example: Gokce E, Erdogan HM: An epidemiological study on neonatal lamb health. Kafkas Univ Vet Fak Derg, 15 (2): 225-236, 2009.

If the reference is a book, it should follow surnames and initial letters of the authors, title of the book, edition number, page numbers, name and location of publisher and year of publication. If a chapter in book with an editor and several authors is used, names of chapter authors, name of chapter, editors, name of book, edition number, page numbers, name and location of publisher and year of publication and the formatting should be performed as shown in the example below.

Example: McIlwraith CW: Disease of joints, tendons, ligaments, and related structures. In, Stashak TS (Ed): Adam's Lameness in Horses. 4th ed. 339-447, Lea and Febiger, Philadelphia, 1988.

In the references can be reached online only, the web address and connection date should be added at the end of the reference information. The generally accepted scientific writing instructions must be complied with the other references.

Abbreviations, such as "et al" and "and friends" should not be used in the list of the references.

7- The Latin expression such as species names of bacterium, virus, parasite and fungus and anatomical terms must be written in italic character keeping their original forms.

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